Plasmid-Mediated Fluoroquinolone Resistance in Pseudomonas aeruginosa and Acinetobacter baumannii

P. V. Geetha1 K. V. L. Aishwarya1 M. Shanthi1 Uma Sekar1

1 Department of Microbiology, Sri Ramachandra Institute of Higher Education and Research, Porur, Chennai, Tamil Nadu, India

Address for correspondence Geetha P. V., MSc, Department of Microbiology, Sri Ramachandra Institute of Higher Education and Research, Porur, Chennai, 600116, Tamil Nadu, India (e-mail: gethu16@gmail.com).

Introduction Pseudomonas aeruginosa and Acinetobacter baumannii are important pathogens in health care–associated infections. Fluoroquinolone resistance has emerged in these pathogens. In this study, we aimed to determine the occurrence of plasmid-mediated quinolone resistance (PMQR) determinants (qnrA, qnrB, qnrS, aac(6′)-Ib-cr, oqxAB, and qepA) by polymerase chain reaction (PCR) and the transmissibility of plasmid-borne resistance determinants in clinical isolates of P. aeruginosa and A. baumannii.

Materials and Methods The study included P. aeruginosa (85) and A. baumannii (45) which were nonduplicate, clinically significant, and ciprofloxacin resistant. Antibiotic susceptibility testing was done by disk diffusion method for other antimicrobial agents, namely amikacin, ceftazidime, piperacillin/tazobactam, ofloxacin, levofloxacin, and imipenem. Minimum inhibitory concentration of ciprofloxacin was determined. Efflux pump activity was evaluated using carbonyl-cyanide m-chlorophenylhydrazone (CCCP). The presence of PMQR genes was screened by PCR amplification. Transferability of PMQR genes was determined by conjugation experiment, and plasmid-based replicon typing was performed.

Results Resistance to other classes of antimicrobial agents was as follows: ceftazidime (86.9%), piperacillin/tazobactam (73.8%), imipenem (69.2%), and amikacin (63.8%). The minimal inhibitory concentration (MIC)50 and MIC90 for ciprofloxacin were 64 and greater than or equal to 256 µg/mL, respectively. There was a reduction in MIC for 37 (28.4%) isolates with CCCP. In P. aeruginosa, 12 (14.1%) isolates harbored qnrB, 12 (14.1%) qnrS, 9 (10.5%) both qnrB and qnrS, 66 (77.6%) aac(6′)-Ib-cr, and 3 (3.5%) oqxAB gene. In A. baumannii, qnrB was detected in 2 (4.4%), 1 (2.2%) harbored both the qnrA and qnrS, 1 isolate harbored qnrB and qnrS, 21 (46.6%) aac(6′)-Ib-cr, and 1 (2.2%) isolate harbored oqxAB gene. Notably, qepA gene was not detected in any of the study isolates. Conjugation experiments revealed that 12 (9.2%) were transferable. Of the transconjugants, seven (58.3%) belonged to IncFII type plasmid replicon, followed by four (33.3%) IncA/C and one (8.3%) IncFIC type.

Conclusion The plasmid-mediated resistance aac(6′)-Ib-cr gene is primarily responsible for mediating fluoroquinolone resistance in clinical isolates of P. aeruginosa and A. baumannii. The predominant plasmid type is IncFII.
Introduction

Fluoroquinolones are synthetic antimicrobial agents with a broad spectrum of activity. They are effective against a wide range of gram-negative and gram-positive pathogenic bacteria. Over the past few years, fueled by their widespread use, resistance to fluoroquinolones has raised globally. An important resistance mechanism to fluoroquinolones is described by mutations in the quinolone resistance-determining regions of gyrA and topoisomerase encoding genes. Another well-known fluoroquinolone resistance mechanism is the decreased intracellular drug accumulation by upregulation of efflux pumps or decreased expression of outer membrane porin. The emergence of plasmid-mediated quinolone resistance (PMQR) has been reported since 1998. These are horizontally transferable plasmid-mediated quinolone resistance (PMQR) has been reported since 1998. These are horizontally transferable genes.

