



Antibacterial and Cytotoxicity of Root Canal Sealer with the Addition of Chitosan Nanoparticle at Various Concentrations

Diatri Nari Ratih¹ Ema Mulyawati¹ Rika Kurnia Santi² Yulita Kristanti¹

¹Department of Conservative Dentistry, Faculty of Dentistry, Universitas Gadjah Mada, Yogyakarta, Indonesia

²Former Student Specialist, Study Program of Conservative Dentistry, Faculty of Dentistry, Universitas Gadjah Mada, Yogyakarta, Indonesia Faculty of Dentistry, Universitas Gadjah Mada, Yogyakarta, Indonesia

Address for correspondence Diatri Nari Ratih, DDs, MDSc, PhD, Jl. Denta 1, Sekip Utara, Yogyakarta, Indonesia (e-mail: diatri@ugm.ac.id).

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Abstract

Objectives The aim of this study was to evaluate the addition of chitosan nanoparticle with concentrations of 0, 10, 20, and 30% to the epoxy resin-based (ERB) sealer on its antibacterial and cytotoxicity effect.

Methods and Materials This research was divided into two studies, the first study was the addition of chitosan with a concentration of 0% (as control), 10, 20, and 30% to an ERB sealer on its antibacterial effect, and the second study was on its cytotoxicity. An agar diffusion test was employed to determine the antibacterial effect on *Enterococcus faecalis*. An MTT (3-[4,5-dimethylthiazol-2-yl]-2,5-diphenyl tetrazolium bromide) assay was utilized to test the cytotoxicity by evaluating cell viability.

Statistical Analysis One-way analysis of variance and Tukey's test ($\alpha = 0.05$) were used to analyze data obtained from each evaluation with a significance level of 95%.

Results The addition of chitosan nanoparticles at concentrations 10, 20, and 30% produced a greater inhibition zone of *E. faecalis* ($p < 0.05$), however, had less cytotoxicity compared with no addition of chitosan (0%) ($p < 0.05$).

Conclusion The addition of chitosan nanoparticles at concentrations 10, 20, and 30% to the ERB sealer produced greater antibacterial and less cytotoxicity compared with no addition of chitosan (0%).

Keywords

- ▶ antibacterial
- ▶ chitosan nanoparticle
- ▶ cytotoxicity
- ▶ root canal sealer

Root canal obturation is an important aspect that influences the successful of root canal treatment. Root canal sealer as material for obturation should have biocompatibility, antibacterial, good apical closure ability, adequate flow, insoluble in tissue fluid, and not staining teeth.¹ Root canal sealer must enable to eradicate the remains of microorganisms that cannot be removed during the root canal preparation and sterilization procedure. Of the numerous types of microorganisms, *Enterococcus faecalis* is a bacteria that is mostly found in the failure of endodontic cases.²

In clinical conditions, the obturation of the root canal can be excessive, resulting in contacting of root canal sealers with the periapical tissue. The tissue response to the material can impair the result of root canal treatment.³ Therefore, the biocompatibility of the sealer is essential for successful root canal treatment since the release of sealer substances can induce reactions in the periapical tissue.⁴

Currently, the epoxy resin-based (ERB) sealer is popular as obturation material due to several beneficial properties such as good apical closure, good flow, low setting time, solubility,

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and inexpensive.⁵ However, previous studies reported that resin sealers have low antibacterial properties; hence, the addition of antibacterial components to sealers has the potential to increase their antibacterial efficacy.^{6,7}

Lately, chitosan has been frequently used in the health field for its many advantageous properties. Chitosan is a nontoxic natural polysaccharide, produced from the deacetylation of chitin obtained from the shells of crustaceans. Chitosan broadly exists in nature, is inexpensive, and possesses chelating properties.⁸ Former studies have explained that chitosan yielded antibacterial properties against *E. faecalis* and *Candida albicans*.^{9,10} Due to the many advantages of chitosan, hence chitosan nanoparticles are incorporated into ERB sealer to enhance its antibacterial efficacy. Previous studies have shown that the addition of chitosan nanoparticles to ERB sealer did not affect the physical properties of ERB sealer, which exhibited acceptable sealer for obturation material.^{11–13}

However, until now there is deliberation about the appropriate concentration of chitosan nanoparticles, which should be added to sealer. Thus, the purpose of this study was to evaluate the addition of nanoparticle chitosan with concentrations of 0, 10, 20, and 30% to the ERB sealer on its antibacterial and cytotoxicity effect. The null hypothesis was that no difference occurred in antibacterial and cytotoxicity effect with the addition of nanoparticle chitosan with concentrations of 0, 10, 20, and 30% to the ERB sealer.

