Use of a novel through-the-needle biopsy forceps in endoscopic ultrasound

Background and aims: Pancreatic cysts are becoming more common. Their differential diagnosis includes benign, premalignant, and malignant lesions. Distinguishing the type of cyst helps in the management decision making. We report on a novel tissue acquisition device for pancreatic cysts.

Methods: Data on two patients who underwent endoscopic ultrasound (EUS)–guided fine-needle aspiration with a new micro forceps device are presented.

Results: Two patients had large pancreatic cystic lesions in the pancreatic head. Linear EUS was performed, and tissue samples were obtained with the Moray micro forceps through a 19-gauge needle. In both patients, mucinous columnar epithelium lined the cystic walls. One patient underwent surgical resection, and the other elected surveillance. Examination of the surgical specimen from the first patient confirmed the cyst was a side-branch intraductal papillary mucinous neoplasm (IPMN), gastric type.

Conclusions: The Moray micro forceps is a new tool that can be used to help determine the nature of pancreatic cysts and aid in their risk stratification and management.
Case 2

A 78-year-old man with morbid obesity was admitted to the hospital for work-up of abdominal pain. Cross-sectional imaging demonstrated numerous pancreatic cystic lesions, the largest of which was a 5.0 × 4.7 × 4.4-cm septate cyst involving the uncinate process and head of the pancreas (Fig. 6).

EUS confirmed the presence of numerous pancreatic cystic lesions. The main pancreatic duct was normal in diameter. A 19-gauge needle (Cook Medical) was advanced into the largest cystic lesion, and fluid was aspirated. A Moray micro forceps (US Endoscopy) was repetitively advanced through the 19-gauge needle to obtain multiple specimens.

The microbiopsy specimens showed fragments of mucinous columnar epithelium and rare fragments of mucus, consistent with the clinical impression of a side-branch IPMN (Fig. 7).

The patient declined surgical intervention after discussion with the pancreatic surgeon because of his multiple medical co-morbidities.

Discussion

Pancreatic cysts are incidentally found in about 2.5% of patients undergoing cross-sectional imaging for reasons not related to the pancreas [2,3], and this finding is strongly correlated with advancing age. The prevalence increases in patients identified as being at high risk for pancreatic tumors based on their genetic and familial background [4]. In an autopsy study, 186 cystic lesions were found in 73 of 300 consecutive examinations (prevalence of 24.3%) [5]. An advantage of EUS over cross-sectional imaging is that tissue samples can be obtained when needed. Not all cysts require tissue sampling; the latest American Gastroenterological Association guidelines suggest that a sample be obtained

Video 1

when a cyst enlarges to more than 3 cm, when there is a solid component, or when the pancreatic duct is enlarged [1]. In a retrospective study of 35 surgically resected cysts larger than 3 cm, EUS-FNA had the highest specificity (88.9%) and magnetic resonance imaging had the highest sensitivity (100%) [6] for the detection of high grade epithelial atypia. Despite the high specificity of EUS-FNA in diagnosing mucinous cysts and high grade atypia, its sensitivity suffers because relatively acellular samples are obtained from pancreatic cysts.

Multiple techniques have been recommended for obtaining tissue samples from pancreatic cysts. Most authorities recommend aspirating fluid from the cyst for carcinoembryonic antigen (CEA) and amylase measurement and cytologic analysis. To increase the yield of EUS-FNA and improve its sensitivity, other technical methods to obtain cyst wall tissue have been studied. In cyst wall puncture, the cyst fluid is completely aspirated, the far wall of the cyst is punctured, and the needle is moved back and forth to obtain tissue. This method has been tested with standard FNA needles [7], showing a 29% improvement in yield ($P=0.001$) compared with CEA and cytology, and with FNA needles that have side fenestration [8, 9], showing a 65% sample adequacy for histopathologic diagnosis. The success rate for obtaining cyst fluid varied from 53.3% to 81%.

Another tool designed to improve the yield of EUS-FNA in pancreatic cystic lesions is the cytology brush (EchoBrush; Cook Medical). Studies of this brush show contradictory results. In one study that evaluated the cytology brush in 30 patients with pancreatic cystic lesions, the brushings improved the diagnostic yield of the specimens (73 vs. 36%, $P=0.08$); however, the technical failure rate was 27%, with 13.6% morbidity and 4.5% mortality [10]. Another study examining this tool showed a significant increase in sample adequacy (85.1% vs. 66.3%, $P<0.05$); however, intra cystic bleeding occurred in three patients, and an abscess developed in one [11].

A pilot study examined the use of a SpyGlass fiberoptic probe (Boston Scientific, Natick, Massachusetts, USA) and a 220-cm × 0.8-mm endoscopic retrograde cholangiopancreatography (ERCP) biopsy forceps (designed for Polyscope; Lumenis Surgical, Santa Clara, California, USA) through a 19-gauge EUS needle in two patients with pancreatic cystic lesions [12]. In both patients, the biopsy sample findings were consistent with mucinous cystic neoplasm. Severe pancreatitis developed in one of the patients 1 month later, but it was presumed that this was unlikely to be related to the biopsy.

**Conclusion**

We report use of the disposable Moray micro forceps, which is intended to sample tissue from lesions occurring within and outside the gastrointestinal tract (e.g., in the pancreas). The device length is 230 cm, with a jaw opening width of 4.3 mm (Fig. 2). The jaws are serrated and attached to a flexible 0.8-mm stainless steel spring sheath that easily passes through a 19-gauge FNA needle.

**Competing interests:** None
References