Materials and Methods

Bacterial Isolates

The study included P. aeruginosa (85) and A. baumannii (45) which were nonduplicate, clinically significant and ciprofloxacin resistant (as determined by disc diffusion test) and obtained from clinical specimens of hospitalized patients at university teaching hospital in South India. They were collected over a period of 1 year from July 2014 to June 2015. They were obtained from clinical specimens such as urine (5), exudative samples (66), respiratory secretions (47), and blood stream (12). The isolates were identified up to species level by automated system (VITEK2 GN-card; BioMerieux, Brussels, Belgium) and/or standard biochemical tests.

Antimicrobial Susceptibility Testing

Antibiotic susceptibility testing was done by Kirby–Bauer disc diffusion method for the following antimicrobials: cefazidime (30 µg), piperacillin/tazobactam (30 µg), imipenem (10 µg), amikacin (30 µg), levofloxacin (5 µg), and ofloxacin (5 µg) (Himedia Laboratories, India). The minimal inhibitory concentration (MIC) of ciprofloxacin was determined by agar dilution technique according to CLSI 2017 guidelines. ATCC Escherichia coli 25922 was used as control for both disc diffusion method and MIC determination.

Phenotypic Detection of Efflux Pump Activity

To detect the presence of efflux pump mechanism, carbonylcyanide m-chlorophenylhydrazone (CCCP), the efflux pump inhibitor was added to each Muller–Hinton (MH) agar plate containing 0.125 to 256 μg/mL of ciprofloxacin. The fixed concentration of CCCP in the MH agar was 20 µg/mL. The MIC with CCCP incorporated was determined in twofold serial dilutions as for the antibiotic (CLSI 2017). A plate without antibiotic and containing only CCCP inhibitor was used as control. The criteria for the presence of efflux pump activity was based on a fourfold decrease in MIC of ciprofloxacin on addition of CCCP.

Polymerase Chain Reaction

The DNA of the study isolates was extracted by the boiling method. The amplification of qnr genes (qnrA, qnrB, and qnrS) was performed by multiplex PCR using the cyclic profile: initial denaturation at 94°C for 7 minutes; denaturation at 94°C for 50 seconds, annealing at 53°C for 40 seconds, and elongation at 72°C for 60 seconds, repeated for 35 cycles, and a final extension at 72°C for 5 minutes. The PCR conditions for acc(6‘)-Ib-cr were: initial denaturation at 94°C for 7 minutes, denaturation at 94°C for 50 seconds, annealing at 55°C for 40 seconds, and elongation at 72°C for 60 seconds, repeated for 35 cycles, and a final extension at 72°C for 5 minutes. The PCR cyclic parameters for oqxAB were as follows: initial denaturation at 95°C for 15 minutes; 30 cycles of 94°C for 30 seconds, 63°C for 90 seconds, and 72°C for 90 seconds, followed by a final extension at 72°C for 10 minutes. The PCR conditions used for qepA were as follows: initial denaturation at 96°C for 1 minute, followed by 30 cycles of amplification at 96°C for 1 minute, annealing
Table 1  Primers used in this study

<table>
<thead>
<tr>
<th>PMQR gene</th>
<th>Primers</th>
<th>Product size</th>
<th>Reference</th>
</tr>
</thead>
<tbody>
<tr>
<td>qnrA</td>
<td>5'-TCAGCAAGAGGGTTTCTCA-3' 5'-GGCAGCACTATACTCCCA-3'</td>
<td>516</td>
<td>16</td>
</tr>
<tr>
<td>qnrB</td>
<td>5'-GATGCGCAGGCGAGGAAGG-3' 5'-AGCATGCTTGGTATGTCG-3'</td>
<td>469</td>
<td>16</td>
</tr>
<tr>
<td>qnrS</td>
<td>5'-ACGACATTCGCTACAAGGCAA-3' 5'-TAATTGGAACCTCTAGGCC-3'</td>
<td>417</td>
<td>16</td>
</tr>
<tr>
<td>acc(6')-ib-cr</td>
<td>5'-TTGGAACGGCGAGGAM-3' 5'-ACACGGCTGGACCATA-3'</td>
<td>260</td>
<td>17</td>
</tr>
<tr>
<td>qoxAB</td>
<td>5'-CGCGACCGATAATATTGTCG-3' 5'-GGCGAGTTTTGATAGGCC-3'</td>
<td>313</td>
<td>18</td>
</tr>
<tr>
<td>qepA</td>
<td>5'-GGA GTCCAAGCGAGAGGATG-3' 5'-CTT CCT GCC GTA TCG TG-3'</td>
<td>218</td>
<td>18</td>
</tr>
</tbody>
</table>

Abbreviation: PMQR, plasmid-mediated quinolone resistance.

at 60°C for 1 minute, extension at 72°C for 1 minute, and the final extension step was at 72°C for 5 minutes. The primers used is given in Table 1.16-18 The PCR by-product was examined by electrophoresis in agarose gel containing ethidium bromide and visualized by gel documentation system.

