First-Trimester Reference Intervals for Thyroid Function Testing among Women Screened at a Tertiary Care Hospital in India

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Abstract

Objectives Due to differences in the method of assay and population-specific factors, each laboratory needs to establish its own gestation-specific reference intervals (GRIs) for thyroid hormones.

Materials and Methods Three-hundred forty-one women with less than 14 weeks gestation were screened at a tertiary care hospital in Chhattisgarh, India. Serum levels of thyroid-stimulating hormone (TSH), free thyroxine (fT4), and thyroid peroxidase antibody (anti-TPO) were measured using an ADVIA Centaur XP immunoassay. GRIs (2.5th and 97.5th percentiles) were determined for TSH and fT4. TSH and fT4 concentrations were converted to multiples of the median (MoM) values. Effect of maternal age, gestational age, and maternal weight was analyzed.

Statistical Analysis Quantitative variables were expressed as means and standard deviations (SD), and qualitative variables were expressed as frequencies and percentages. Normality of the data was checked using the Kolmogorov–Smirnov test. Values that were normally distributed were expressed only as means and SD. Those that were not normally distributed were expressed as medians and interquartile range. For all statistical analysis, \( p < 0.05 \) was considered as statistically significant.

Results First-trimester GRI was 0.245 to 4.971 mIU/L for TSH, 10.2 to 18.9 pmol/L for fT4, and 27.0 to 56.89 kIU/L for anti-TPO. There was no significant difference in the mean serum TSH (\( p = 0.920 \)), fT4 (\( p = 0.714 \)), or anti-TPO (\( p = 0.754 \)) values among women in 4 to 7th week and 7 to 14th week of gestation. The 1st and 99th centile MoMs were 0.03 and 4.09 for TSH and 0.66 and 1.39 for fT4. There was a significant positive correlation between the maternal weight and TSH MoM values (\( p = 0.027, r = 0.120 \)).

Conclusion These laboratory- and first-trimester-specific GRI for TSH and fT4 shall help in proper diagnosis and treatment of subclinical thyroid dysfunctions. TSH and fT4 MoM values can be used to indicate high or low values in a quantitative manner independent of the reference ranges and may be used by other laboratories.

Keywords
► thyroid-stimulating hormone
► thyroxine
► reference range
► pregnancy
► multiple of median

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Introduction

Thyroid hormones (TH) are crucial for embryogenesis and fetal development. During the first half of the pregnancy, they are sourced entirely from the TH circulating in the maternal blood. Thyroid dysfunction during pregnancy increases the risk of complications such as pregnancy loss, premature delivery, gestational diabetes, gestational hypertension, low newborn birth weight, and impaired neuropsychological development.

During pregnancy, the increased demand for TH for fetal development, increased concentration of thyroxine-binding globulin due to increase in estrogen levels, and inactivation of TH by placental iodothyronine deiodinase type 3 necessitate the increased production of maternal TH. The pregnancy hormone, β-human chorionic gonadotropin, is a weak thyroid-stimulating hormone (TSH) receptor agonist. It also stimulates the thyroid gland to compensate for the increased degradation of TH. TSH level decreases and the free thyroxine (fT4) level in the maternal blood increases. Hence, there is a need for determining gestation-specific reference intervals (GRIs) for normal thyroid function. This may help to identify women who can potentially benefit from timely treatment.

Studies done at a few centers cannot be extrapolated to all centers globally due to differences in method of assay as well as population-specific factors such as ethnicity, body mass index (BMI), and iodine intake. The reference intervals for thyroid function tests in the same woman also vary from one trimester to another. The International Federation of Clinical Chemistry (IFCC), the International Committee for Standardization in Hematology, the Endocrine Society, the European Thyroid Association, and the American Thyroid Association (ATA) advise every center to determine their own population-specific GRIs.

Initial recommendations by ATA and the European Endocrinology Society had set the TSH reference limits of 0.1 to 2.5 mIU/L for the first trimester. The ATA guidelines revised in 2017 recommend that pregnancy-specific and laboratory-specific reference ranges for TSH and fT4 should be calculated. If that is not possible, a reference range can be used from the literature that is derived from a population with similar ethnicity, BMI, iodine status, etc. If both are not possible, ATA recommends the first-trimester upper cutoff limit 0.5 mIU/L less than the pre-pregnancy TSH value. If the pre-pregnancy value is unavailable, the upper limit is taken to be 4.0 mIU/L.

