Spinal Dysraphisms: A New Anatomical–Clinicoradiological Classification

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Indian J Radiol Imaging 2021;31:809–829.

Abstract

Background  Spinal dysraphisms refer to the congenital abnormalities of the spine and spinal cord due to aberrations in the processes of gastrulation, primary neurulation, and secondary neurulation. Embryology of many complex spinal dysraphisms are yet poorly understood and there is no agreeable anatomical–clinicoradiological classification with inclusion of recently documented and complex spinal dysraphisms.

Aims and Objectives  The main objective of this study was to review the imaging features of spinal dysraphisms with a better understanding of embryological abnormalities and propose a new classification inclusive of all complex and unusual dysraphisms based on anatomical and clinicoradiological correlation.

Materials and Methods  This was a retrospective single institutional observational study of 391 cases of spinal dysraphism for 10 years in our institution. Of 391 cases included in the study, 204 were males and 187 were females. Also, 123 cases belonged to the 0–6 months age group, 38 cases belonged to the 7–12 months age group, 156 belonged to the 1–5 years age group, 39 cases belonged to the 6–10 years age group, and 35 cases belonged to 10–20 years age group.

Results  An anatomical–clinicoradiological analysis of cases yielded a high proportion of cases of spinal lipomas, including lipomyelocoeles and lipomyelomeningoceles (31.3%) and posterior myelomeningocele (14.2%). Anterior myelocoele (0.2%), sacral chordoma (0.2%), and intrasacral meningocele (0.2%) formed the least proportion of cases. A new classification was proposed based on the analysis of acquired data.

Conclusion  A structured approach in imaging spinal dysraphism is necessary for imaging evaluation in recent years. The proposed new classification based on clinicoradiological correlation and anatomic location is inclusive of unusual and complex dysraphisms.

Keywords

► spinal dysraphism
► embryology
► magnetic resonance imaging
► open spinal dysraphism
► closed spinal dysraphism

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**Introduction**

Spinal dysraphism is a group of diverse conditions that have variable imaging patterns. The purpose of this study was to analyze unusual and complex dysraphism and propose a new classification based on clinicoradiological correlation and anatomical location. There is a necessity for detailed embryological knowledge for a better understanding of imaging.

**Embryology**

Spinal dysraphism results from aberrations in the closely linked embryological processes occurring between the gestational weeks 2 and 6, namely, gastrulation (2–3 weeks), primary neurulation (3–4 weeks), and secondary neurulation (5–6 weeks). The level of coordination between these processes explains why the abnormal development of one structure will be associated with the maldevelopment of the other. The spinal cord up to the level of S2 (9/10th of the cord) is formed by primary neurulation and the spinal cord distal to it (terminal 1/10th) is formed by secondary neurulation.

**Gastrulation**

By the end of the second gestational week (post-fertilization), the inner cell mass of the blastocyst produces a bilaminar disc composed of two layers—epiblast facing the amniotic cavity and the hypoblast facing the yolk sac (Fig. 1). Gastrulation is the process of conversion of the bilaminar disc into a trilaminar disc initiated by a primitive streak. A narrow group of cells arises from the epiblast at the caudal end of the embryo forming the primitive streak (Fig. 2). A rapidly proliferating group of cells forms at one end of the primitive streak. This nodular proliferation surrounding a small primitive pit, known as Hensen’s node, defines the cephalic end of the primitive streak. The primitive node, which is a depression at the cranial end of the streak, contains cells that are important for establishing the embryonic axes. Epiblast cells migrate toward and through the streak and node, detach and form two new layers ventral to the remaining epiblast. The early cells passing through the streak displace the original hypoblast to form the endoderm, whereas cells migrating slightly later create a new middle layer, the mesoderm (Fig. 3). The mesodermal cells organize in the midline to form the notochordal process, which develops to become the notochord (Fig. 4). The sequential stages of notochordal development include one where the notochordal process fuses with the endoderm (interecalation), creating a communication of the central canal of the notochordal process with the yolk sac. This communication is called the primitive neureric canal/canal of Kovalevsky. Notochord initiates the process of neurulation by inducing the formation of the neural plate from overlying ectoderm cells. Thus, the neural plate is derived from the ectoderm and forms in the central

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**Fig. 1** Bilaminar Disc. (A) Bilaminar disc with the development of the amniotic cavity and the primary yolk sac followed by formation of the primitive streak in (B) and (C).

**Fig. 2** Formation of the primitive streak (A) showing the formation of the extra embryonic mesoderm and (B) and (C) showing the development of the prochordal plate from the hypoblast, cranially and the primitive streak from the cells of the epiblast, caudally. The formation of the Hensen’s node at one end of the primitive streak establishes it to be the cranial end of the primitive streak.
part of this upper layer. The remainder of the ectoderm surrounding the neural plate forms the epidermis. Thus, gastrulation and neurulation continue simultaneously in the human embryo.

### Primary Neurulation

The neural plate (Fig. 5) elevates to form the neural folds creating a neural groove in the midline. Pairs of somites, representing collections of underlying mesoderm that appear on either side of the neural groove, form the vertebrae. This mesoderm helps support the elevating neural folds. Lateral borders of the neural plate elevate into neural folds, which later fuse in the midline to form the neural tube. Elevation occurs by the proliferation of underlying mesoderm and hyaluronic acid. After elevation, the neural folds furrow to form a neural groove, the edges of which eventually close to form a cord (Fig. 6). This occurs through the formation of hinge points using microfilaments and microtubules. Neural tube closure first occurs near the junction of the hindbrain and spinal cord at the level of the fifth somite. It then proceeds in a zipper-like fashion cranially and caudally (Fig. 6E). The open regions of the neural tube are called the anterior (cranial) and posterior (caudal) neuropores. Cranial neuropore is closed on day 25, whereas the caudal neuropore is closed on day 28. On neural tube closure, overlying non-neural epidermal cells form the ectodermal layer of the skin, whereas the neuroectoderm cells reorganize to form the roof of the neural tube. Primary neurulation is responsible for establishing brain and spinal cord regions down to the lowest sacral levels (probably up
to the level of S2). Neural crest cells on either side form accessory neural tissue\(^4\) (**Fig. 7**).

### Secondary Neurulation

This process helps in the development of the cord caudal to the S2 level. A caudal cell mass of undifferentiated, totipotent mesodermal cells initially appears at the caudal end at the end of primary neurulation; it subsequently undergoes vacuolization, canalization (**Fig. 8C**) and finally become continuous with the cranial part of the tube initially formed by primary neurulation.\(^5\) Part of the caudal cell mass undergoes retrogressive differentiation to form filum terminale, terminal ventricle, the tip of conus medullaris, the sacrum, and coccyx.\(^5\) By the third month of gestation, the spinal cord extends the entire length of the developing spinal column. There is an apparent ascent of the cord causing the conus medullaris to recede cranially due to rapid growth of the vertebral column. A ventriculus terminalis of the central canal is found within the lumbar enlargement during secondary neurulation.