Materials and Methods

The research protocol was approved by the institutional ethics committee under the number 00483/KKEP/FKG-UGM/EC/2020. This study evaluated the addition of chitosan nanoparticles (NHI, Tangerang, Indonesia) with concentrations of 0 (as control), 10, 20, and 30% to ERB sealer (AH 26, De Trey, Dentsply, Konstanz, Germany), and was divided into two evaluations, namely, antibacterial and cytotoxicity test.

Antibacterial Evaluation

The antibacterial study used 24 samples assigned into 4 groups of each 6 samples. The methodology for evaluating the antibacterial effect was performed using agar diffusion test (ADT). The procedure for the ADT was modified from Silva et al.¹⁴ A 24-hour culture of *E. faecalis* (strain ATCC 29212) in Brain Heart Infusion agar was employed to make a bacterial suspension encompassing 108 viable bacterial cells per milliliter. Standardization of 0.5 suspensions with a spectrophotometer and McFarland scale was then performed. *E. faecalis* suspension was planted using a sterile swab on Muller Hinton Agar plates and incubated for 24 hours at 37°C. Afterward desiccating for 10 minutes at 36°C, a sterile glass tube was employed to produce 4 agar wells (6 mm diameter × 4 mm depth) of each petri.

The wells were filled with the materials described as follows: group 1: 0% chitosan nanoparticle + ERB sealer as a control group, group 2: 10% chitosan nanoparticles + ERB sealer, group 3: 20% chitosan nanoparticles + ERB sealer, and group 4: 30% chitosan nanoparticles + ERB sealer. Before

placing the material into the well of each petri, the sealer was mixed concordant to the instruction of the manufacturer until a homogeneous consistency was obtained. All sealers were then put in the well according to their respective groups. Furthermore, all petri were stored in an incubator at 37°C for 24 hours. After incubation, the zones of microbial growth inhibition were assessed at the radical zone using a sliding caliper on a millimeter scale with a precision of 0.02 mm, based on ▶Fig. 1 and the formula developed by Levinson.¹⁵

The measurement of radical zone of the well: $\frac{1}{2} (AB - ab) + \frac{1}{2} (CD - cd) + \frac{1}{2} (EF - ef)/3$.

Cytotoxicity Evaluation

Samples Preparation

All samples were created according to the International Organization for Standardization (ISO) 10993–12.¹⁶ This evaluation used 48 samples divided into four concentration groups of chitosan nanoparticles as in antibacterial evaluation. Each concentration group consisted of 12 samples. The mixed sealers were located into 48 Teflon molds (5 mm in diameter and 2 mm in height), permitted to set in an incubator at 37°C for 24 hours, crushed into tiny particles with mortar and pestle, and located in the cell culture dish.

Cell Culture

This present study used Vero cells (ECACC, Public Health England, London, U.K.) for cytotoxicity evaluation, which was attained from Cell Culture Laboratory, LPPT UGM, Yogyakarta, Indonesia. The procedure for the cytotoxicity evaluation was adapted from Catunda et al.¹⁷ The cells were cultured in M199 medium (Gibco, Waltham, MA) at 37°C in a humidified 5% CO₂ atmosphere. The culture medium

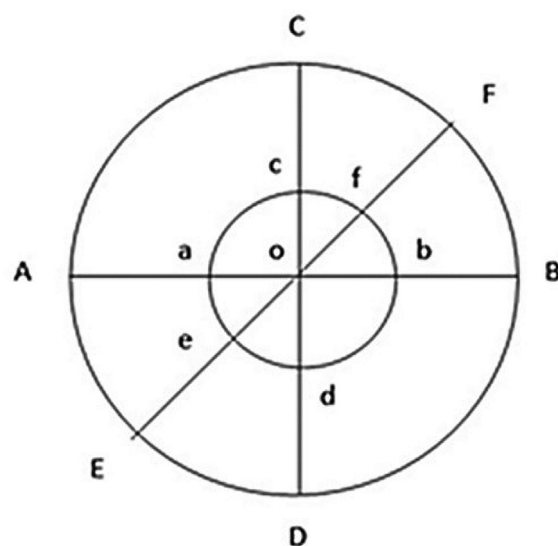


Fig. 1 The measurement of inhibition zone; Point O: center point of the well; Line AB, CD, and EF: diameter of radical zone; Line ab, cd, and ef: diameter of the well (6 mm).

