Peculiar laboratory phenotype/genotype relationship due to compound inherited protein C defects in a child with severe venous thromboembolism

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Keywords
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Summary
A 7-years-old child who developed unprovoked deep vein thrombosis (DVT) and pulmonary embolism (PE) was tested for inherited thrombophilia. Protein C (PC) antigen level (87 %) and PC coagulometric and amidolytic activities (12 % and 11 %, respectively) were consistent with a homozygous PC type IIA phenotype.

The patient was carrier of two heterozygous missense mutations causing p.Arg32Cys substitution associated with a type I PC defect (“null allele”, from the paternal side) and p.Gly433Ser substitution responsible for a type IIA PC defect (from the maternal side).

Thus, the apparently normal PC antigen level in the proband was misleading in the interpretation of phenotype/genotype relationship of this compound PC defect. The child was also carrier of heterozygous prothrombin G20210A variant.

Severe venous thromboembolism can occur in otherwise healthy children with complex inherited thrombophilia. Careful laboratory characterization of the phenotype/genotype relationship can be crucial to correctly classify PC defects and for their management with anticoagulants or replacement therapy.

Introduction
Protein C (PC) is a natural anticoagulant whose deficiency is associated with an increased risk of venous thromboembolism (VTE). PC deficiency can be transmitted as an autosomal dominant or autosomal recessive trait. The prevalence of mild (heterozygous forms) PC deficiency in the healthy population ranges from 1/200 to 1/500 (1). Heterozygous carriers are often asymptomatic, but may experience recurrent thrombotic events in the presence of additional inherited prothrombotic risk factors, such as factor V (FV) Leiden or...
prothrombin (PT) G20210A mutations (2). In contrast, severe PC deficiency (homozygous and double heterozygous forms) is very rare (from 1/40000 to 1/250000) (3) and manifests in the neonatal period by purpura fulminans, disseminated intravascular coagulation and massive venous thrombosis (4) with rare exceptions.

Based on the functional and immunological assays, PC deficiency can be classified in three types. Type I deficiency, the more common form, is quantitative (“true” defect) and type II deficiency is qualitative (dysfunctional molecules). Type I deficiency is characterized by a concordant reduction in PC antigen and PC coagulometric and amidolytic activities. In contrast, type II deficiency exhibits normal PC antigen but the function of the molecule is impaired. Type II is further classified into two subtypes: type IIA, in which both coagulometric and amidolytic activities are reduced (when the lesion affects the catalytic site) (5), and type IIB, in which only the coagulometric activity is reduced (when the lesion affects the gamma-carboxyglutamatic acid-rich (Gla) domain or impairs the interaction of PC with its physiological substrates factors V and VIII) (6–8). Type III deficiency results from the combination in the heterozygous form of a type I defect with a type II defect, which leads to a reduced synthesis of a dysfunctional PC molecule. Type III deficiency shows a discrepancy between PC antigen and PC activities with a greater decrease in the level of both activities compared to the antigen level (9) (Tab. 1).

<table>
<thead>
<tr>
<th>Classification</th>
<th>PC antigen</th>
<th>PC amidolytic activity</th>
<th>PC coagulometric activity</th>
</tr>
</thead>
<tbody>
<tr>
<td>Type I (quantitative or true defects of PC)</td>
<td>Reduced</td>
<td>Reduced</td>
<td>Reduced</td>
</tr>
<tr>
<td>Type IIA (qualitative or dysfunctional defects of PC)</td>
<td>Normal</td>
<td>Reduced</td>
<td>Reduced</td>
</tr>
<tr>
<td>Type IIB (qualitative or dysfunctional defects of PC)</td>
<td>Normal</td>
<td>Normal</td>
<td>Reduced</td>
</tr>
<tr>
<td>Type III or Hypo-dys (quantitative and qualitative defects of PC)</td>
<td>Reduced (lower than the antigen level)</td>
<td>Reduced (lower than the antigen level)</td>
<td>Reduced (lower than the antigen level)</td>
</tr>
</tbody>
</table>

Here, we present a peculiar compound PC defect in combination with heterozygous PT G20210A mutation found in an Italian child with severe thrombotic manifestations.

**Case report and results**

A 7-year-old boy was admitted to our Pediatric Emergency Department in 2008 for a painful swelling of his leg appeared about 7 days after the onset of varicella.

Compression ultrasonography of the lower limbs revealed complete thrombosis of the right popliteal, superficial femoral, common femoral and iliac veins, with involvement of the right small and great saphenous veins, and almost complete thrombosis of the left small and great saphenous veins up to the cross with the left superficial femoral vein. Perfusion lung scan did not show any significant perfusion defect.