The various anomalies occurring due to aberrations in each stage of embryological development have been summarized in **Table 1**.

### Materials and Methods

#### Study Design

This was a retrospective single institutional observation study.
Table 1: A review of embryological events and their abnormalities

<table>
<thead>
<tr>
<th>Time period</th>
<th>Embryological stages</th>
<th>Probable embryological abnormalities</th>
<th>Spinal dysraphism</th>
</tr>
</thead>
</table>
| Day 12–20         | **Gastrulation:** The paired mesodermal primordia formed during gastrulation subsequently integrate along the midline to form the notochordal process. The notochord helps in the beginning of neurulation. Later, the notochord persists as the nucleus pulposus of the intervertebral discs | 1. **Disorders of notochord formation**  
A) Caudal notochord and caudal cell mass formation affected resulting in a high position and abnormal termination of conus medullaris.  
B) Intermediate notochord dysgenesis  
Error in positional imprinting of migration of chorda: mesodermal cells trough the primitive pit and into the ectoderm-endoderm interface could activate the abnormal apoptosis resulting in dysgenetic segment | Caudal regression syndrome  
(Type I)  
Segmental Spinal Dysgenesis |
|                   | **The transient neuenteric canal/canal of Kovalevsky is a transient communication of the amnion through the notochordal canal to the yolk sac during notochordal formation at days 16–17** | 2. **Disorders of notochord integration**  
A) Persistent connection between ectoderm and endoderm results in splitting of the spinal cord. Abnormalities during these stages produce the spectrum of dorsalmostic spinal anomalies. Currently, the most widely accepted theory suggests a primary notochord defect, resulting in secondary changes to the paraxial mesoderm, which is responsible for the formation of the spinal column, giving rise to a medial intersosseous space. Through this space, the endoderm and the underlying primitive intestine herniate, adhere to the dorsal ectoderm, and eventually rupture out.  
B) **Splitting of the spinal cord into two hemicords**  
In type I diastematomyelia, the intervening primitive streak develops into a bony bar or cartilage, resulting in two hemicords in different dural sacs separated by an osseocartilaginous septum. Each of them includes a central canal, one dorsal, and one ventral root. In type II diastematomyelia, the primitive streak is resorbed or forms a fibrous septum with the hemicords lying within the same dural sac | Dorsal enteric fistula  
Neuroenteric cysts  
Dorsal enteric sinus  
Dorsal enteric diverticulum  
Diastematomyelia |
| Weeks 3 and 4     | **Primary neurulation** The spinal cord up to the level of S2 is formed by primary neurulation. The process of neural tube formation (neurulation) occurs simultaneously with the separation of the neuroectoderm from the surface ectoderm, which is called disjunction | 3. **Disorders of notochord duplication**  
Duplication of notochord results in the extreme end of split cord malformations (split notochord syndrome). | Vertebral duplication  
- Vertebral duplication implies the presence of double cords. Unlike type 1 Diastematomyelia, Each cord had its own spinal canal within its own set of vertebrae |
| (Day 17–27)       |                                                                                       | 1. **Premature disjunction**  
Inclusion of mesoderm in the neural tube secondary to premature separation of the ectoderm and neural placode, with mesoderm preventing tube closure results in this spectrum. | Lipomyelomeningocele  
Lipomyelocele  
Type 1 Spinal Lipoma  
Intramedullary Lipoma |

(Continued)
<table>
<thead>
<tr>
<th>Time period</th>
<th>Embryological stages</th>
<th>Probable embryological abnormalities</th>
<th>Spinal dysraphism</th>
</tr>
</thead>
<tbody>
<tr>
<td>Week 5 and 6 (Day 26)</td>
<td>Secondary neurulation and retrogressive differentiation</td>
<td><strong>Caudal cell mass formation is affected</strong>&lt;br&gt;Interim focal expansion of the central canal known as ventricle terminalis is found within the lumbar enlargement during secondary neurulation.&lt;br&gt;Syrinx herniating through the posterior vertebral defect into a posterior meningocele&lt;br&gt;Intrathecal cystic lesion arising as a protrusion of the arachnoid through a congenitally weak area in the dura. Mostly these are extradural in location, causing compression of the bony sacral canal, associated with Tarlov cysts and caudal regression syndrome&lt;br&gt;Defective apoptosis during late secondary neurulation results in a persistent medullary cord&lt;br&gt;Abnormal junctional neurulation between primary and secondary neurulation&lt;br&gt;Abnormal mesenchymal differentiation/inclusion</td>
<td>Caudal regression syndrome&lt;br&gt;Ventriculus terminalis&lt;br&gt;Posterior myelocystocele&lt;br&gt;Intrasacral meningocele&lt;br&gt;Retained Medullary Cord&lt;br&gt;Type 2 Spinal Lipomas&lt;br&gt;Type 3, Type 4 Spinal Lipomas, Intramedullary Lipomas</td>
</tr>
</tbody>
</table>

| Weeks 3–5 | The development of vertebrae begins at the time of development of notochord. The mesoderm surrounding the notochord separates into paraxial, lateral, and intermediate mesoderm. The paraxial mesoderm develops into 42 pairs of somites. Each somite then differentiates into a sclerotome and a dermomyotome. Sclerotome forms a skeleton of the vertebral column while dermomyotome forms muscle and dermis of the skin | **Mesodermal maldevelopment around the neural tube and notochord**<br>-failure of development of mesoderm around the neural tube and notochord, results in anterior/lateral vertebral body defects along with anterior dysplasia of meninges resulting in these spectra of anomalies.<br>Another possible theory is midline persistent neurenteric canal (failure of closure), through which there is herniation of cord structures<sup>11</sup> | Anterior myelomeningocele<br>Anterior meningocoele<br>Anterior myelocele<br>Anterior myelocystocele<br>Anterior lipomyelocele<br>Anterior lipomyelomeningocele |
Study Population
A total of 391 children, whose imaging and clinical features were consistent with spinal dysraphism from January 2011 to January 2020 in Stanley Medical College, Chennai, were included in the study. Postoperative spinal dysraphism is excluded from the study. Adults (>20 years) were not included in the study. Cutaneous and subcutaneous hemangiomas, lipomas, other fibrovascular lesions, and acrochordons were included under cutaneous stigmata.