utilized was Dulbecco's modified Eagle medium (DMEM, Sigma Chemical Co., St Louis, MO) accompanied with 10% fetal bovine serum (Sigma Aldrich, St Louis, MO) and 1% antibiotic-antimycotic solution (10,000 UI of penicillin, 10 mg of streptomycin in 0.9% sodium chloride (Sigma Chemical Co.). Cultures were delivered with fresh medium every 3 days until an adequate number of cells was attained. Following dilution with a ratio of 1:10 in Trypan Blue Dye (10 μ L of cells in 90 μ L of Trypan Blue), the cells were calculated in a Neubauer chamber. The cells (2×10^5 cells/mL of DMEM per well) were then moved to the culture plate (Sigma-Aldrich, Munich, Germany) and incubated for 24 hours at 37°C in 5% CO₂ and 95% air. All samples were prepared by the same operator in a laminar flow chamber (Biobase, Jinan, Shandong, China), and exposed to ultraviolet light for 45 minutes.

Extracts and Experimental Groups

The extract was made by submerging the sample in DMEM kept in a Falcon tube for 24, 48, or 72 hours to condition the media. The conditioned DMEM was filtered (0.22 μ m syringe filter; TPP, Darmstadt, Germany) to eradicate dense components.

Cytotoxicity Assay

In 96-well culture plates (Thermo-Fisher Scientific, Waltham, MA), 2×10^5 cells in 1 mL of DMEM per well were cultured and grown to subconfluent monolayers for 24 hours. The culture medium was then altered with equal volumes (25 μ L) of sealer extracts (conditioning medium), using the culture medium itself as a negative control. The assessment of the cytotoxic activity was substantiated by the colorimetric method bromide (3-{4,5-dimethylthiazol-2-yl}-2,5-diphenyl tetrazolium bromide) (MTT). After 24 hours of incubation, 25 μ L (5 mg/mL) of MTT solution was inserted to each well, and the plates were incubated for 3 hours. The MTT was then detached and 25 μ L per well dimethyl sulfoxide (Sigma Chemical Co.) was put into each well to dissolve the formazan crystals.

Based on ISO 10993-12,¹⁶ a reduction in the number of alive cells leads to a decline in the metabolism in the sample. Such reduction is directly associated with the quantity of blue-violet formazan created as observed by the optical density at 570 nm using enzyme-linked immunosorbent assay reader (Tecan Spark, Tecan Trading AG, Switzerland). The percentage of viable cells in each well was calculated as below¹⁸:

Absorbance of Sample

% Cell viability = (Absorbance of sample/Absorbance of control) \times 100%.

Absorbance of Control

The lesser viability % value means higher cytotoxic potential. The cell viability was categorized as noncytotoxic (more than 90% cell viability), slightly cytotoxic (60–90% cell viability), moderately cytotoxic (30–59% cell viability), and severely cytotoxic (less than 30% cell viability).¹⁹

Statistical Analysis

Data obtained from each evaluation were assessed separately using the Shapiro–Wilk normality test for normal data distribution and Levene's test for homogeneity of variances. Then, each evaluation data were analyzed using analysis of variance (ANOVA), followed by Tukey's test with $p < 0.05$ considered as a significant difference. The statistical analysis was processed and analyzed using the SPSS Version 23 program.

Results

The mean inhibition zone diameter of *E. faecalis* was the highest at 10% concentration and the lowest was at 0% concentration, whereas the greatest toxicity was at 0% concentration and the least toxicity was at 30% concentration (►Table 1). The cell viability of representative samples can be seen in ►Fig. 2, which exhibits the more the quantity of formazan that occurred (apparently directly proportional to the number of viable cells) the more the cell viability.