The definition of a thyroid disorder tends to be dependent on the TH level because of which the top 2.5% and the bottom 2.5% of people are believed to have thyroid dysfunction. This can be misleading and lead to unnecessary treatment for many people. Expressing TH levels as multiple of median (MoM) values can allow for interlaboratory assay variations, gestational age, and other factors that may affect the marker levels. To unitize laboratory reports and aid in uniform interpretation of results, MoM values may be incorporated in reporting. MoM measures how far the result deviates from the median. This value is the ratio of the patient’s result and the median result appropriate for the gestational period.

Maturation of the fetal thyroid function involves proper functional development of the hypothalamus, pituitary gland, and the thyroid gland. This process is more or less complete by the 12th to 14th week of gestation. After the 14th week of gestation, there may be irreversible effect on the fetal brain development due to lack of adequate TH in the maternal blood.

Some studies show that the median TSH in early first trimester (4–6 weeks) is significantly higher (p < 0.001) than that in the late first trimester (7–12 weeks). The Polish Society of Endocrinology recommends universal thyroid screening of all women either preconception or at their first antenatal visit (4–8 weeks). However, the current ATA guidelines cite unclear evidence in support of universal thyroid screening. They strongly recommend clinical evaluation of all patients, who seek pregnancy or are newly pregnant, for identifying risk factors of thyroid disorders. Only those with risk factors for thyroid disorders are then tested for thyroid function.

In this study, we determined the first-trimester GRIs of TH for pregnant women in Chhattisgarh state in India. We studied if the GRIs changed depending on whether the TH levels were determined in the early weeks (4–7th week) or the late weeks (7–14th week) of the first trimester. We also calculated the MoM values of TSH and fT4.

Materials and Methods

Subjects

Study Design and Setting

This cross-sectional study was conducted at a tertiary care and medical college facility in Chhattisgarh, India. The institutional ethics committee approved the study protocol and due written consent was obtained from the study participants before including them in the study.

Participants

Healthy pregnant women of more than 18 years of age, with uncomplicated single intrauterine gestations in first trimester, attending the Out Patient Department of Obstetrics and Gynecology, for antenatal care between June 2017 and September 2019, were consecutively enrolled for the study. The cutoff gestational age for the first trimester was considered to be up to 14 completed weeks, that is, 98 days. Selected subjects were assessed by the obstetrician for demographic and clinical details. Gestational age was calculated from the first day of last menstrual period (LMP). If LMP was unknown, gestational age was determined by ultrasonographic scan. Participants in early first trimester (4th–7th week) were categorized in Group A and those in late first trimester (7th–14th week) in Group B.

Women with one or more of the following were excluded from the study: (i) history and signs or symptoms of thyroid dysfunction; (ii) past or present history of intake of antithyroid drugs or thyroxine; (iii) any chronic medical disease such as hypertension, diabetes mellitus or presence of any other autoimmune disorders; (iv) history of recurrent miscarriages; (v) multiple gestations; (vi) positive thyroid
peroxidase antibody (anti-TPO) status indicated by anti-TPO levels > 60 kIU/L; (vii) TSH > 10 mIU/mL; or (viii) urinary iodine levels of ≤ 150 µg/L.

**Sample Size**
Because of the high interindividual variability and skewness for TSH and also to some extent fT4, we followed the recommendation of Clinical and Laboratory Standards Institute and IFCC, and took around 400 individual measurements for calculation of the reference interval.\(^\text{15,16}\)

**Methods**
Fasting venous blood (5 mL) was collected. Serum was separated and stored at −20°C until analysis of TSH, fT4, and anti-TPO, by chemiluminescence technique. We used commercially available kits from Siemens HealthCare Diagnostics, United States, with Siemens ADVIA Centaur XP immunoassay analyzer.

The ADVIA Centaur TSH3-Ultra assay is a third-generation assay that measures TSH concentrations from 0.008 to 150 µIU/mL (mIU/L). As per the manufacturer’s kit insert, the precision of TSH for within run coefficient of variation (CV%) and run to run CV% was (1.93–4.69) and (1.2–6.64), respectively. fT4assay is a competitive immunoassay, with sensitivity and assay range of 0.1 to 12.0 ng/dL (1.3–155 pmol/L). The within run CV% ranged from 2.23 to 3.33 and run to run CV% from 2.33 to 4.00. The ADVIA Centaur anti-TPO assay is a competitive immunoassay with sensitivity and assay range of 15 to 1,300 U/mL or kIU/L of within run CV% (1.3–6.8) and run to run CV% (2.8–3.4).