Data Acquisition
Magnetic resonance imaging (MRI) spine was performed as per the institutional protocol through (MRI, MR Magnetom aera, 1.5 T, 48 channel Siemens, Erlangen, Germany). The MRI spine protocol was as follows:

1) MR myelogram in sagittal and coronal planes
2) T1, T2, short T1 inversion recovery images, T1 fat-suppression in sagittal sections with the field of view including the entire spine with a slice thickness of 3 mm and slice gap of 0.3 mm
3) T1- and T2-weighted images in the axial section of the spine. In all cases, in addition to the whole spine sagittal images, focused axial sections of malformed cord segment, with the same 3 mm—slice thickness and 0.3 mm gap, were imaged for clear visualization of the spinal cord.

The MR images were systematically analyzed by two experienced radiologists. Clinical manifestations in other systems were not analyzed. Association with Arnold Chiari malformations and other vertebral, rib anomalies were not analyzed.

Demographic Data
Out of 391 cases included in the study, 204 were males and 187 were females.

In total, 123 cases belonged to the 0–6 months age group, 38 cases belonged to the 7–12 months age group, 156 belonged to the 1–5 years age group, 39 cases belonged to the 6–10 years age group, and 35 cases belonged to 10–20 years age group.

Results
(►Table 2)

Discussion
Terminology and Classification
Spina bifida: The term spina bifida classically refers to the defective fusion of posterior spinal bony elements. The spectrum of cases in the study included bony defects in the anterior arch as well.

Neural placode: Segment of non-neurulated embryonic neural tissue, which is arrested at the neural plate stage.

Open and closed spinal dysraphism: The neural placode is exposed through a skin defect in the midline in an open

Table 2 Proportion of spinal dysraphisms in the study*

<table>
<thead>
<tr>
<th>Condition</th>
<th>Number of cases</th>
<th>Proportion</th>
</tr>
</thead>
<tbody>
<tr>
<td>Posterior myelomeningocele</td>
<td>64</td>
<td>14.2%</td>
</tr>
<tr>
<td>Posterior myelocoele</td>
<td>7</td>
<td>1.5%</td>
</tr>
<tr>
<td>Anterior myelocoele</td>
<td>1</td>
<td>0.2%</td>
</tr>
<tr>
<td>Anterior myelomeningocele</td>
<td>3</td>
<td>0.66%</td>
</tr>
<tr>
<td>Posterior Hemimyelomeningocele</td>
<td>2</td>
<td>0.4%</td>
</tr>
<tr>
<td>Posterior Lipomyelomeningocele</td>
<td>15</td>
<td>3.33%</td>
</tr>
<tr>
<td>Posterior lipomyelocele</td>
<td>25</td>
<td>5.5%</td>
</tr>
<tr>
<td>Anterior lipomyelocele</td>
<td>3</td>
<td>0.66%</td>
</tr>
<tr>
<td>Anterior myelocystocele</td>
<td>7</td>
<td>1.55%</td>
</tr>
<tr>
<td>Posterior myelocystocele</td>
<td>10</td>
<td>2.22%</td>
</tr>
<tr>
<td>Posterior meningocoele</td>
<td>20</td>
<td>4.44%</td>
</tr>
<tr>
<td>Type 1, Type 2, Type 3 spinal lipomas</td>
<td>56</td>
<td>12.4%</td>
</tr>
<tr>
<td>Type 4 Spinal Lipoma</td>
<td>19</td>
<td>4.2%</td>
</tr>
<tr>
<td>Intramedullary lipoma</td>
<td>3</td>
<td>0.56%</td>
</tr>
<tr>
<td>Low lying cord</td>
<td>20</td>
<td>4.44%</td>
</tr>
<tr>
<td>Limited dorsal myeloschisis (saccular and Flat)</td>
<td>24</td>
<td>5.3%</td>
</tr>
<tr>
<td>Retained medullary cord</td>
<td>15</td>
<td>3.3%</td>
</tr>
<tr>
<td>Persistent terminal ventricle</td>
<td>6</td>
<td>1.13%</td>
</tr>
<tr>
<td>Dorsal dermal sinus</td>
<td>23</td>
<td>4.3%</td>
</tr>
<tr>
<td>Dorsal enteric fistula</td>
<td>2</td>
<td>0.4%</td>
</tr>
<tr>
<td>Neurenteric cyst</td>
<td>8</td>
<td>1.7%</td>
</tr>
<tr>
<td>Dermoid/epidermoid cysts</td>
<td>6</td>
<td>1.3%</td>
</tr>
<tr>
<td>Cervical diastematomyelia</td>
<td>3</td>
<td>0.66%</td>
</tr>
<tr>
<td>Thoracic diastematomyelia</td>
<td>17</td>
<td>3.7%</td>
</tr>
<tr>
<td>Lumbar diastematomyelia</td>
<td>27</td>
<td>6.0%</td>
</tr>
<tr>
<td>Type 1</td>
<td>4</td>
<td>0.8%</td>
</tr>
<tr>
<td>Type 2</td>
<td>14</td>
<td>3.1%</td>
</tr>
<tr>
<td>Complex</td>
<td>3</td>
<td>0.6%</td>
</tr>
<tr>
<td>Partial</td>
<td>6</td>
<td>1.3%</td>
</tr>
<tr>
<td>Caudal regression syndrome</td>
<td>13</td>
<td>2.9%</td>
</tr>
<tr>
<td>Segmental spinal dysgenesis</td>
<td>11</td>
<td>2.4%</td>
</tr>
<tr>
<td>Vertebral duplication</td>
<td>3</td>
<td>0.66%</td>
</tr>
<tr>
<td>Intrasacral meningocoele</td>
<td>1</td>
<td>0.2%</td>
</tr>
<tr>
<td>Sacrococcygeal teratoma</td>
<td>9</td>
<td>2.0%</td>
</tr>
<tr>
<td>Sacral chordoma</td>
<td>1</td>
<td>0.2%</td>
</tr>
</tbody>
</table>

*Of the total 391 cases analyzed, two or more simultaneously occurring aberrations were seen to occur in 71 cases bringing the total number of pathological entities that we could analyze, to 450 which could then be analyzed and classified under the proposed new classification (►Table 5).
spinal dysraphism. There is no exposed neural tissue in the case of a closed spinal dysraphism.

**Posterior spinal dysraphism:** These are defects of the laminae and the spinous process of the vertebrae and/or the intervening mesenchymal elements.

**Anterior spinal dysraphism:** These are defects in the vertebral body/pedicles/neural foramina of the vertebrae and/or the intervening mesenchymal elements.

Dysraphism is confined to the spinal canal with or without a dural defect.

**Primary defects involving multiple (more than 2) anatomical compartments:** (prevertebral mesenchyme, anterior or spinal bony elements, spinal canal, posterior spinal bony elements, postvertebral mesenchyme, and other organ systems)

**Nondysraphic masses of the spinal region:** Tumors with a close association with the notochord and the neural tube with neurulation remaining unaffected.