One-way ANOVA analysis exhibited that the addition of chitosan nanoparticles at concentrations 10, 20, and 30% to ERB sealer generated a greater inhibition zone of *E. faecalis* but less cytotoxicity than no addition of chitosan (0%) ($p < 0.05$). Tukey's test revealed that the addition of chitosan nanoparticles at concentrations 10, 20, and 30% caused significant increase in antibacterial efficacy to *E. faecalis* compared with no addition of chitosan nanoparticles ($p < 0.05$); however, no significant differences occurred among concentrations 10, 20, and 30% ($p > 0.05$). The addition of chitosan nanoparticles at concentrations 10, 20, and 30% to the EBS sealer produced less cytotoxicity compared with concentration 0% ($p < 0.05$), while between 20 and 30% concentrations, no significant difference occurred ($p > 0.05$).

Table 1 Mean and standard deviation of *Enterococcus faecalis* inhibition zone and cell viability of chitosan nanoparticles addition at concentration 0, 10, 20, and 30% to epoxy resin-based sealer

Chitosan nanoparticles concentrations	<i>Enterococcus faecalis</i> inhibition zone (mm)	Cell viability (%)
0%	4.82 \pm 0.54 ^a	61.90 \pm 1.20 ^a
10%	9.94 \pm 0.63 ^b	73.08 \pm 1.07 ^b
20%	9.63 \pm 0.35 ^b	74.08 \pm 0.90 ^c
30%	9.60 \pm 0.25 ^b	75.42 \pm 1.43 ^c

Note: Different letters indicate that there were statistically significant differences.

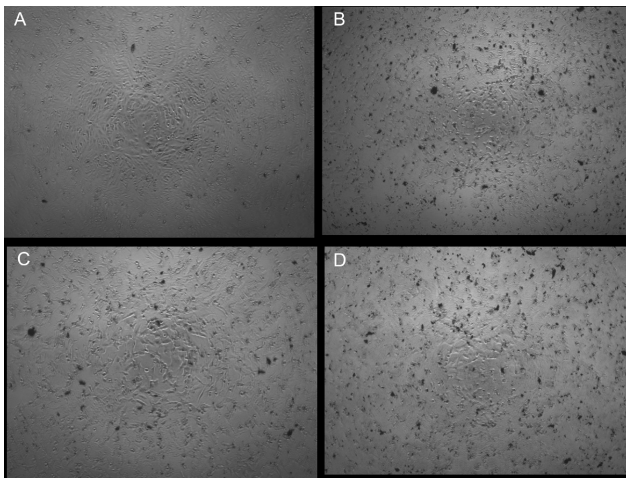


Fig. 2 Micrographs showed cell viability of chitosan nanoparticles addition at concentration 0% (A), 10% (B), 20% (C), and 30% (D) to epoxy resin-based sealer. The quantity of formazan (related to the number of viable cells) is showed as black spot as monitored by the optical density (OD) at 570 nm.

Discussion

The most crucial requirements of sealers for root canal obturation are antibacterial and biocompatible; therefore, this study was conducted to evaluate the antibacterial effect and biocompatibility with cytotoxicity test.²⁰ Previous studies stated that the ERB sealer has a minimal antibacterial property.⁵ However, in the clinical condition, this minimal antibacterial property of ERB sealer is inadequate to protect against persistent bacterial infections such as *E. faecalis* since this microorganism is usually related to the etiology of tenacious periradicular lesions.²¹ Additionally, in most studies of chitosan, the antibacterial efficacy of chitosan nanoparticles has already been proven^{6,9,14}; therefore, in this study, it was verified that chitosan nanoparticles with the concentration of 10, 20, and 30% added to ERB sealer would enhance the antibacterial efficacy of ERB sealer.

The efficacy of chitosan to eliminate microorganisms can be explained by two mechanisms. The first mechanism is that chitosan nanoparticles bind to the target cell membrane through electrostatic forces resulting in changes in the membrane, depolarization, and loss of membrane integrity. The important cell functions of the bacteria are disrupted such as respiration, nutrient transport, and energy transduction resulting in bacterial cell death. The second mechanism is the production of free radicals such as reactive oxygen species which can affect the resistance of bacterial cells by inhibiting protein function and damaging deoxyribonucleic acid.²²

This present study revealed the development of the inhibition zone of ERB sealer also occurred with no addition of chitosan nanoparticles. It might be due to the presence of antibacterial components, such as hexamethylenetetramine and formaldehyde in the ERB sealer used in this study (AH 26).²³ The more concentration addition of the chitosan to ERB sealer may not affect the antibacterial activity as shown in this study.²⁴

