Cardio-Active Steroid Glycosides Against Heart Failure: Ups and Downs

Plants that contain cardio-active steroid glycosides (CGs) have been therapeutically used since ancient times. For instance, the usage of squill, Drimia maritima (L.) Stearn (Asparagaceae), was reported in the Ebers Papyrus [1]. At the beginning of the early modern history, the German botanist Leonhard Fuchs, who created the scientific name Digitalis for foxglove, described the plant in detail and recommended it for diuresis. However, due to the very difficult dosage, which is based on the narrow therapeutic index of CGs [2], digitalis therapy did not succeed at that time.

The successful usage of foxglove, Digitalis purpurea L. (Plantaginaceae; • Fig. 1), started in 1785 with a publication by the English physician William Withering. Due to his investigations, he could make clear recommendations on the preparation and dosage of the herbal remedy against dropsy (heart failure-associated edema) [1]. Unfortunately, the digitalis therapy became too successful: It was carelessly propagated as a universal remedy, which led to severe reservations among physicians and, consequently, to a shift away from this herbal remedy [3].

The next revival of digitalis started in the middle of the 19th century and was based on the experimental and clinical research by Ludwig Traube in Berlin and on the isolation of CGs by Oswald Schmiedeberg in Strasbourg [1]. With the beginning of the 20th century, digitalis therapy became the gold standard for the treatment of heart failure. A further research milestone was achieved in 1965 when the German pharmacologist Kurt Repke identified the cell membrane-located ion pump Na⁺-K⁺-ATPase as the molecular target of CGs [4]. Besides their action on this pump, they were also found to exert sympathoinhibitory effects and to increase the vagal tone [5].
The discovery of the binding of CGs to the Na⁺-K⁺-ATPase nourished speculations that endogenous CGs, so-called digitalis- or ouabain-like factors, might exist that serve as ligands of this target. In the late 1980s, hints to the existence of endogenous CGs intensified [6–8] and in the early 1990s, these compounds were eventually discovered [9–13]. Their biosynthesis, tissue of origin, and precise physiological as well as pathophysiological function is an ongoing matter of research and debate [14]. Interestingly, in critically ill patients (not treated with CGs) Berendes et al. [15] found that the occurrence of endogenous CGs in the blood was associated with an increased risk of mortality. The authors speculated that the secretion of endogenous CGs depends not only on the impaired myocardial function and the volume overload in these patients, but also on systemic inflammation. However, it is an open question whether the endogenous CGs are a mechanism of protection or part of the detrimental pathology.

Surprisingly, despite their intensive clinical use, randomized controlled trials with CGs have not been performed until the end of the 20th century. The results of these trials and of the respective meta-analyses were sobering: Both the efficacy and the safety of the digitalis-based therapy of heart failure were questioned –984. Since well-tolerated alternative drugs, such as angiotensin-converting enzyme inhibitors and β-blockers, that show clear benefits on the mortality of heart failure patients are available, the use of cardiac glycosides has declined increasingly.

Cardio-Active Steroid Glycosides as Antitumor Agents: A Well Evolved Field

CGs currently experience a fascinating renaissance in a completely different field: They are evaluated as cancer-preventive as well as anticancer drugs (Fig. 2). Their antitumor action was discovered 50 years ago: In 1967, Osamu Shiratori [23] from the Japanese company Shionogi reported about the antiproliferative action of several CGs isolated from Digitalis purpurea on neoplastic cells in vitro and in vivo. Moreover, an epidemiologic study published in 1979 indicated that women who were treated with CGs and who developed breast cancer showed a less aggressive tumor growth and lower metastases formation [24]. These discoveries triggered huge research efforts and, most importantly, fueled the idea of the Na⁺-K⁺-ATPase as a promising drug target for the treatment of different types of cancer. The mechanistic basis of these actions has been intensively studied in the last years: CGs inhibit cancer cell growth by various complex and cell-type dependent mechanisms, e.g. by inducing apoptosis or autophagy, by triggering mitochondrial dysfunction due to an accumulation of calcium, and/or by interfering with the mammalian target of rapamycin (mTOR) and NfκB signaling. Recent comprehensive reviews that summarize this vibrant field of research can be found under the references [25–30].