Spot urine sample (5 mL) was collected and stored at −20°C. Urinary iodine concentration was determined by the wet digestion method on the basis of Sandell–Kolthoff reaction.\(^\text{17}\)

**Statistical Analysis**
All statistical analysis was done by using SPSS trial version and in MS-Excel 2007. Quantitative variables were expressed as means and standard deviations and qualitative variables were expressed as frequencies and percentages. Normality of the data was checked by using Kolmogorov–Smirnov test. Values that were normally distributed were expressed only as means and standard deviation. Those that were not normally distributed were expressed as medians and interquartile range. Analysis of variance was used as a test of difference of more than two means and linear regression analysis was used for finding the correlation. Chi-squared test was used for examining the categorical variables. For all statistical analysis, \( p < 0.05 \) was considered as statistically significant.

The 2.5th and 97.5th percentiles for TSH, fT4, and anti-TPO were determined in the first trimester to find the reference range for each of these measures. MoM values were calculated for TSH and fT4.

**Results**

**Study Population**
A total of 412 consecutive women in their first trimester were screened for the study. Nine of them were either on TH medication or had hypothyroid blood profile, one had a twin pregnancy, and one was diagnosed with sickle-cell trait. Seven women had a TSH > 10 mIU/mL and 56 had anti-TPO > 60 kIU/L. After excluding these patients, the data of 341 women was analyzed (\(<\text{Table 1}\>\).

The median age was 25 years (range: 18–42 years) and the mean BMI was 20.69 kg/m\(^2\) (range: 13.01–36.20 kg/m\(^2\)). Of the 341 women analyzed, 150 (44.0%) were primigravida and 175 (51.3%) were nulliparous.

**Table 1** Descriptive characteristics of the study population \((n = 341)\)

<table>
<thead>
<tr>
<th>Parameter</th>
<th>Value</th>
</tr>
</thead>
<tbody>
<tr>
<td>Median age in years (range)</td>
<td>25 (18–42)</td>
</tr>
<tr>
<td>Mean BMI in kg/m(^2) ± SD (range)</td>
<td>20.69 ± 3.474 (13.01–36.20)</td>
</tr>
<tr>
<td>Obstetric history</td>
<td></td>
</tr>
<tr>
<td>Mean gestational age (in days) ± SD at presentation (range)</td>
<td>63.19 ± 16.768 (31–98)</td>
</tr>
<tr>
<td>Number of women in 4–7 weeks of gestation (%)</td>
<td>86 (25.2)</td>
</tr>
<tr>
<td>Number of women in 7–14 weeks of gestation (%)</td>
<td>255 (74.8)</td>
</tr>
<tr>
<td>Gravida (%)</td>
<td></td>
</tr>
<tr>
<td>1</td>
<td>150 (44.0)</td>
</tr>
<tr>
<td>2</td>
<td>125 (36.6)</td>
</tr>
<tr>
<td>3</td>
<td>51 (15.0)</td>
</tr>
<tr>
<td>4</td>
<td>12 (3.5)</td>
</tr>
<tr>
<td>5</td>
<td>3 (0.9)</td>
</tr>
<tr>
<td>Para (%)</td>
<td></td>
</tr>
<tr>
<td>0</td>
<td>175 (51.3)</td>
</tr>
<tr>
<td>1</td>
<td>131 (38.4)</td>
</tr>
<tr>
<td>2</td>
<td>34 (10.0)</td>
</tr>
<tr>
<td>3</td>
<td>1 (0.3)</td>
</tr>
<tr>
<td>Living (%)</td>
<td></td>
</tr>
<tr>
<td>0</td>
<td>192 (56.3)</td>
</tr>
<tr>
<td>1</td>
<td>129 (37.8)</td>
</tr>
<tr>
<td>2</td>
<td>19 (5.6)</td>
</tr>
<tr>
<td>3</td>
<td>1 (0.3)</td>
</tr>
<tr>
<td>Abortion (%)</td>
<td></td>
</tr>
<tr>
<td>0</td>
<td>255 (74.8)</td>
</tr>
<tr>
<td>1</td>
<td>70 (20.5)</td>
</tr>
<tr>
<td>2</td>
<td>12 (3.5)</td>
</tr>
<tr>
<td>3</td>
<td>4 (1.2)</td>
</tr>
</tbody>
</table>

Abbreviations: BMI, body mass index; SD, standard deviation.