### Cutaneous Stigmata of Spinal Dysraphism: Skin Predicts the Spine

Larger defects are usually obvious, whereas small or occult malformations causing tethering of the cord may be asymptomatic at birth. Also, 69.1% of patients in our study had a cutaneous marker of spinal dysraphism, most commonly in the lumbosacral region. Hence, the first steps in diagnosis are these clinical cues such as the presence or absence of an exposed neural placode, the presence of a subcutaneous mass, or a dermal sinus in case of closed dysraphisms. A combination of cutaneous markers is possible (Table 3). A low-lying (<2.5 cm from anal verge) sacral dimple may not always be associated with underlying spinal dysraphism (Table 4).

#### Table 3 Skin Stigmata and commonly associated conditions

<table>
<thead>
<tr>
<th>Cutaneous marker</th>
<th>Associated condition</th>
</tr>
</thead>
<tbody>
<tr>
<td>Hypertrichosis (Faun tail)</td>
<td>Split cord malformation</td>
</tr>
<tr>
<td>Sacral dimple</td>
<td>Dorsal dermal sinus</td>
</tr>
<tr>
<td>Subcutaneous lipoma</td>
<td>Lipomyelomeningocele</td>
</tr>
<tr>
<td>Absent or asymmetric gluteal cleft</td>
<td>Sacral agenesis</td>
</tr>
<tr>
<td>Others</td>
<td>Non-specific occult dysraphism</td>
</tr>
</tbody>
</table>

#### Table 4 Skin stigmata of occult spinal dysraphism

<table>
<thead>
<tr>
<th>High probability of underlying dysraphism</th>
<th>Low probability</th>
</tr>
</thead>
<tbody>
<tr>
<td>Hypertrichosis (Faun tail nevus)</td>
<td>Capillary malformation (port-wine stain)</td>
</tr>
<tr>
<td>Dimples (atypical large dimples &gt; 2.5 cm from the anal verge)</td>
<td>Telangiectasia</td>
</tr>
<tr>
<td>Acrochordons/pseudotails/true tails</td>
<td>Hyperpigmentation</td>
</tr>
<tr>
<td>Subcutaneous lipomas</td>
<td>Melanocytic nevi</td>
</tr>
<tr>
<td>Hemangioma</td>
<td>Low-lying sacral dimple (&lt;2.5 cm from the anal verge)</td>
</tr>
<tr>
<td>Aplasia cutis or scar</td>
<td></td>
</tr>
<tr>
<td>Dermoid cyst or sinus</td>
<td></td>
</tr>
</tbody>
</table>

An embryological knowledge of these complex interlinked processes, although necessary, is often confusing while arriving at a diagnosis. Hence, the need for a simpler and more practical anatomical-neuroradiological classification (Table 5).

#### Table 5 Anatomical–neuroradiological classification of spinal dysraphism (Stanley Hospital classification)

1) Posterior spinal dysraphism
   - Posterior: Open spinal dysraphism:
     - Myelomeningocele
     - Myelocoele
     - Hemi myelomeningocele
     - Hemi myelocoele
   - Posterior: Closed spinal dysraphism
     - With subcutaneous mass:
       - Meningocele
       - Sacral limited dorsal myeloschisis
       - Myelocystocele
   - Posterior lipomyelomeningocele (spinal lipoma Type IB, IIB, IIB)
   - Posterior lipomyelomingocele (spinal lipoma type VB)

2) Anterior spinal dysraphism:
   - Anterior myelomeningocele
   - Anterior meningocoele
   - Anterior myelocoele
   - Anterior myelocystocele
   - Anterior lipomyelocoele (spinal lipoma type IC, IIC, IIC)
   - Anterior lipomyelomingocele (spinal lipoma type VA)
   - Intracranal meningocele
   - Prevertebral neurenteric cysts

3) Dysraphism confined to the spinal canal:
   - Intramedullary spinal lipoma
   - Intradural lipomas
     - Type Ia,
     - Type Ib, IId
     - Type IIIa
     - Type IVa–IVA-apical, IVb: mid filar and IV c: filar tip Lipomas
     - Tight filum terminale (type IVD)
   - Intracranal meningocele
   - Persistent terminal ventricle
   - Retained medullary cord
   - Split cord malformations
   - Types 1 and 2 diastematomyelia
   - Composite type diastematomyelia
   - Partial type diastematomyelia
   - Neuroenteric cysts
     - (Epi)dermoid/dermoid
   - Segmental spinal dysgenesis: type 1

4) Dysraphism involving multiple anatomical compartments:
   - Dorsal enteric fistula
   - Vertebral duplication
   - Caudal regression syndrome
   - Segmental spinal dysgenesis: type 2
   - OEIS, VACTERL, Currarino’s triad associations

5) Nondysraphic masses of the spinal region:
   - Sacrococcygeal teratoma
   - Sacral chordoma
A Review of the Anomalies and Associated Findings

Myelomeningocele and Myelocele: These are classified under open dysraphisms. The most common type of open spinal dysraphism is myelomeningocele (Fig. 9). The neural placode protrudes above the skin surface in case of a myelomeningocele due to cystic dilatation of subarachnoid space whereas a myelocele does not protrude above the skin surface.

Most are located at the lumbosacral level, and the placode is terminal (below the S2 level). A midline skin defect causes the external surface of the placode to be directly visible on inspection.

The nerve roots originate from this surface and course obliquely through the subarachnoid space to reach their corresponding neural foramina. Because both are exposed to the skin surface, ulceration and risk of infection are more, which increases the mortality rate. Hence, surgery is needed soon after birth or even before birth with exit. Preoperative imaging with MRI helps in better anatomic characterization and assessing the associated abnormalities Chiari malformations, syrinx, hydrocephalus, and split cord malformations.

In the proposed classification, meningoceles and myelomeningoceles have been further classified into those extending anteriorly and those extending posteriorly.

Myelomeningocele is almost always associated with Chiari 2 malformation.

McLone and Knepper proposed a theory to explain this consistent association. CSF leaks freely through the spinal defect into the amniotic sac because the neural tube remains non-neurulated. The resulting chronic CSF hypotension causes failure of expansion of the rhombencephalic vesicle, causing a lack of induction of the perineural mesenchyme of the posterior cranial fossa. Both the cerebellum and brainstem eventually are forced to develop within a smaller than normal posterior fossa and consequently herniate through the tentorial groove and the foramen magnum.

Hemimyelocele and Hemimyelomeningocele

These are extremely rare. These occur when a myelomeningocele or myelocele is associated with diastematomyelia and one hemicord fails to neurulate.