A New Frontier: The Anti-Inflammatory Potential of Cardio-Active Steroid Glycosides

A schematic overview of the subsequently described actions of CGs is provided in Fig. 2.

Cardio-active steroid glycosides exert anti-inflammatory actions in vivo

Acute inflammation

The first report on the influence of CGs on inflammatory processes was published in 1967: Lancaster and Vegad [31] found that pretreatment with ouabain (also known as g-strophanthin) (100 µg/kg bodyweight) decreased zymosan-induced early signs of inflammation (edema formation) in the skin and pleural cavity of sheep. De Vasconcelos et al. [32] tested the action of ouabain in different murine models of inflammation and algesia: The authors reported that ouabain (310 or 560 µg/kg bodyweight) decreased paw edema formation in mice induced by carrageenan, compound 48/80, zymosan, prostaglandin E2, and bradykinine. Also the concanavalin A-evoked peritoneal inflammation and the associated presence of leukocytes in the peritoneal cavity was reduced by the CG. Moreover, ouabain exhibited nociceptive actions in the acetic acid-induced writhing response and in the hot plate model. These findings were corroborated and expanded in a study by Leite et al. [33], who showed that ouabain (560 µg/kg bodyweight) decreased zymosan-induced peritonitis in mice. The compound reduced fluid extravasation (edema formation), leukocyte infiltration and the levels of the cytokines IL-1β and tumor necrosis factor α (TNFα), but did not influence the viability or function (phagocytosis) of leukocytes. The reduced leukocyte extravasation was attributed to a reduced migratory capacity evoked by ouabain. Moreover, ouabain was found to inhibit the zymosan-induced DNA-binding activity of the prototypical proinflammatory transcription factor NfκB. Taken together, these studies provide clear evidence that ouabain effectively prevents acute inflammation in vivo.
Infection-induced inflammation

The activation of inflammatory processes is a prerequisite for the successful elimination of pathogens. Anthony Esposito [34] published his findings on the action of digoxin (4 µg/kg bodyweight per day; ▶ Fig. 4) on the inflammatory response in murine pneumococcal pneumonia. The digoxin-treated animals exhibited a higher mortality rate, since they were unable to clear the infection. Interestingly, these mice showed a decreased recruitment of granulocytes and macrophages into the lung tissue, whereas, at least in vitro, digoxin did not affect the bacteria-killing action of alveolar macrophages. Jacob et al. [35] investigated the action of ouabain in murine leishmaniosis. The CG effectively prevented the infiltration of the peritoneal cavity with neutrophils and reduced the levels of TNFα and interferon γ (IFNγ) in the peritoneal exudate.

Chronic inflammatory autoimmune diseases

Regarding autoimmune diseases, digoxin was first evaluated in an animal model resembling multiple sclerosis: Huh and coworkers [36] were seeking for small molecule inhibitors of the retinoid acid receptor-related orphan nuclear receptor RORγt, a transcription factor required for the differentiation of TH17 cells. In the last years, these cells have been recognized to play a crucial role in autoimmune diseases. In a compound screening approach digoxin was discovered as specific inhibitor of RORγt. In fact, digoxin was found to inhibit murine T117 cell differentiation. In murine experimental autoimmune encephalomyelitis, a model of multiple sclerosis, digoxin (40 µg per mouse) delayed the onset and reduced the severity of the disease. Moreover, it strongly decreased the infiltration of the spinal cord with T117 cells. To overcome the limitations of CGs regarding their toxicity, the authors searched for chemical modifications of digoxin with a decreased affinity to the Na+-K+-ATPase and an augmented affinity to RORγt and identified the two compounds 20,22-dihydropinogoxin-21,23-diol and digoxin-21-salicylidene. Inspired by these important findings on the influence of CGs on T117 cells, the action of digoxin was tested in two further models of autoimmune diseases: Lee et al. [37] provided evidence for both a preventive and therapeutic action of digoxin (2 and 5 mg/kg bodyweight) on collagen-induced arthritis in mice. Digoxin lowered the joint inflammation and reduced the expression of IL-17, IL-1β, IL-6, TNFα, and IL-21. The digoxin-treated mice showed a decreased amount of T117 cells and an increased number of regulatory T cells. In vitro, digoxin suppressed the differentiation of T117 cells and reduced the production of immunoglobulins from murine B cells. Furthermore, digoxin was tested in experimental autoimmune uveitis in mice. Hinshaw et al. [38] found that digoxin (2 mg/kg bodyweight) reduced uveal inflammation and inhibited the differentiation of T117 cells. Unfortunately, this anti-inflammatory action was accompanied by a severe retinal degeneration process that led to the loss of vision. Obviously, digoxin is an improper compound to treat uveitis. These findings strengthen the approach of Huh et al. [36] to separate the actions on RORγt from those on the Na+-K+-ATPase.