ADT was selected to evaluate antibacterial properties in this study since this technique has been widely used to evaluate the antibacterial activity of sealers and offers many advantages, such as simplicity, low cost, the ability to test enormous numbers of microorganisms and antimicrobial agents, and the ease to interpret results provided.²⁵ In addition, this method allows measurement of the activity of soluble and degradable components of the tested material, such as chitosan nanoparticles used in this study incorporated to ERB in the surrounding medium, indicated by an inhibition halo.²⁶

Besides antibacterial properties, sealers should be biocompatible since they enable to contact with periapical tissue.⁴ This study showed that all concentrations of chitosan addition produced a cytotoxic effect, although with different degrees of toxicity, and all concentrations including without the addition of chitosan were classified as slightly cytotoxic (cell viability ranged from 60 to 90%).¹⁹ ERB sealer without addition of chitosan (0%) produced the highest cytotoxicity compared with the addition of other concentrations of chitosan nanoparticles. It can be explained that the ERB sealer employed in this study (AH 26) consisted of formaldehyde, epoxy resin, and hexamethylenetetramine, besides being antibacterial, they are also toxic.^{23,27}

This cytotoxic effect in this study was only observed after 24 hours. This effect may decline if the longer observation is undertaken because the release of formaldehyde is reduced on the 7th day.²⁸ The cytotoxic influence may also be associated with the solubility of the material used, thus the leaching out of sealer components is influenced by its solubility.²⁹ In this study, the addition of 30% chitosan nanoparticles induces the lowest cytotoxic effect, perhaps the more chitosan, which is biocompatible in nature, added to the ERB sealer generated less quantity of the cytotoxicity components (formaldehyde, epoxy resin, and hexamethylenetetramine) in the sealer mixture. Consequently, the leaching of the cytotoxic components also decreased.³⁰

Conclusion

The addition of chitosan nanoparticles at concentrations 10, 20, and 30% to the ERB sealer produced greater antibacterial and less cytotoxicity compared with no addition of chitosan (0% concentration).

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Conflict of Interest

None declared.

References

- Chandra SS, Shankar P, Indira R. Depth of penetration of four resin sealers into radicular dentinal tubules: a confocal microscopic study. *J Endod* 2012;38(10):1412–1416
- Dalmia S, Gaikwad A, Samuel R, Aher G, Gulve M, Kolhe S. Antimicrobial efficacy of different endodontic sealers against