DNA Sequencing
The PCR positive amplicons were sequenced at SciGenome Labs Pvt., India and analyzed with BLAST tools (www.ncbi.nlm.nih.gov). The assigned GenBank accession number for the submitted sequences are: (1) MH709266 (qnrA); (2) KY130487 (qnrB); (3) KY130488 (qnrS); (4) MH709269 (acc(6')-ib-cr), and (5) MN273774 (qoxAB).

Conjugation
Conjugation experiments were performed for all PMQR positive isolates. Escherichia coli J53 Azir strain was used as the recipient and PMQR positive isolates as donor. The donor and recipient cells (0.5 mL each) in logarithmic phase were added to 3 mL of LB broth and incubated at 37°C overnight. Transconjugants were selected by plating on MacConkey agar plates containing sodium azide (100 µg/mL) and ciprofloxacin (0.5 µg/mL).19 The transfer of PMQR genes in transconjugants was confirmed by PCR.

Incompatibility Grouping of Plasmid Encoding Resistance for PMQR Genes
Plasmid Inc group for the transconjugants was determined by PBRT. Five sets of multiplex PCR [(HI1, HI2, I1); [X, L/M, N]; [FIA, FIB, W]; [YP FIC]; [A/C, T, FIIS]] and three simplex PCR (FrepB, K/B, B/O) were performed.13 The primers employed is depicted in Table 2.13

Results
Antimicrobial Susceptibility Testing
All the study isolates were resistant to other fluoroquinolones—levofloxacin and ofloxacin. Resistance to other classes of antimicrobial agents was as follows: ceftazidime (86.9%), piperacillintazobactam (73.8%), imipenem (69.2%), and amikacin (63.8%). The MIC of ciprofloxacin ranged from 4 greater than or equal to 256 µg/mL. The MIC<sub>50</sub> and MIC<sub>90</sub> values were 64 and greater than or equal to 256 µg/mL, respectively.

Detection of Efflux Pump Activity
Among 130 isolates, twofold reduction was evident in 46 (35.8%) and fourfold or more reduction was observed in 37 (28.4%). Fourfold was evident in 11 (12.9%), 8-fold in 5 (5.8%), 16-fold in 7 (8.2%), 32-fold in 3 (3.5%), and 128 fold in 2 (2.3%) among P. aeruginosa. In A. baumannii, 4-fold reduction was observed in one (2.2%) isolate, 8-fold in three (6.6%), 16-fold in two (4.4%), and 64-fold in three (6.6%), respectively (Table 3).

Polymerase Chain Reaction
Among P. aeruginosa, qnr genes were detected in 36 (27.6%) isolates, of which 12 (14.1%) isolates harbored qnrB, 12 (14.1%) carried qnrS gene, and 9 (10.5%) isolates harbored both qnrB and qnrS genes. Among A. baumannii, qnrB was detected in two (4.4%) isolates and only one (2.2%) harbored both the qnrA and qnrS; 77.6% (66) of P. aeruginosa and 46.6% (21) of A. baumannii isolates harbored acc(6')-ib-cr gene; 3.5% (3) of P. aeruginosa and 2.2% (1) of A. baumannii isolates harbored qoxAB gene. qepA gene was not detected in any of the study isolates. The PMQR genes encountered is depicted in Table 4.

PMQR Gene Transfer and Distribution of Plasmid Replicons
In P. aeruginosa, 9.2% (12/130) were transferred successfully. All the 12 transconjugants were positive only for acc(6')-ib-cr gene in A. baumannii, none of them was transferable.