Neuroinflammation

Forshammar et al. [39] reported about the beneficial action of CGs on LPS-activated astrocytes. Both a very low (1 pM) and a very high concentration (10 µM) of ouabain prevented the downregulation of the Na+-K+-ATPase, restored the disorganized F-actin cytoskeleton, and reduced the secretion of IL-1β. Forshammar and colleagues [40] also investigated the action of ouabain on rat microglia cells which represent resident brain macrophages. Interestingly, in contrast to astrocytes ouabain did not exert any influence on the LPS-induced secretion of cytokines from microglia. In vivo experiments in the context of neuroinflammation were performed by Kinoshita et al. [41] who evaluated the effect of oua-
bain (1.8 µg/kg bodyweight) on the hippocampus in a rat model of LPS-induced systemic inflammation/sepsis. Ouabain reduced the LPS-triggered activation of astrocytes, inhibited the raise of the proapoptotic protein Bax, and maintained the expression of the brain-derived neurotrophic factor which indicates a neuroprotective action. Furthermore, ouabain decreased the nuclear translocation of NFkB p65 and the degradation of IkB as well as the expression of IL-1β and of iNOS. Surprisingly, the elevated serum levels of corticosterone and TNFα were not altered by ouabain. Although neither LPS nor ouabain affected the activity of the hippocampal Na⁺-K⁺-ATPase, an alternative target which might be responsible for the observed pharmacological actions of ouabain was not presented.

Cystic fibrosis

Cystic fibrosis is a genetic disease caused by the lack or a malfunction of the chloride channel cystic fibrosis conductance regulator (CFTR). It is characterized by severe lung inflammation and high levels of the proinflammatory cytokine IL-8 in lung tissues. In 2004, Srivastava et al. [42] reported that different CGs (e.g. digitoxin, ouabain, oleandrin) suppress IL-8 hypersecretion in vitro in cystic fibrosis lung epithelial cells. With an IC₅₀ of 0.9 nM, digitoxin was the most active CG. Moreover, the authors tested the influence of digitoxin on the global gene expression in these cells and compared the gene expression patterns with those obtained upon restoral of the lacking CFTR ion channel. Interestingly, digitoxin could mimic the effect of this genetic restoral and the overlapping regulated genes were associated with the TNFα/NFκB pathway. This work was followed-up one year later: In a detailed mechanistic analysis, CGs were found to block the TNFα-dependent recruitment of the adaptor protein TNF receptor-associated death domain (TRADD) to the TNF receptor 1 (TNFR1) [43]. This explains the observed inhibitory action of digitoxin on the activation of IKK and on the phosphorylation and degradation of IkB.

Inspired by these two in vitro studies, Zeitlin et al. [44] initiated an explorative randomized controlled trial: 24 patients suffering from mild to moderate cystic fibrosis were treated over a period of 28 days with digitoxin (50 µg or 100 µg daily) or with placebo. As primary parameters, the concentration of IL-8 and the count of neutrophils in the sputum were analyzed. The therapy was safe, but steady state blood concentrations of digitoxin could not be achieved until the last few days of the treatment period. Since the duration of the trail was too short, there was no chance to reveal statistically significant outcomes. In fact, digitoxin did not alter IL-8 levels and only slightly reduced the neutrophil count. Larger and much longer studies are needed to answer the question as to whether CGs can beneficially influence inflammatory processes in cystic fibrosis patients.

Ouabain exerts immunomodulatory actions on leukocytes

A schematic overview of the subsequently described effects of CGs on the different types of leukocytes is provided in Fig. 5.