Meningocele

Meningoceles are herniation of CSF-filled sacs lined by dura and arachnoid. When it occurs through a posterior bony spina bifida, they are called posterior meningoceles. The most common are lumbar or sacral posterior meningocele (Fig. 10); therefore, thoracic and even cervical meningoceles may be found. The spinal cord is not contained within the meningocele although it may be tethered to its neck. Most of these are extradural in location, causing compression of the bony sacral canal. A few reported meningoceles may be a dome stalk variety of saccular limited dorsal myeloschisis.

Anterior Neural Tube Defects

Myelocystocele, myelomeningoceles, myelocystoceles, and lipomyelocoele usually occur posteriorly. When these lesions occur anteriorly or laterally with defects in the vertebral body, pedicles or neural foraminae, they are collectively called anterior neural tube defects. Golden and Chernoff hypothesized multiple sites of anterior neural tube closure and that anterior neural tube defects occur either due to failure of closure to occur or from the failure of two closures to meet. Another possibility would be due to CSF pulsation resulting in herniation through anteriorly located cleft in the vertebra.

Anterior Myelocystocele

Herniation of dilated cord with syrinx (hydro melic cord) through anterior vertebral defect is called anterior myelocystocele (Fig. 11). Meningocele requires a simple surgical repair, whereas myelocystocele requires untethering of cord,
decompression of syrinx along with excision, and repair of myelocystocele.\textsuperscript{25}

**Anterior Myelomeningocele**

Posterior myelomeningoceles are more common in the lumbo-sacral region (80\%). Anterior myelomeningoceles (\textsuperscript{\textasteriskcentered}Fig. 12) are rarely seen in the sacral region and very rarely seen in the thoracic/cervical region. The embryology behind this is an incomplete development of mesoderm around the neural tube, which results in the anterior defect through which there is herniation of meninges and other contents.

Multiple lateral thoracic meningoceles are usually associated with neurofibromatosis (85\%). Patients may present with sudden breathlessness secondary to rupture of meningocele resulting in hydrothorax.

**Myelocystocele**

Myelocystoceles occur due to herniation of dilated central canal forming terminal syringohydromyelia (syringocele) through a posterior (\textsuperscript{\textasteriskcentered}Fig. 13) or anterior vertebral defect into an expanded CSF-filled dural sheath (meningocele). It usually results from defective secondary neurulation that affects the CSF flow dynamics. The inner terminal syrinx communicates with the central canal of the spinal cord and the outer meningocele is continuous with the spinal subarachnoid space. The syringocele and meningocele usually do not communicate with each other.\textsuperscript{14} Although classified as terminal and non-terminal myelocystoceles earlier, many of the “non terminal” myelocystoceles were found to be the saccular type of limited dorsal myeloschisis. Thus, it is recommended to stick to the term “myelocystocele (anterior/posterior).”

**Spinal Lipomas**

The embryonic genesis of spinal lipoma cases is marked by a premature disjunction between the neural and cutaneous ectoderm during the process of primary neurulation. Embryologists often consider closed caudal spinal lesions to be the result of failed secondary neurulation. But still, some confusion in terms of the embryogenesis of spinal lipomas remains.

**Spinal lipomas:** Embryologically, Type I is defined as pure primary neurulation failure; Type II ranges from primary to secondary neurulation failure; Type III consists of secondary neurulation failure (early phase); and Type IV is defined as secondary neurulation failure (late phase).\textsuperscript{12} This classification of spinal lipomas based on the embryonic stage has the potential for clinical use and agrees well with
both clinical and surgical findings. However, we have now added subtypes to include the extension of the lipoma posteriorly (IB/IIIB/IIIB) and anteriorly (IC/IIC/IIIC) respectively as lipomyelolceles to add on to the previous classification. The rare intramedullary lipomas could be classified as type VI Spinal lipomas (Table 6).

**Type I: Primary Neurulation Failure**

Type I spinal lipoma is one in which the lipoma–cord interface occurs on the dorsal surface of the spinal cord (Type IA), while the conus medullaris is located distally or ventrally with respect to the caudal end of the lipoma. It may be found extending dorsally (Type IB) or ventrally (Type IC) through a spinal defect as lipomyelolecele and be associated with a tethered cord. They are usually seen in the lumbosacral region. Large lipomas may cause cord displacement. On MRI, lipomas follow the signal intensity of subcutaneous fat on all sequences (Fig. 14).

**Type II: Failed Neurulation Between the Primary and Secondary Stages**

Type II spinal lipomas include the transitional and chaotic lipomas that have a clearly discernible demarcation with the cord surface cranially but caudally either involve the conus medullaris, entangle the nerve roots of the cauda equina, or along with unpredictable involvement of the ventral fat as in the case of chaotic lipomas (Type IID). They may extend dorsally (Type IIB) or ventrally as lipomyeloleceles too (Type IIC) (Fig. 14).

The Type I spinal lipomas are formed purely by failed primary neurulation; the conus medullaris is left unaffected. The tapered shape of the end of the spinal cord suggests the presence of the conus medullaris. In contrast, a sharp tapered end to the spinal cord is not clearly discernible in Type II spinal lipomas. Dural lipomatosis occurring due to factors such as steroid abuse is a differential to be considered in the case of diffuse fatty proliferation.

**Type III: Early Phase Secondary Neurulation Failure**

This type of spinal lipoma involves the conus medullaris and is always distal to the origin of the last branch of the spinal nerves on the cord. The caudal end of the conus is unformed and directly connects to the spinal lipoma. The lipoma extends in the caudal direction and usually remains intradural (Type IIIa). Earlier classifications propounded that in contrast to Type II, in Type III spinal lipomas, there is never a pathological spina bifida or dural defect due to the lipoma. A few of our cases showed extension of these caudal lipomas posteriorly and anteriorly as lipomyeloceles and hence, we have added types IIIb and IIIc, respectively (Fig. 15), to indicate this extension.

**Type IV: Late Phase Secondary Neurulation Failure**

In this type, the spinal lipoma is located in the filum terminale. The caudal end of the conus medullaris is clearly discernible (Fig. 15). On imaging, filar lipoma appears hyperintense on T1- and T2-weighted images.

Lipomas of the filum terminale could be further classified (Fig. 15) as Type IVA: abutting the apex of the conus medullaris; apical lipoma that requires meticulous dissection during surgery; Type IVB: mid filar lipomas with normal-appearing/thickened filum terminale cranially and caudally; and Type IVc: filar tip lipomas that were always associated with a low-lying cord and tethered cord syndromes among the cases analyzed.

**Tight Filum Terminale (Type IVD Lipoma)**

It is characterized by shortening and hypertrophy of filum terminale that cause tethering of cord and impairs the ascent of conus medullaris. Embryologically, the defect lies in retrogressive differentiation during secondary neurulation.