The plasmid incompatibility types of the transconjugants were recognized by PBRT. Of the 12 transconjugants, 7 (58.3%) belonged to IncFII type plasmid replicon, 4 (33.3%) were IncA/C, and 1 (8.3%) IncFIC type.
Table 2: Primers for PCR-based replicon typing

<table>
<thead>
<tr>
<th>Replicon type</th>
<th>Primer sequence (5′–3′)</th>
<th>Amplicon size (bp)</th>
</tr>
</thead>
<tbody>
<tr>
<td>HI1</td>
<td>F-GGAGCCGTGGATACCTCCTTCGACTA</td>
<td>471</td>
</tr>
<tr>
<td>HI2</td>
<td>F-TTCTTCGATTGTGATGTTACCTTCGACTA</td>
<td>644</td>
</tr>
<tr>
<td>I1</td>
<td>F-CGAAAGCCGACGCGGCAAA</td>
<td>139</td>
</tr>
<tr>
<td>X</td>
<td>F-AACTTACAGGCTTATTAAGTTCGTGAT</td>
<td>376</td>
</tr>
<tr>
<td>L/M</td>
<td>F-GGATGAAAGCTATCGATTCGAAAG</td>
<td>785</td>
</tr>
<tr>
<td>N</td>
<td>F-GTCGAGGTGTCGTTGACCGAGAG</td>
<td>559</td>
</tr>
<tr>
<td>FIA</td>
<td>F-CCTAGTCTGGTCTGAGAGGTG</td>
<td>462</td>
</tr>
<tr>
<td>FIB</td>
<td>F-GGAGTTTCGACACACATTGCTTATTG</td>
<td>308</td>
</tr>
<tr>
<td>W</td>
<td>F-CCTAAGAACAAGAAAGCCCCCG</td>
<td>242</td>
</tr>
<tr>
<td>Y</td>
<td>F-AATTCAGAACAAGACTCCTGCATG</td>
<td>765</td>
</tr>
<tr>
<td>P</td>
<td>F-CTATGGCCCTGAAACGCGGAGAAATCCGAC</td>
<td>534</td>
</tr>
<tr>
<td>FIC</td>
<td>F-GTGAAGTTCCGCATCAGACGAT</td>
<td>262</td>
</tr>
<tr>
<td>A/C</td>
<td>F-GAGAAAGCAGAAAGACTCTTGAGAAGACAAGAATGCCTTGCAAGAA</td>
<td>465</td>
</tr>
<tr>
<td>T</td>
<td>F-CTTGGAAGGCATTACCTCTGAAATGCCTTCGACTGCAAA</td>
<td>750</td>
</tr>
<tr>
<td>FIIa</td>
<td>F-CTCTTGCTAACGCTGGCAGAGGAG</td>
<td>270</td>
</tr>
<tr>
<td>FexpII</td>
<td>F-TGATCGTTAGGAAAATTTGGAAGAAGAATTCG</td>
<td>270</td>
</tr>
<tr>
<td>K/B</td>
<td>F-CCGCGCGAGGCGGACAG</td>
<td>160</td>
</tr>
<tr>
<td>B/O</td>
<td>F-CGGCGCCGAGGCGGACAGAAGAGAGA</td>
<td>159</td>
</tr>
</tbody>
</table>

Abbreviation: PCR, polymerase chain reaction.

Discussion

Fluoroquinolones are potent antibiotics active against a broad range of bacteria. The global increase in the prevalence of clinical strains with reduced susceptibility to fluoroquinolones constitutes a major concern. The emergence of fluoroquinolone resistance among P. aeruginosa and A. baumannii presents a serious challenge in clinical management of bacterial infections. In this study, antibiotic susceptibility testing revealed that all the test isolates were resistant to the other fluoroquinolones also: levofloxacin and ofloxacin. Resistance to other class of antibiotics was ceftazidime (86.9%), piperacillin/tazobactam (73.8%), imipenem (69.2%), and amikacin (63.8%). In this study, 32.9% (28) of P. aeruginosa and 37.7% (17) of A. baumannii isolates had an MIC of greater than or equal to 256 μg/mL to ciprofloxacin and similar observation has also been documented by Zaki et al.22

In this study, the MIC decrease factor value of fourfold and more reduction was evident in 28.4% (37/130) of isolates. In one isolate (4.5%) (P. aeruginosa), it resulted in loss of ciprofloxacin resistance thus reflecting a highly active efflux activity as evidenced by the decrease in MIC from 256 to 0.5 μg/mL on addition of CCCP inhibitor. Similar inhibition of efflux pump activity has been described previously. Researchers in the United States, Iran, and Bahrain have observed nonsignificant twofold reduction in MIC on addition of efflux pump inhibitor CCCP in P. aeruginosa. Helmy et al in their study reported significant efflux pump activity in A. baumannii and P. aeruginosa (46.1% and 41.1%).