**Lymphocytes**

In 1968, Quastel and Kaplan [45] reported that ouabain (≥ 10 nM) blocks DNA synthesis induced by the mitogen phytohaemagglutinin in isolated human lymphocytes. As expected due to the inhibition of the Nα⁺-K⁺-ATPase, the effect could be reversed by increasing potassium concentrations in the culture medium. In a follow-up article [46], the authors corroborated and expanded these findings: ouabain (≥ 10 nM) did not only inhibit the mitogen-triggered DNA, but also the RNA and protein synthesis in lymphocytes. Of note, the inhibitory action of ouabain (1–10 μM) on the de novo protein synthesis was confirmed in a recent work by Takada et al. [47], who additionally showed that ouabain decreases the Nα⁺-dependent amino acid transport across cell membranes. However, the authors used a human lung carcinoma cell line in this study. After these initial reports, the field was not advanced for a decade. Then, in 1980, Szamel et al. [48] published an article in which they refined the findings of Quastel and Kaplan on the inhibition of ouabain on lymphocytes [45,46]. They analyzed the action of ouabain (20 nM) in dependence of the cell cycle status and showed that lymphocytes were most sensitive to ouabain within the S phase. In a further article, Szamel et al. [49] reported that ouabain (5 µM) is also able to block a very early event in the activation of lymphocytes, i.e. the altered metabolism of plasma membrane phospholipids (e.g. oleate incorporation), by inhibiting the acyl-CoA lyssolecithin-acyltransferase due to a functional and spatial association of this enzyme with Na⁺-K⁺-ATPase. Furthermore, ouabain (100 nM) reduced both the phytohaemagglutinin- and the phorbol ester-induced proliferation of peripheral blood lymphocytes [50]. Despite this inhibition of proliferation, the lymphocytes could still be activated.

As a consequence of the discovery of IL-2 as an important regulator of T cells in the late 1970s, Stoeck et al. [51] wondered...
whether the actions of ouabain on T lymphocytes could depend on alterations in the production of IL-2 and/or the response to this cytokine. Indeed, ouabain (100 µM) clearly decreased the mitogenic activity of IL-2 on T cells, but only moderately reduced the concanavalin A-triggered IL-2 production. These findings were confirmed by Lillehoj and Shevach [52] and studied in great detail by Dornand et al. [53]. Of note, ouabain, up to a concentration of 10 mM and a treatment period of 30 h, did not alter the viability of isolated human lymphocytes despite inhibition of the Na⁺-K⁺-ATPase [54].

In thymocytes, which represent hematopoietic progenitor cells in the thymus, a high concentration of ouabain (5 µM) inhibited the mitogen-induced proliferation [55]. Also a much lower concentration (100 nM) was able to decrease thymocyte activation via reducing the phosphorylation of MAPK p38 and the levels of the nuclear factor of activated T cells (NFAT) c1 [56]. In contrast, ouabain (100 nM) was reported to augment intracellular calcium concentrations and to enhance the expression of CD69, an indicator of thymocyte activation [57]. Moreover, ouabain and glucocorticoids synergistically increased the death of thymocytes [58, 59]. The pathophysiological meaning of these results, however, remains elusive.

Besides T lymphocytes, also B cells were found to be susceptible to ouabain: Mithorp et al. [60] demonstrated that ouabain (10 µM) inhibits the production of antibodies in splenocytes from pigs and rabbits, but not from mice.

Peripheral blood mononuclear cells

In the late 1990s, the influence of CGs on the cytokine secretion from PBMCs (monocytes and lymphocytes) was investigated in detail. These studies provided an interesting and very important insight into the differential actions of CGs: the influence of ouabain on the cytokine production of PBMCs strongly depends on their activation status. Based on the finding that the activity of the Na⁺-K⁺-ATPase is decreased in PBMCs from rheumatoid arthritis patients [61], Foey et al. [62] showed that mimicking this pathological situation in isolated human PBMCs from healthy donors by treating these cells with ouabain (100 nM) strongly augments IL-1β and TNFα, but suppresses IL-6 secretion. Thus, an impaired function of the Na⁺-K⁺-ATPase might be associated with the increased levels of proinflammatory cytokines in patients with rheumatoid arthritis. Matsumori et al. [63] also obtained PBMCs from healthy human donors. In their setting, the expression (mRNA and protein) of all three proinflammatory cytokines, IL-1β, TNFα, and IL-6, was increased by ouabain (100 nM). After activation of PBMCs with LPS, however, ouabain suppressed the production of IL-6 and TNFα. This was in line with the in vivo results: ouabain (1 mg/kg bodyweight) protected mice from the LPS-induced lethal toxicity and reduced the levels of IL-6 and TNFα. In a follow-up study, Matsumori et al. [64] could show that the calcium channel blocker amlodipine blocked the ouabain (1 µM)-triggered increase of IL-1α, IL-1β, and IL-6 in PBMCs from healthy human donors.