On imaging, it is characterized by a thick filum terminale (Fig. 16) due to fibrous or fatty infiltration (thickness
measuring >2 mm at L5–S1) and a low-lying conus medullaris.

Also, 1.5% to 5% of the normal adult population may show fat within filum terminale on MRI and hence the finding is considered a normal variant unless it is associated with tethered cord syndrome.\textsuperscript{15,16}

\section*{Lipomyelocele and Lipomyelomeningocele}

Lipomyelocoles and lipomyelomeningoceles are characterized by a subcutaneous fatty mass located above the intergluteal crease and usually extending asymmetrically into one buttock.

In a lipomyelocele (lipomyeloschisis), the placode–lipoma interface lies within the spinal canal (Fig. 17) and may extend over several vertebral levels. There is continuity of the intraspinal lipoma with the subcutaneous fat through a posterior spina bifida.\textsuperscript{17} Any of the types I, II, and III spinal lipomas may present as a lipomyelocele.

In lipomyelomeningoceles, the placode–lipoma interface lies outside the spinal canal because of expansion of the underlying subarachnoid spaces, resulting in a posterior meningocele. Anterior and posterior lipomyelomeningoceles can be classified as Va and Vb, respectively, (Fig. 19).

\subsection*{Type VI: Intramedullary lipoma (Fig. 18)}

integumentation of menenchymal elements within the developing spinal cord results in the unusual intramedullary lipomas.\textsuperscript{18}

\subsection*{Retained Medullary Cord}

Defective apoptosis during late secondary neurulation results in a persistent medullary cord (Fig. 21). An absent filum terminale is characteristic of most RMCs.\textsuperscript{19} They appear like a robust cord-like structure attached from the conus to the cul-de-sac. An altered shape of the conus is visible.

They are often misread as a low-lying conus or a tight filum terminale (Table 7). Although MRI signal intensity characteristics are the same as a low-lying conus making preoperative diagnosis difficult, intraoperative
neuromonitoring helps distinguish between the two as an RMC is non-functional with no electrical activity. The symptoms are essentially due to associated cord tethering. Few variants (►Fig. 21) may show an hourglass shape with the non-functional medullary cord located caudal to the conus with an intervening short filum terminale. A few cases of association with a subcutaneous meningocele or a dermal sinus have been known.28 A few other cases showing an RMC associated with a lipoma are known.

**Filar cysts:** The etiology of filar cysts is unclear; however, it is postulated that it may be a pseudocyst-like structure formed by arachnoid reflections. Another theory says that could be a true cyst that is a different remnant of the terminal ventricle. Filar cysts are thought to be of no clinical significance and are a normal variant. They are located within the filum, in the midline, just distal to the conus medullaris. The difference in location helps to distinguish between the persistent terminal ventricle. Sometimes, these cysts are not well demonstrated on MRI.

**Filar cysts:**

Table 6 Spinal lipomas (►Fig. 20)

<table>
<thead>
<tr>
<th>Type</th>
<th>Description</th>
</tr>
</thead>
<tbody>
<tr>
<td>Type I</td>
<td>Intradural lipomas above the level of Conus (Lipoma: placode interface within the dura)</td>
</tr>
<tr>
<td>IA:</td>
<td>Those lipomas that are dorsal to the cord or extend cranially/caudally/around the spinal cord; entirely confined to the intradural space</td>
</tr>
<tr>
<td>IB:</td>
<td>Dorsal extension as a lipomyelocele</td>
</tr>
<tr>
<td>IC:</td>
<td>Ventral extension as a lipomyelocele</td>
</tr>
<tr>
<td>Type II</td>
<td>Transitional and chaotic lipomas</td>
</tr>
<tr>
<td>IIA:</td>
<td>Transitional lipoma above the last dorsal root entry zone of the conus</td>
</tr>
<tr>
<td>IIB:</td>
<td>Dorsal extension as a lipomyelocele</td>
</tr>
<tr>
<td>IIC:</td>
<td>Ventral extension as a lipomyelocele</td>
</tr>
<tr>
<td>Type III</td>
<td>Conal lipomas arising from the level of conus medullaris below the level of last dorsal root entry zone</td>
</tr>
<tr>
<td>III A:</td>
<td>Confined to intradural space</td>
</tr>
<tr>
<td>III B:</td>
<td>Dorsal extension as a lipomyelocele</td>
</tr>
<tr>
<td>III C:</td>
<td>Ventral extension as a lipomyelocele</td>
</tr>
<tr>
<td>Type IV</td>
<td>Filar lipomas: Lipomas of the filum terminale</td>
</tr>
<tr>
<td>IV A:</td>
<td>Apical lipoma</td>
</tr>
<tr>
<td>IV B:</td>
<td>Mid filar lipoma</td>
</tr>
<tr>
<td>IV C:</td>
<td>Filar tip lipoma</td>
</tr>
<tr>
<td>IV D:</td>
<td>Tight filum terminale</td>
</tr>
<tr>
<td>Type V</td>
<td>Lipomyelomeningocele: Lipoma-placode interface outside the spinal canal</td>
</tr>
<tr>
<td>V A:</td>
<td>Anterior lipomyelomeningocele</td>
</tr>
<tr>
<td>V B:</td>
<td>Posterior lipomyelomeningocele</td>
</tr>
<tr>
<td>Type VI</td>
<td>Intramedullary lipoma</td>
</tr>
</tbody>
</table>

**Dermal Sinus**

It is an epithelial-lined fistulous communication between the CNS or its meningeal covering and skin. It results from the focal incomplete disjunction between neuroectoderm and cutaneous ectoderm. Clinically, a midline dimple or ostium is found on the cutaneous surface and is commonly associated with cutaneous stigmata of underlying occult spinal dysraphism-like hairy nevus, hemangioma, or hyperpigmentation. The tract then ascends and opens into the spinal canal. Dermal sinus may be associated with intraspinal dermoid or epidermoid, which shows variable imaging findings depending on their contents. Dermoids usually appear hyperintense on both T1- and T2-weighted images, while epidermoids are hypointense on T1-weighted and hyperintense on T2-weighted images. CNS infection is a common complication because of fistulous communication and hence these cases require early surgical repair.

The dermal sinus is lined by squamous epithelium which is absent in the case of limited dorsal myeloschisis. Cord tenting is unusual at the site of attachment (►Fig. 23).
Limited Dorsal Myeloschisis

Limited dorsal myeloschisis (LDM) is a closed form of spinal dysraphism, characterized by the presence of a fibroneural or fibrovasculoneural tract connecting the skin lesion to the underlying spinal cord. It may be confused with congenital dermal sinus20 (Fig. 24).