First-Trimester Thyroid Reference Ranges  Nanda et al.
in Table 2. According to our study, in the first trimester, the reference interval of serum TSH was 0.25 to 4.97 mIU/L and of serum fT4 was 10.2 to 18.9 pmol/L. The reference interval for anti-TPO was 27.0 to 56.89 kIU/L.

There was no significant difference in the mean serum TSH, fT4, or anti-TPO values among women in Group A and Group B (p = 0.920, 0.714, and 0.754, respectively). The median values and the reference ranges of TSH, fT4, or anti-TPO were similar between the two groups (Table 3). There was a significant (r = −0.189, p < 0.001) inverse correlation between TSH and fT4 levels and a significant (r = 0.167, p = 0.002) positive correlation between urinary iodide and anti-TPO levels. There was no statistically significant difference in either serum TSH level (p = 0.265, r = 0.164, Supplementary Fig. S1A) or serum fT4 (p = 0.059, r = 0.176, Supplementary Fig. S1B) level across increasing gestational age in the first trimester.

Table 3 shows selected centiles of TSH and fT4 in conventional units and their MoM values. MoM values for 1st and 99th centiles for TSH were 0.03 and 4.09, respectively. Corresponding values for fT4 were 0.66 and 1.39, respectively.

We found a significant positive correlation between the maternal weight and TSH MoM values (p = 0.027, r = 0.120) but not between maternal age and TSH MoM values (p = 0.164, r = 0.076) (Supplementary Figs. S2 and S3). In contrast, there was no significant correlation between maternal weight and fT4 MoM values (p = 0.749, r = −0.017) and between maternal age and fT4 MoM values (p = 0.176, r = 0.073) (Supplementary Figs. S2 and S3).

**Discussion**

In our study, in the first trimester, the GRI of serum TSH was 0.25 to 4.97 mIU/L. The upper limit was higher than that mentioned in ATA guidelines as well as obtained from several other studies (Table 5). This may be due to the difference in study design, sample size used, ethnicity, region, and/or assay method.

Low maternal TSH that is still detectable is unlikely to be of clinical significance. However, since the normal upper limit of maternal TSH in our study population is higher than that set by ATA guidelines, the use of the GRI of ATA for pregnant women attending our institute could lead to misdiagnosis and mistreatment in many women. Overtreatment with TH can hinder proper fetal brain development and increase the risk of child mental health disorder, ADHD, and autism spectrum disorder.18 Therefore, we needed to set our own institutional GRI.
The definition of thyroid dysfunction is also dependent on the fT4 concentration. In our study, serum fT4 reference range during the first trimester was from 10.2 to 18.8 pmol/L. Rajput et al.\textsuperscript{19} reported first trimester serum fT4 range as 0.88 to 1.78 ng/mL that is equivalent to 1,132.560 to 2,290.860 pmol/L. These values are almost 100 times more than ours and most other studies (\textsuperscript{►}Table 5). We wonder if there was an error in the use of units by Rajput et al.\textsuperscript{19} They report their fT4 values in ng/mL and not in ng/dL as done by researchers of other Indian studies.\textsuperscript{20–22}

When fT4 is within normal limits but TSH level is persistently above the normal limit, a biochemical diagnosis of subclinical hypothyroidism can be confirmed. TSH levels within normal range of laboratory do not always mean the

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**Table 4** Selected centiles of TSH and fT4 in conventional units and their MoM values

<table>
<thead>
<tr>
<th>Centile</th>
<th>TSH in mIU/L</th>
<th>TSH MoM</th>
<th>fT4 in pmol/L</th>
<th>fT4 MoM</th>
</tr>
</thead>
<tbody>
<tr>
<td>1</td>
<td>0.06</td>
<td>0.03</td>
<td>9.525</td>
<td>0.66</td>
</tr>
<tr>
<td>2</td>
<td>0.203</td>
<td>0.11</td>
<td>10.01</td>
<td>0.69</td>
</tr>
<tr>
<td>2.5</td>
<td>0.245</td>
<td>0.13</td>
<td>10.17</td>
<td>0.71</td>
</tr>
<tr>
<td>5</td>
<td>0.416</td>
<td>0.23</td>
<td>10.97</td>
<td>0.76</td>
</tr>
<tr>
<td>10</td>
<td>0.56</td>
<td>0.31</td>
<td>11.97</td>
<td>0.83</td>
</tr>
<tr>
<td>50</td>
<td>1.82</td>
<td>1</td>
<td>14.42</td>
<td>1</td>
</tr>
<tr>
<td>90</td>
<td>3.78</td>
<td>2.08</td>
<td>17.12</td>
<td>1.19</td>
</tr>
<tr>
<td>95</td>
<td>4.309</td>
<td>2.37</td>
<td>17.76</td>
<td>1.23</td>
</tr>
<tr>
<td>97.5</td>
<td>4.971</td>
<td>2.73</td>
<td>18.83</td>
<td>1.31</td>
</tr>
<tr>
<td>98</td>
<td>5.978</td>
<td>3.28</td>
<td>19.19</td>
<td>1.33</td>
</tr>
<tr>
<td>99</td>
<td>7.446</td>
<td>4.09</td>
<td>20.03</td>
<td>1.39</td>
</tr>
</tbody>
</table>