His family history was positive for thromboembolic events, since his father had suffered two episodes of deep vein thrombosis (DVT) in the right leg complicated by pulmonary embolism (PE).

Routine coagulation tests, performed in the patient on admission, yielded normal values except for the PC. PC antigen was 87% (normal range, 80–120%) whereas PC activity, measured by both coagulometric and amidolytic assays, were markedly reduced to 12% and 11%, respectively (normal range for both activities, 80–120%).

The child was also heterozygous for PT G20210A mutation. These preliminary findings suggested that the patient suffered from severe PC deficiency.

Therapeutic dose of subcutaneous low molecular weight heparin (LMWH) was started and after a few days the warfarin therapy was initiated. Compression ultrasonography was performed after a few days and, despite a partial recanalization of the veins of the right leg, an extension of thrombosis to the deep veins of the left leg was observed.

Because both PC coagulometric and amidolytic activities had fallen to undetectable levels, PC concentrates (Ceprotin®, Baxter, Vienna, Austria) were administered at the dose of 501U/Kg in association with intravenous heparin. Six days later, warfarin was added to the heparin treatment and, after an additional overlap of 1 week, international normalized ratio (INR) values were in therapeutic range (INR 2.0 to 3.0) and both heparin and PC concentrates were discontinued.

After 2 weeks of hospitalization, the child presented with sudden right-sided chest pain, fever, cough and difficulty breathing. Because of the clinical suspect of PE, lung perfusion scinti-scan was performed, which showed no perfusion of the right lung in the presence of a normal chest X-ray (mismatch). Warfarin was adjusted to achieve and maintain a therapeutic INR range of 2.5–3.5 and new infusions of PC concentrates (2000IU) were scheduled every 3 days. At this dosage, a trough level of PC activity was kept around 20% in this child. The patient’s symptoms improved in the subsequent 2 weeks and he was discharged.

No recurrences were observed in the following 6 months and PC concentrates were then discontinued without further VTE.

All available family members (Fig. 1) underwent coagulation screening. The levels of PC antigen and activities found in the proband (III.1) and his relatives are summarized in Tab. 2.

The proband’s father (II.1) (who was under warfarin therapy) and the proband’s grandfather (I.1) showed a concordant decrease in PC antigen and activities reflecting a heterozygous type I PC defect. The proband’s father was also found to be heterozygous for PT G20210A mutation.
levels half of normal and may develop thromboembolic events in early adulthood or remain asymptomatic throughout life (12). Homozygous or compound heterozygous individuals have very low or undetectable PC levels and may present with purpura fulminans and disseminated intravascular coagulation in the neonatal period (13, 14).

In this study, a severe PC defect was detected in a child with DVT in both lower limbs complicated by PE. In order to characterize the defect, family study was performed, which revealed two different PC gene lesions resulting in peculiar phenotypes. The proband’s father and other family members on this side showed a classical type I heterozygous PC deficiency due to p.Arg32Cys mutation (15–17). The proband’s mother and other family members on this side showed a type IIA heterozygous PC deficiency due to p.Gly433Ser mutation (16). Interestingly enough, the carriers of this mutation in our family presented with both PC coagulometric and amidolytic activities reduced to about 40–50% of normal, whereas antigen levels were higher than normal upper limit (134% and 139%, respectively). One explanation could be an overexpression of PC related to the mutation in itself or to genetic variations present in the PC gene promoter. DNA sequencing of the promoter region in carriers of p.Gly433Ser mutation revealed three common polymorphisms, known to influence the plasma antigen levels of PC (rs1799808, rs1799809 and rs1799810). However, these polymorphisms have been associated with low PC antigen levels (18, 19). Notably, different genome-wide association studies (GWAS) support the idea that polymorphisms in other genes may affect circulating levels of PC (20, 21). Another possible explanation of the increased antigen levels may be related to the mutation in itself or to genetic variations present in the PC gene promoter. DNA sequencing of the promoter region in carriers of p.Gly433Ser mutation revealed three common polymorphisms, known to influence the plasma antigen levels of PC (rs1799808, rs1799809 and rs1799810). However, these polymorphisms have been associated with low PC antigen levels (18, 19). Notably, different genome-wide association studies (GWAS) support the idea that polymorphisms in other genes may affect circulating levels of PC (20, 21). Another possible explanation of the increased antigen levels may be related to the mutation in itself or to genetic variations present in the PC gene promoter. DNA sequencing of the promoter region in carriers of p.Gly433Ser mutation revealed three common polymorphisms, known to influence the plasma antigen levels of PC (rs1799808, rs1799809 and rs1799810). However, these polymorphisms have been associated with low PC antigen levels (18, 19). Notably, different genome-wide association studies (GWAS) support the idea that polymorphisms in other genes may affect circulating levels of PC (20, 21). Another possible explanation of the increased antigen levels may be related to the type of antibodies used in the ELISA test (22) and their possible interaction with PC in plasma of carriers of p.Gly433Ser mutation. In other words, the mutation could result in an increased affinity for the antibodies causing an apparent higher PC antigen level in the ELISA test. Very interestingly, higher PC antigen levels were previously reported by Reitsma et al. in some probands affected by the same heterozygous mutation (16). Needless to say,
different anti-PC antibodies (monoclonal/polyclonal) in the ELISA assay could result in different levels of PC antigen.