Saccular LDMs (Fig. 25) occur due to accompanying hydromyelia of the fibroneural tract (swelling of the basal neural nodule with CSF or a dome-stalk type of saccular LDM). Nonsaccular “flat” LDMs may have cutaneous stigmata such as a pit, flat membrane, or a crater too. The fibroneural tract tethers the spinal cord (Fig. 26) causing characteristic tenting and the associated neurological symptoms.

A pearly white membranous crater is a characteristic cutaneous marker called “Cigarette burn mark.” Other common stigmata are dimple, pit, hypertrichosis, and hemangioma.

Low-lying cord: Persistent cord termination below L2–L3 after the first month of life in a full-gestation infant is abnormally low-lying. Axial T1-weighted images are most accurate in determining the conus level. The probability of tethered cord syndrome is more when a low-lying cord is associated with a tight filum terminale (Fig. 27).

Tethered cord: It is a clinical syndrome usually associated with closed spinal dysraphism-like spinal lipomas (Fig. 14).
Fig. 22 Retained medullary cord (A) Thickened, abnormally low termination of cord without a clearly demarcated conus. (B) With apical lipoma. (C) With ventriculus terminalis cyst (D) Hour glass configuration of thickened conus. (E) Myelocystocele with herniation of the cord. (F) Partial herniation of the cord into the myelocystocele sac.

Table 7 RMCs and a close differential diagnosis: tight filum terminale

<table>
<thead>
<tr>
<th>Retained Medullary Cord</th>
<th>Tight Filum Terminale</th>
</tr>
</thead>
<tbody>
<tr>
<td>The shape of the conus altered/not demarcated</td>
<td>Conus shape unaffected</td>
</tr>
<tr>
<td>Low-lying conus</td>
<td>Low-lying/normal conus</td>
</tr>
<tr>
<td>Usually associated with tethered cord syndrome</td>
<td>Usually associated with tethered cord syndrome</td>
</tr>
<tr>
<td>Histological findings of RMCs include a dense glioneuronal core along with fibrofatty elements, an ependymal-lined central cavity, and vestigial nerve roots19</td>
<td>No evidence of glioneuronal elements on Histopathology</td>
</tr>
</tbody>
</table>

Table 8 Differences between a Limited dorsal myeloschisis and Congenital Dermal sinus

<table>
<thead>
<tr>
<th>Limited dorsal myeloschisis (LDM)</th>
<th>Congenital dorsal dermal sinus (CDS)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Closed skin defect</td>
<td>Tract with lumen communicating with the skin opening</td>
</tr>
<tr>
<td>Fewer chances of infection and fewer neurological defects; hence, surgery needed only in cases of tethering</td>
<td>More chances of infection, more number of neurological defects, surgery even in asymptomatic cases in view of ascending infections</td>
</tr>
<tr>
<td>Attachment site: Clearly seen as hypointense round structure adhering to the dorsal aspect of the spinal cord above conus medullaris</td>
<td>Attachment site: variable and not clearly visible</td>
</tr>
<tr>
<td>Dorsal tenting of the spinal cord seen in nearly all patients</td>
<td>Dorsal tenting of the spinal cord is rare</td>
</tr>
<tr>
<td>Tethering is more common</td>
<td>Tethering is less common</td>
</tr>
<tr>
<td>Dermoid and (epi)dermoids are rare</td>
<td>Dermoid and (epi)dermoids are more common</td>
</tr>
</tbody>
</table>

Fig. 23 Dorsal dermal sinus. (A) Patient presented with a midline opening above the gluteal cleft. (B) T1 axial MinIP, (C) T2 sagittal images showing a hypointense tract extending from the skin opening into the spinal canal. (D) Different patient shows thickened T2 hypointense thickened tract with an ill-defined T2 hyperintensity within the spinal canal. (E) On contrast, there is thick peripheral enhancement. This is a dorsal dermal sinus with intraspinal abscess, due to infection.
The tight filum terminale (Fig. 16 D), split cord malformations (Fig. 32), caudal regression syndrome (Fig. 35), limited dorsal myeloschisis (Fig. 25A) meningoceles/ meningomyeloceles (Fig. 9) due to anchoring of the spinal cord/neural placode or filum terminale to the surrounding tissues occurring primarily or secondary to scarring or formation of a dermoid post-surgery and not just due to a low-lying cord or a thick filum terminale. The tethering causes functional neurological deterioration resulting in motor and sensory dysfunction, muscle atrophy, urinary incontinence, decreased or hyperactive reflexes, spastic gait, and other deformities.

**Persistent fifth ventricle:** Evidence of a fifth ventricle not accompanied by other pathologies is a frequent finding that does not have pathological significance during the first 5 years of life. It is located immediately above the filum terminale and without contrast enhancement, differentiating it from other cystic lesions of the conus medullaris (Fig. 22C).

**Disorders of Midline Notochordal Integration**

The process of fusion of paired notochordal anlagen to form a single midline notochordal process is called midline notochordal integration. Any abnormality at this stage results in longitudinal cord splitting. The most important conditions in this group are neurenteric cysts, dorsal enteric fistula (Fig. 28), and diastematomyelia.

**Dorsal enteric fistula and neurenteric cyst:** The most severe form of disorder of midline notochordal integration is a dorsal enteric fistula, which is a fistulous communication between dorsal skin surface and bowel.

The spectrum of dorsal enteric remnants consists of (1) dorsal enteric fistula, (2) dorsal enteric sinus, (3) dorsal enteric cyst (neurenteric cyst), and (4) dorsal enteric diverticulum.

The dorsal enteric fistula (Fig. 29) is due to a persistent connection between the endoderm and ectoderm, resulting in the splitting of the notochord. The fistula traverses the prevertebral soft tissue, the vertebral bodies, and the spinal canal with its contents. Any portion of this tract may involute or become fibrous, leading to a fistula or a cyst.

A Neurenteric cyst (Fig. 30) is a localized form of dorsal enteric fistula. These cysts are typically seen in the extra-medullary intradural compartment of the cervicothoracic spine; however, may be seen in other locations such as subcutaneous, intramedullary, or prevertebral planes. On MRI, neurenteric cysts usually appear iso-to-hyperintense to CSF on both T1- and T2-weighted images due to high protein content and show absent contrast enhancement.

An isolated (epi)dermoid appearing T2 hyperintense is an unusual differential in the case of intraspinal neurenteric cysts. The dermoid/epidermoid cyst occurs due to epithelial rests within the neural tube (Fig. 31).

The dorsal enteric sinus, a remnant of the posterior portion of the tract, has an opening on the skin surface. Dorsal enteric cysts are trapped remnants of the middle portion of the tract, found in the intraspinal or paraspinal compartments.

The dorsal enteric diverticulum is a tubular diverticulum arising from the bowel and represents a remnant of the anterior portion of the tract.

There exists a high incidence of associated urogenital malformations and anorectal malformations.