Abbreviations: fT4, free thyroxine; MoM, multiples of median; TSH, thyroid stimulating hormone.

1 pmol/L = 0.078 ng/dL; 1 kIU/L = 1 mIU/L.

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**Table 5** Comparison of reference ranges for thyrotropin and fT4 during early pregnancy

<table>
<thead>
<tr>
<th>Author(s)\textsuperscript{Ref}, country</th>
<th>n</th>
<th>Gestation week</th>
<th>Assay used</th>
<th>TSH, mIU/L</th>
<th>fT4, ng/dL (pmol/L)</th>
</tr>
</thead>
<tbody>
<tr>
<td><strong>Studies done outside India</strong></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Bestwick et al,\textsuperscript{9} Italy</td>
<td>5,505</td>
<td>&lt; 16</td>
<td>CL (Auto DELFIA\textsuperscript{a})</td>
<td>1.07</td>
<td>0.04–3.19</td>
</tr>
<tr>
<td>Bestwick et al,\textsuperscript{9} UK</td>
<td>16,334</td>
<td>&lt; 16</td>
<td>CL (ADVIA Centaur\textsuperscript{b})</td>
<td>1.11</td>
<td>0.06–3.50</td>
</tr>
<tr>
<td>Karakosta et al,\textsuperscript{26} Greece</td>
<td>141</td>
<td>&lt; 13</td>
<td>CL (IMMULITE 2000\textsuperscript{b})</td>
<td>1.02</td>
<td>0.05–2.53</td>
</tr>
<tr>
<td>Li et al,\textsuperscript{11} China</td>
<td>1,024</td>
<td>4–12</td>
<td>CobasElesys 601\textsuperscript{d}</td>
<td>1.66</td>
<td>0.14–4.87</td>
</tr>
<tr>
<td>Zhang et al,\textsuperscript{27} China</td>
<td>132</td>
<td>1–12</td>
<td>CL (Centaur CP\textsuperscript{b})</td>
<td>NR</td>
<td>0.02–3.78</td>
</tr>
<tr>
<td><strong>Studies done in India</strong></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Maji et al,\textsuperscript{20} India</td>
<td>125</td>
<td>&lt; 13</td>
<td>ELISA (Acubind\textsuperscript{e})</td>
<td>1.8</td>
<td>0.25–3.35</td>
</tr>
<tr>
<td>Mankar et al,\textsuperscript{21} India</td>
<td>50</td>
<td>8–10</td>
<td>CL Immunoassay (NR)</td>
<td>NR</td>
<td>0.24–4.17\textsuperscript{a}</td>
</tr>
<tr>
<td>Rajput et al,\textsuperscript{19} India</td>
<td>301</td>
<td>≤ 12</td>
<td>ECL (ADVIA Centaur CP\textsuperscript{b})</td>
<td>1.40</td>
<td>0.37–3.69</td>
</tr>
<tr>
<td>Pramanik et al,\textsuperscript{22} India</td>
<td>80</td>
<td>NR</td>
<td>CL (IMMULITE 1000\textsuperscript{b})</td>
<td>NR</td>
<td>0.19–4.34</td>
</tr>
<tr>
<td>Our study, India</td>
<td>341</td>
<td>≤ 14</td>
<td>CL (ADVIA Centaur XP\textsuperscript{b})</td>
<td>1.820</td>
<td>0.25–4.97</td>
</tr>
</tbody>
</table>

Abbreviations: CL, chemiluminescence; ECL, electrochemiluminescence; ELISA, enzyme-linked immunosorbent assay; fT4, free thyroxine; NR, not reported; TSH, thyroid stimulating hormone.