- Enterococcus faecalis*: an *in vitro* study. J Int Soc Prev Community Dent 2018;8(02):104–109
- 3 Torres-Rosas R, Torres-Gómez N, Moreno-Rodríguez A, García-Contreras R, Argueta-Figueroa L. Anti-Inflammatory and antibacterial activity of the chitosan/chlorhexidine gel commercial preparation for postexodontia treatment: an *in vitro* study. Eur J Dent 2020;14(03):397–403
 - 4 Kaur A, Shah N, Logani A, Mishra N. Biotoxicity of commonly used root canal sealers: a meta-analysis. J Conserv Dent 2015;18(02):83–88
 - 5 Fonseca DA, Paula AB, Marto CM, et al. Biocompatibility of root canal sealers: a systematic review of *in vitro* and *in vivo* studies. Materials (Basel) 2019;12(24):1–34
 - 6 Huang Y, Li X, Mandal P, et al. The *in vitro* antimicrobial activities of four endodontic sealers. BMC Oral Health 2019;19(01):118–125
 - 7 AL-Haddad A, Che Ab Aziz ZA. Bioceramic-based root canal sealers: a review. Int J Biomater 2016;2016:9753210
 - 8 Devadiga SB, Shetty D, Lakshmi G, Packiyam JE. Synthesis of chitosan silver nanoparticles from chitin of crustacean shell and its applications. Int J Curr Res Chem Pharmac Sci 2016;3(05):1–5
 - 9 Del Carpio-Perochena A, Kishen A, Shrestha A, Bramante CM. Antibacterial properties associated with chitosan nanoparticle treatment on root dentin and 2 types of endodontic sealers. J Endod 2015;41(08):1353–1358
 - 10 Aydın ZU, Özyürek T, Keskin B, Baran T. Effect of chitosan nanoparticle, QMix, and EDTA on TotalFill BC sealers' dentinal tubule penetration: a confocal laser scanning microscopy study. Odontology 2019;107(01):64–71
 - 11 Enggardipta RA, Untara RTE, Santosa P, Kartikaningtyas AT, Widyastuti A, Ratih DN. Apical sealing ability of chitosan nanoparticles in epoxy-resin-based endodontic sealer. MKGI 2019;5(02):69–74
 - 12 Ratih DN, Enggardipta RA, Kusumo ANH, Hadriyanto W. Setting time, flowability, and solubility of epoxy resin-based sealer mixed with chitosan nanoparticles. Int J Appl 2021;13(02):122–126
 - 13 Putranto AW, Suprastiwi E, Meidyawati R, Agusnar H. Characterization of novel cement-based carboxymethyl chitosan/amorphous calcium phosphate. Eur J Dent 2022;16(04):809–814
 - 14 Silva AR, Santos EB, Pinto SCS, Gomes JC, Vaz IP, Carvalho MF. Antimicrobial effect and transdentinal diffusion of new intracanal formulations containing nitrofurantoin or doxycycline. Braz Dent J 2014;25(05):425–429
 - 15 Levinson W. Review of Medical Microbiology and Immunology. 9th ed. New York: McGraw Hill; 2006:278–300
 - 16 International Organization For Standardization. Biological evaluation of medical devices - Part 12: Sample preparation and reference materials. Geneva ISO 10993–12; 2012
 - 17 Catunda RQ, Vieira JRC, de Oliveira EB, da Silva EC, Brasil VLM, Perez DC. Cytotoxicity evaluation of three dental adhesives on Vero cells *in vitro*. J Clin Exp Dent 2017;9(01):e61–e66
 - 18 Badole GP, Warhadpande MM, Meshram GK, et al. A comparative evaluation of cytotoxicity of root canal sealers: an *in vitro* study. Restor Dent Endod 2013;38(04):204–209
 - 19 Ahrari F, Tavakkol Afshari J, Poosti M, Brook A. Cytotoxicity of orthodontic bonding adhesive resins on human oral fibroblasts. Eur J Orthod 2010;32(06):688–692
 - 20 Monajemzadeh A, Ahmadi Asoor S, Aslani S, Sadeghi-Nejad B. *In vitro* antimicrobial effect of different root canal sealers against oral pathogens. Curr Med Mycol 2017;3(02):7–12
 - 21 Gjorgievska ES, Nicholson JW, Coleman NJ, Booth S, Dimkov A, Hurt A. Component release and mechanical properties of endodontic sealers following incorporation of antimicrobial agents. BioMed Res Int 2017;2017:2129807
 - 22 Nair N, James B, Devadathan A, Johny MK, Mathew J, Jacob J. Comparative evaluation of antibiofilm efficacy of chitosan nanoparticle and zinc oxide nanoparticle-incorporated calcium hydroxide-based sealer: an *in vitro* study. Contemp Clin Dent 2018;9(03):434–439
 - 23 Soliman AY, Roshdy NB, Lutfy RA. Assessment of the effect of chitosan nanoparticles on the ultra-structure of dentinal wall: a comparative *in vitro* study. Acta Sci Dent Sci 2020;4(01):32–38
 - 24 Poggio C, Trovati F, Ceci M, Colombo M, Pietrocola G. Antibacterial activity of different root canal sealers against *Enterococcus faecalis*. J Clin Exp Dent 2017;9(06):e743–e748
 - 25 Balouiri M, Sadiki M, Ibnsouda SK. Methods for *in vitro* evaluating antimicrobial activity: a review. J Pharm Anal 2016;6(02):71–79
 - 26 Matalon S, Weiss EI, Gorfil C, Noy D, Slutzky H. *In vitro* antibacterial evaluation of flowable restorative materials. Quintessence Int 2009;40(04):327–332
 - 27 Teixeira L, Basso FG, Hebling J, et al. Cytotoxicity evaluation of root canal sealers using an *in vitro* experimental model with roots. Braz Dent J 2017;28(02):165–171
 - 28 Jung S, Sielker S, Hanisch MR, Libricht V, Schäfer E, Dammaschke T. Cytotoxic effects of four different root canal sealers on human osteoblasts. PLoS One 2018;13(03):e0194467
 - 29 Graunaite I, Lodiene G, Arandarcikaite O, Pukalskas A, Machiulskiene V. Leachables and cytotoxicity of root canal sealers. J Oral Sci 2018;60(03):381–387
 - 30 Sajani AR, Hegde MN. Leaching of monomers from bulk-fill composites: an *in vitro* study. J Conserv Dent 2016;19(05):482–486