In 2008, Ihenetu et al. [65] confirmed the earlier reports on the action of CGs on human PBMCs: Both digoxin (<100 nM) and the endogenous digoxin-like immunoreactive factors (DLIF) inhibited the LPS-triggered activation of NFκB and the release of TNFα, IL-6, and IL-8.

Monocytes and macrophages

Sowa and Przewlocki [66] analyzed the release of nitric oxide (NO), an important defense mechanism of the innate immunity, from rat peritoneal macrophages. They found that ouabain (125–500 µM) does not affect basal NO generation, however, it strongly enhanced the LPS-induced NO formation. In monocytes purified from human PBMCs (healthy donors), ouabain treatment (100 nM) resulted in a decreased expression of the membrane-bound form of CD14 [67]. This effect was based on the activation of the epidermal growth factor receptor and p38 MAPK. Furthermore, ouabain (10 nM) augmented the intracellular calcium concentration, the surface levels of the activation markers CD69, HLA-DR, CD86 and CD80, the expression of IL-1β and TNFα, and the endocytic activity of these cells [68].

Neutrophils, natural killer cells and dendritic cells

Ward and Becker [69] found that ouabain inhibits the chemotactic migration of rabbit neutrophils, but the IC₅₀ value was quite high, ranging between 1–100 µM. De Moraes et al. [70] wondered whether natural killer cells are also prone to the action of ouabain. However, they could clearly demonstrate that ouabain does not influence these cells. Nascimento et al. [71] were the first to investigate the action of ouabain (100 nM) on human dendritic cells (DCs). Interestingly, the TNF-induced expression of CD83 and the production of IL-12, two features of DC maturation, were reduced by ouabain, whereas the capability of DCs to activate lymphocytes remained unaltered.

### Cardiac Glycosides Decrease the Inflammatory Activation of Endothelial Cells

Endothelial cells are not only a crucial part of the vascular system, but also regulate inflammatory processes, such as the extravasation of immune cells from the blood into the tissue. Up to now, only two studies have dealt with the actions of CGs on endothelial cells in the context of inflammation. Bereta et al. [72] showed that

<table>
<thead>
<tr>
<th>FIG. 5 Summary of the effects of ouabain on leukocytes.</th>
</tr>
</thead>
<tbody>
<tr>
<td><strong>lymphocytes</strong></td>
</tr>
<tr>
<td>• inhibition of proliferation as well as DNA, RNA and protein synthesis</td>
</tr>
<tr>
<td>• alterations in the metabolism of plasma membrane phospholipids</td>
</tr>
<tr>
<td><strong>peripheral blood mononuclear cells</strong></td>
</tr>
<tr>
<td>• stimulation of IL-1β, IL-6 and TNFα synthesis in non-activated cells</td>
</tr>
<tr>
<td>• inhibition of IL-6 and TNFα synthesis under pro-inflammatory conditions</td>
</tr>
<tr>
<td><strong>other types of leukocytes</strong></td>
</tr>
<tr>
<td>• inhibition of neutrophil migration</td>
</tr>
<tr>
<td>• no effect on natural killer cells</td>
</tr>
<tr>
<td>• modulation of dendritic cell maturation</td>
</tr>
</tbody>
</table>