What could it be the result of the combination of these two PC gene lesions in the same patient in terms of laboratory phenotype? Sequencing results in the proband showed the presence of one null allele (p.Arg32Cys mutation) and a second allele (p.Gly433Ser mutation) responsible for the synthesis of a dysfunctional PC molecule. Contrary to what expected, however, PC antigen was about 87% and PC activities around 10%. These findings are related to the effect of p.Gly433Ser mutation on laboratory tests for PC and no contribution to the effect of p.Gly433Ser mutation on laboratory phenotype suggestive of a severe inherited thrombophilia (24, 25) such as in our proband. Healthy children who are carriers of thrombophilia, however, may remain asymptomatic also in the presence of common inherited thrombophilic conditions (26).

Conclusion

The identification of the specific mutations underlying severe thrombotic manifestations, particularly in children, may help making an accurate diagnosis and providing patients with appropriate therapies. In many cases, both laboratory genotyping and phenotyping are required for a correct classification of PC defects together with an extensive investigation of family members, which includes both laboratory testing and collection of accurate clinical information.

References


Tab. 2  Protein C phenotype and genotype of the proband and family members

<table>
<thead>
<tr>
<th>Individual</th>
<th>PC antigen, %1</th>
<th>PC amidolytic activity, %1</th>
<th>PC coagulometric activity, %1</th>
<th>c.94C&gt;T p.Arg32Cys</th>
<th>c.1297G&gt;A p.Gly433Ser</th>
<th>Promoter haplotypes</th>
<th>Symptoms</th>
</tr>
</thead>
<tbody>
<tr>
<td>III.1, proband</td>
<td>87</td>
<td>11</td>
<td>12</td>
<td>Heterozygous</td>
<td>Heterozygous</td>
<td>CC,GG,TT</td>
<td>DVT + PE</td>
</tr>
<tr>
<td>II.1, father2</td>
<td>30</td>
<td>32</td>
<td>12</td>
<td>Heterozygous</td>
<td>Normal</td>
<td>CC,TA</td>
<td>DVT</td>
</tr>
<tr>
<td>II.2, mother</td>
<td>134</td>
<td>49</td>
<td>45</td>
<td>Normal Heterozygous</td>
<td>CT,GA</td>
<td>Asymptomatic</td>
<td></td>
</tr>
<tr>
<td>II.3, sister</td>
<td>98</td>
<td>94</td>
<td>82</td>
<td>Normal Heterozygous</td>
<td>CT,AA</td>
<td>Asymptomatic</td>
<td></td>
</tr>
<tr>
<td>I.1, paternal grandfather</td>
<td>49</td>
<td>52</td>
<td>46</td>
<td>Heterozygous</td>
<td>Normal</td>
<td>CT,GA</td>
<td>Asymptomatic</td>
</tr>
<tr>
<td>I.4, maternal grandmother</td>
<td>75</td>
<td>22</td>
<td>24</td>
<td>Normal</td>
<td>Heterozygous</td>
<td>CT,GA</td>
<td>Asymptomatic</td>
</tr>
<tr>
<td>II.3, maternal uncle</td>
<td>139</td>
<td>47</td>
<td>43</td>
<td>Normal</td>
<td>Heterozygous</td>
<td>CC,GG,TT</td>
<td>Asymptomatic</td>
</tr>
</tbody>
</table>

Nucleotide variations and amino acid changes were described according to the Human Genome Variation Society. 1Normal range for protein C antigen and activities, 80–120%. 2On oral anticoagulant therapy at the time of blood sampling. PC, protein C; DVT, deep vein thrombosis; PE, pulmonary embolism.