Gomaa et al in Egypt recorded a high percentage of efflux pump-mediated resistance (77.8%) in A. baumannii. In contrast, in South Africa, CCCP inhibitors did not affect MIC in A. baumannii.

In this study, among qnr genes, 14.1% (12) were qnrB and 14.1% (12) were qnrS in P. aeruginosa which is a higher rate compared with a previous study from Egypt, which documented the presence of qnrB and qnrS genes in 1.8 and 2.7% of Pseudomonas spp. In contrast, El-Badawy et al and Rafig et al documented high prevalence of qnrS (79.5 and 24%) gene, respectively. qnrA and qnrB were not detected in any of their isolates of Pseudomonas spp. Similar to the earlier study, qnrA gene was not detected in any of the isolates of P. aeruginosa in the present study. In China, a single isolate of P. aeruginosa with qnrA has been observed. In many other studies, qnr determinants were not detectable in clinical isolates of P. aeruginosa.

In this study, among qnr genes, 14.1% (12) were qnrB and 14.1% (12) were qnrS in P. aeruginosa which is a higher rate compared with a previous study from Egypt, which documented the presence of qnrB and qnrS genes in 1.8 and 2.7% of Pseudomonas spp. In contrast, El-Badawy et al and Rafig et al documented high prevalence of qnrS (79.5 and 24%) gene, respectively. qnrA and qnrB were not detected in any of their isolates of Pseudomonas spp. Similar to the earlier study, qnrA gene was not detected in any of the isolates of P. aeruginosa in the present study. In China, a single isolate of P. aeruginosa with qnrA has been observed. In many other studies, qnr determinants were not detectable in clinical isolates of P. aeruginosa.

Yang et al examined the prevalence of qnr genes among 39 isolates of A. baumannii where 7.7% (3/39) isolates harbored qnrB and 2.6% (1/39) qnrS. Hamed et al also reported the presence of qnrS gene in one isolate of A. baumannii. Touati et al observed qnrA gene in only one isolate in their study. In the present study, qnrB was encountered in two isolates singly. One isolate harbored qnrB and qnrS, while another carried both qnrA and qnrS.

Table 3: Effect of CCCP on the ciprofloxacin MIC

<table>
<thead>
<tr>
<th>Organism (n = 130)</th>
<th>Fold reduction in MIC + CCCP (μg/mL)</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>0</td>
</tr>
<tr>
<td>P. aeruginosa (n = 85)</td>
<td>21</td>
</tr>
<tr>
<td>A. baumannii (n = 45)</td>
<td>26</td>
</tr>
</tbody>
</table>

Abbreviations: CCCP, carbonyl-cyanide m-chlorophenylhydrazone; MIC, minimal inhibitory concentration.
Table 4 Distribution of PMQR genes

<table>
<thead>
<tr>
<th>PMQR genes</th>
<th>Pseudomonas aeruginosa (n = 85)</th>
<th>Acinetobacter baumannii (n = 45)</th>
<th>Total prevalence (n = 130)</th>
</tr>
</thead>
<tbody>
<tr>
<td>aac(6′)-Ib-cr</td>
<td>31 (36.4%)</td>
<td>17 (37.7%)</td>
<td>48 (36.9%)</td>
</tr>
<tr>
<td>qoxAB</td>
<td>1 (1.1%)</td>
<td>1 (2.2%)</td>
<td>2 (1.5%)</td>
</tr>
<tr>
<td>qnrB + aac(6′)-Ib-cr</td>
<td>12 (14.1%)</td>
<td>2 (4.4%)</td>
<td>14 (10.7%)</td>
</tr>
<tr>
<td>qnrS + aac(6′)-Ib-cr</td>
<td>12 (14.1%)</td>
<td>0</td>
<td>12 (9.2%)</td>
</tr>
<tr>
<td>qnrA + qnrS + aac(6′)-Ib-cr</td>
<td>0</td>
<td>1 (2.2%)</td>
<td>1 (0.7%)</td>
</tr>
<tr>
<td>qnrB + qnrS + aac(6′)-Ib-cr</td>
<td>9 (10.5%)</td>
<td>1 (2.2%)</td>
<td>10 (7.6%)</td>
</tr>
<tr>
<td>qnrB + qnrS + oqxAB + aac(6′)-Ib-cr</td>
<td>1 (1.1%)</td>
<td>0</td>
<td>1 (0.7%)</td>
</tr>
<tr>
<td>oqxAB + aac(6′)-Ib-cr</td>
<td>1 (1.1%)</td>
<td>0</td>
<td>1 (0.7%)</td>
</tr>
<tr>
<td>Total</td>
<td>67 (78.8%)</td>
<td>22 (48.8%)</td>
<td>89 (68.5%)</td>
</tr>
</tbody>
</table>

Abbreviation: PMQR, plasmid-mediated quinolone resistance.