**Summary of the effects of ouabain on leukocytes.**
ouabain activates NFκB and increases the expression of the NFκB target genes iNOS and VCAM-1 in murine brain endothelial cells. Furthermore, digitoxin was found to strongly inhibit the IL-1β-induced expression of an important proinflammatory mediator, the monocyte chemoattractant protein-1 (MCP-1), and of VCAM-1 [73]. Most importantly, this study also showed that digitoxin inhibits the adhesion of monocytes to endothelial cells. Regarding the underlying mechanisms, digitoxin blocked the NFκB cascade as well as the activation of the extracellular signal-regulated kinase (ERK), but not of p38 MAPK and c-Jun N-terminal kinase (JNK). Moreover, digitoxin augmented the activity of the endothelial NO synthase (eNOS) and prevented the TNFα-induced expression of an important proinflammatory mediator, the monocyte chemoattractant protein-1 (MCP-1), and of VCAM-1 [73]. Most importantly, this study also showed that digitoxin inhibited the adhesion of monocytes to endothelial cells. Regarding the underlying mechanisms, digitoxin blocked the NFκB cascade as well as the activation of the extracellular signal-regulated kinase (ERK), but not of p38 MAPK and c-Jun N-terminal kinase (JNK). Moreover, digitoxin augmented the activity of the endothelial NO synthase (eNOS) and prevented the TNFα-caused apoptosis of endothelial cells via activation of the phosphatidyl-inositol-3-kinase (PI3K)/Akt pathway.

Summary and Conclusion

The first report on the anti-inflammatory potential of CGs dates back 50 years. Since then, CGs have been tested in several animal models of inflammation ranging from acute and infection-triggered models to chronic inflammatory disease. Despite this knowledge, a translation into clinical applications has not yet occurred. This might be due to the following unresolved issues that call for further research:

CGs have been found to exert anti-inflammatory actions not only by targeting the Na⁺-K⁺-ATPase, but also via different systems, such as RORγt. Thus, the role of the ATPase for the anti-inflammatory action of CGs is still unclear. This issue is of high relevance, since the toxic activities of CGs are mainly ascribed to this enzyme. The separation of the cardiovascular/Na⁺-K⁺-ATPase-inhibiting effects from the anti-inflammatory activities has been shown to be feasible. Further approaches into this direction are, thus, of great importance and will strongly advance the field.

Most of the studies analyzed the actions of CGs on leukocytes. A major drawback of these analyses is the fact that highly different concentrations of CGs which do not properly reflect the in vivo situation were used. Physiological levels of endogenous CGs range from 10 pM to 1 nM. In contrast, both the plasma concentrations of endogenous CGs during stress conditions and the serum levels of administered CGs during pharmacotherapy are much higher, namely in the range of 100 nM [74, 75]. Also species differences have not yet been considered systematically. Thus, from the clinical perspective, some of the reported effects and presented mechanisms might only be of limited relevance.

Up to now, only one very small and preliminary clinical trial has investigated the anti-inflammatory potential of CGs (in the context of cystic fibrosis). Unfortunately, due to the very limited period of drug treatment, the study could not demonstrate significant benefits of CGs. However, the effort to perform clinical trials is highly appreciated and needs to be expanded urgently.

Interestingly, although the pathogenesis of many types of tumors is closely associated with inflammatory processes, no data about the action of CGs on the interface between cancer and inflammation exist. As a future direction, it would be of great interest to investigate both the pathophysiological role of endogenous CGs and the pharmacological action of exogenously applied CGs in the context of inflammation-associated tumorigenesis.

Taken together, CGs possess a very interesting anti-inflammatory potential. However, the usage of unmodified CGs seems to be very unlikely due to their narrow therapeutic index. The rational design of CG analogs with decreased cardiac and augmented anti-inflammatory actions is a young but promising research area that will greatly advance the field.

Conflict of Interest

The authors declare no conflict of interest.

References


Fürst R et al. New Knowledge About... Planta Med 2017; 83: 977–984

This document was downloaded for personal use only. Unauthorized distribution is strictly prohibited.
Calderón-Montaño JM, Burgos-Morón E, Orta ML, Maldonado-Navas D, Fürst R et al. New Knowledge About... 


Shiratori O. Growth inhibitory effect of cardiac glycosides and aglycones on neoplastic cells: *in vitro* and *in vivo* studies. Gan 1967; 58: 521–528


Samzel M, Resch K. Inhibition of lymphocyte activation by ouabain. Interference with the early activation of membrane phospholipid metabolism. Biochim Biophys Acta 1981; 702: 297–301


Samzel M, Schneider S, Resch K. Functional interrelationship between (Na(+) + K(+)−)−ATPase and lysocellitin acyltransferase in plasma membranes...