**Diastematomyelia**

It is the most common form of defective midline notochordal integration. Due to defective midline integration, there are
two notochordal processes each of which induces the formation of separate neural plates with primitive streak tissue in between them. The development of the primitive streak tissue decides the type of diastematomyelia.

Pang et al. coined a new term called split cord malformations and divided it into two types. In type I diastematomyelia, the intervening primitive streak develops into a bone or cartilage, resulting in two hemicords in different dural sacs separated by an osseocartilaginous septum. Each of them includes a central canal, one dorsal, and one ventral root (Fig. 32). Type I is more common than type II. In type II diastematomyelia, the primitive streak is reabsorbed or forms a fibrous septum with the hemicords lying within the same dural sac. Diastematomyelia is commonly associated with vertebral anomalies and hydromyelia. A high-lying hairy tuft over a child’s back is a reliable indicator for underlying diastematomyelia.

Diastematomyelia can be classified into three types depending on location 1) lumbar (most common), 2) lumbosacral, 3) thoracolumbar, and 4) cervicothoracic.

Composite types of split cord malformations (multi-level split cord malformations) are those with multiple levels of split cord in the same patient. Another classification, depending on the length of involvement, is divided into short-segment and long-segment types. When the split cord malformation is partial, it is called partial diastematomyelia (spinal cord cleft), seen associated with Klippel–Feil syndrome. Split cord malformations include a wide
spectrum of spinal abnormalities, which may range from simple diastematomyelia to complete duplication of the spinal column with two hemicords.

**Vertebral Duplication**

Vertebral duplication of split cord malformation (SCM) is a rare abnormality. Affected individuals present with clinical pictures ranging from mild-to-very severe symptoms which...
may be associated with gastrointestinal abnormalities such as neurenteric fistula, omphalocele or neurologic abnormalities such as myelomeningocele. Dorsolumbar and lumbar regions are the most affected sites. Vertebral duplication as such has double cords which are not separated by a septum unlike in Type 1 diastematomyelia but may be associated with diastematomyelia at a different level (Fig. 32F, G). Each cord had its own spinal canal within its own set of vertebrae.

**Segmental spinal dysgenesis:** Segmental spinal dysgenesis is characterized by a localized deformity of the thoracolumbar, lumbar, lumbosacral spine, which is associated with localized agenesis/dysgenesis spine and spinal cord. This occurs due to an abnormality of an intermediate segment of notochord occurring during gastrulation. SSD is a rare, complex, congenital, closed spinal dysraphism that meets the following criteria:

1. presence of congenital paraparesis/paraplegia with lower limb abnormalities;
2. multiple (more than one) formation and segmentation anomalies of the vertebra with or without kyphoscoliosis;
3. absent or malformed segment of the spinal cord and underlying nerve roots involving any spinal segment from the cervical to sacral region; and
4. visualization of the segment of spinal cord distal to the interrupted cord (Fig. 33).

It usually requires a multidisciplinary management approach based not only on the extent and severity of the dysgenetic cord but also on the cause (type 1 primary cord hypoplasia or type 2 secondary cord hypoplasia due to dysmorphic vertebra) of the spinal dysgenesis for further surgical management.

**Caudal agenesis/caudal regression syndrome:** It is characterized by partial or total agenesis of the spinal column and is commonly associated with genital anomalies, anal imperforation, pulmonary hypoplasia, renal aplasia or dysplasia, and limb abnormalities. This may be associated with syndromic complexes such as OEIS, VACTERL and the Currarino triad (partial sacral agenesis, anorectal malformation, and presacral mass), wherein aberrations from the stage of midline notochordal integration and mesodermal proliferation result in combined abnormalities (Fig. 34).

A low crown–rump length may be the first sign of CRS in the first trimester. At later stages, there is the abrupt interruption of the spine at dorsal, lumbar, or sacral level with femurs fixed in a “V” pattern.

Caudal regression syndrome is broadly divided into two types. In type I caudal regression syndrome, both caudal cell mass and notochord formation are affected, resulting in the high position (D12 vertebra) and abnormal termination of conus medullaris. There is an association of vertebral aplasia in the majority of patients.
In type II caudal regression syndrome, there is abnormal development of the caudal cell mass with unaffected true notochord formation. Hence, there is defective secondary neurulation with normal primary neurulation. As a result, only the most caudal part of conus medullaris is absent in type II CA (►Fig. 35). Vertebral dysgenesis is less severe in these cases and these patients present with tethered cord syndrome as the conus in these cases is stretched and tethered. In the case of lumbosacral classification, Pang’s classification (1993) describes five types:

- Type I: Total sacral agenesis (some lumbar vertebrae are missing)
- Type II: Total sacral agenesis (lumbar vertebrae are not involved)
- Type III: Subtotal sacral agenesis, S1 present
- Type IV: Hemisacrum
- Type V: Coccyegeal agenesis

**Sacroccygeal Teratoma**

Although sacrococcygeal teratoma is a tumor, the embryological origin is from the pluripotent cells of caudal cell mass. This tumor usually presents as a large complex solid–cystic mass caudal to the coccyx. Most of them are benign and contain derivatives from all three germ layers. Based on the presence of external and internal components, they are divided into four types: Type 1: primarily external, type 2: Equal external and internal portions, Type 3: primarily internal, and Type 4: entirely internal.

This could be diagnosed in antenatal ultrasound. In MRI, the mass has variable appearance depending on the internal contents, namely, fat, soft tissue, fluid, or calcium with a heterogeneous enhancement of solid components of the mass on post-contrast study.

The migration of pluripotent cells to unusual locations such as the level of cervical, dorsal, or lumbosacral vertebrae may also result in teratomas.

**Spinal/Sacral Chordoma**

Spinal/sacral chordomas are aggressive rare malignant masses of notochordal origin presenting as a firm mass. They occur most commonly in the sacrococcygeal region (30–35%). They present as a large destructive T1/T2 heterogeneous lesion with SWI/GE showing variable intralosomal hemorrhage, suggested by the presence of a blooming artifact.

**Limitations**

We have not analyzed the cutaneous manifestations and other system manifestations. We have not made a detailed analysis of vertebral anomalies and associated Arnold Chiari malformations. We have not analyzed the follow-up images of the patients after surgery.

**Conclusion**

We have tried to provide a structured approach in imaging of spinal dysraphism, with special emphasis and inclusion of recently documented unusual and complex dysraphisms, and proposed a new classification. Further study and verification are necessary to establish the clinical utility of the proposed system.

Presentation at a Meeting

Classification of Spinal Dysraphisms  Kathirivelu et al.  829

Place: Bengaluru
Date: Jan 21 to 24, 2021

Authors’ Contributions
Conception and design, acquisition of data, data analysis and interpretation, manuscript editing and reviewing and final approval.

References