In Brazil, a low prevalence of aac(6′)-Ib-cr gene (2.6%) was found in *P. aeruginosa*. Studies from Turkey and Egypt reported a high prevalence of 56.4 and 79.5% in *P. aeruginosa*, respectively. This is similar to the findings of the present study (66.9%). In this study, only four (3%) isolates harbored oqxAB. Notably, qepA gene was not encountered. oqxAB and qepA genes were not identified in many other studies too.

Conjugation experiments demonstrated that in 14.1% (12/85) of *P. aeruginosa*, PMQR determinants were successfully transferred and all the transconjugants harbored the aac(6′)-Ib-cr gene. In *A. baumannii*, none of them was transferable. Jiang et al in their study documented that in 33.3% of nonfermenting gram negative bacteria (NFGNB), the transconjugants harbored the same PMQR determinants as their donors. In this study, more than one half of PMQR determinants, 59.2% were non-conjugative, and this suggests that these genes may be of chromosomal location. Among the PMQR genes, high incidence of aac(6′)-Ib-cr (66.9%) was encountered and when conjugated, the transferability rate was 100% for this gene. This emphasizes that aac(6′)-Ib-cr gene plays a major role in mediating fluoroquinolone resistance. In the present study, of the 12 transconjugants, 33.3% belonged to IncA/C type plasmid replicon. In Nigeria, IncFII plasmid harboring aac(6′)-Ib-cr gene has been described in *P. aeruginosa*. A recent study in Argentina has demonstrated plasmid IncR group in *P. aeruginosa*.

In this study, the prevalence rate of PMQR determinants is (68.5%), which is higher than the rates in China (1.7%), Egypt (4.5%), and Nigeria (61%). However, increasing rates of fluoroquinolone resistance have limited the treatment option. The approach of combined antibiotic therapies is an alternative to this phenomenon.

The ciprofloxacin resistance isolates which were negative for PMQR genes in our study probably harbor the chromosomal mutation genes (gyrA, gyrB, parC, and parE). These genes were not looked for in the present study.

Conclusion
Plasmid-mediated fluoroquinolone resistance is encountered in (78.8%) of *P. aeruginosa*, while in *A. baumannii*, it is present in a proportion of 48.8% of clinical isolates. Single clinical isolate can harbor multiple PMQR genes. Plasmid-mediated efflux fluoroquinolone resistance is responsible for a small proportion of resistance to fluoroquinolone in clinical isolates of *P. aeruginosa* and *A. baumannii*. Of the plasmid efflux pump genes, only oqxAB is present in 3% of isolates. It is reasonable to assume that of the plasmid-mediated resistance, aac(6′)-Ib-cr is primarily responsible for mediating a major proportion of resistance to fluoroquinolones. IncFII is the predominant plasmid type followed by IncAC and IncFIC type.

Funding
This study is funded by Department of Science & Technology (DST) under Women Scientists Scheme (WOS-A).

Conflict of Interest
None declared.

References
8. Rodríguez-Martínez JM, Díaz de Alba P, Briales A, et al. Contribution of OqxAB efflux pumps to quinolone resistance in extended-
Plasmid-Mediated Fluoroquinolone Resistance  
Geetha et al.

spectrum-β-lactamase-producing *Klebsiella pneumoniae*. J Anti-
microb Chemother 2013;68(01):68–73


22 Zaki MES, Abou ElKheir N, Mofreh M. Molecular study of quinolone resistance determining regions of gyrA and parC genes in clinical isolates of *Acinetobacter baumannii* resistant to fluoroquinolones. Open Microbiol J 2018;12:116–122


44 Elena A, Quinteros M, Di Conza J, Gutkind G, Cejas D, Radice MA. Full characterization of an IncR plasmid harboring *qnrS1*