1 pmol/L = 0.078 ng/dL; 1 kIU/L = 1 mIU/L.
\textsuperscript{a}PerkinElmer Life and Analytical Sciences.
\textsuperscript{b}Siemens Healthcare Diagnostics.
\textsuperscript{c}Ortho Clinical Diagnostics.
\textsuperscript{d}Roche Diagnostics.
\textsuperscript{e}Monobind Inc.
\textsuperscript{f}5th and 95th centiles were used to determine the reference range.
\textsuperscript{g}FT4 values were calculated in 139 women.
\textsuperscript{h}Seems like an error in the cited study as ADVIA Centaur CP uses CL and not, ECL.
\textsuperscript{i}Mentioned as 0.88–1.78 ng/mL in the report.

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individual has normal thyroid function. The TSH value may be outside the individual reference range. However, it is impractical to take several samples and measure individual reference range for each patient. Laboratory-specific reference range is preferred. It is not difficult to calculate laboratory-specific reference range. Every hospital where perinatal care is provided can collaborate with its clinical biochemistry department and conduct a study to determine their own laboratory-specific GRIs for TSH and fT4.

Andersen et al.13 and Laurberg et al.1 report that the use of uniform reference range for TSH and fT4 in early pregnancy may be too simple and can lead to frequent misclassification and incorrect use of therapy. They suggest the use of a reference range stratified by the week of pregnancy because of the dynamic physiological changes seen over weeks. We observed no significant difference in the TSH and fT4 reference ranges among women in early or women in late first trimester. Khalil et al.24 had also found no difference in the 4 to 6 weeks and 7 to 12 weeks gestation groups.

TSH ranges can vary slightly depending on the method of analysis used. This variability can be reduced by using MoM values.13 MoM is often used for screening test results. Computer software is available that can convert marker levels of Down syndrome or neural tube defects to MoM values. Similar software can be used to calculate MoM values for TSH and fT4 levels. Data from approximately 100 patients would be sufficient.5 We have used data from 341 patients to determine the MoM values.

An individual value is divided by the population median and the resulting value is not influenced by the method of assay used. Reporting the MoM value along with the centile, it corresponds to, does not put a woman into the abnormal category that can happen when only the reference range is used.9 MoM values also take into account certain factors such as gestational age and maternal weight that can have a relationship with TSH and fT4 values.9

Considering obesity is increasing in many populations, it may be an important finding that maternal weight correlated with TSH MoM values (p = 0.027, r = 0.120). Bestwick et al.9 had found that for every 10 kg increase in weight, there was 0.025 MoM increase in TSH MoM values and 0.009 MoM decrease in fT4 MoM values. In contrast, we found no significant correlation between maternal weight and fT4 MoM values (p = 0.749, r = -0.017).

The 1st to 99th percentiles of TSH and fT4 MoM values in our study were 0.03 to 4.09 and 0.66 to 1.39, respectively. In a study by Kianpour et al.,25 among pregnant Iranian women with gestational age ≤ 14 weeks, the corresponding values were 0.06 to 4.62 and 9.00 to 18.02, respectively. In the study by Bestwick et al.,9 among pregnant women (gestational age less than 16 weeks) from United Kingdom, the 1st to 99th percentiles of TSH and fT4 MoM values were 0.02 to 4.13 and 0.75 to 1.38, respectively. Among those from Italy, corresponding values were 0.01 to 3.93 and 0.76 to 1.46, respectively. Both our TSH and fT4 MoM results match those from the two populations studied by Bestwick et al.,9 whereas only our TSH MoM values match those from the study by Kianpour et al.25 The first-trimester–specific reference interval for serum fT4 is similar in both our study and that by Kianpour et al, but the fT4 MoM values are grossly different. A careful look at the fT4 MoM values published by them in a 2019 issue of Hormone and Metabolic Research suggests that there might have been some error as the numerical values of MoM are exactly the same as the individual centile values for fT4.25

To the best of our knowledge, ours is the largest Indian study of women in their first trimester that determines the laboratory-specific GRIs for TSH and fT4. It is also probably the first one in South East Asia to determine the MoM values for TSH and fT4 during the first trimester. Another strength of our study was the measurement of anti-TPO, urine iodide, and detection of multiple gestations through ultrasound scanning. It helped in a clearer definition of the normal population for reference ranges. We could also have attempted to determine the GRIs for the second and third trimester using this sample population. Though our sample size of 341 pregnant women in their first trimester is perhaps the largest in India, a larger sample size could have provided more accurate reference intervals.

To obviate the slightly different, although overlapping, reference ranges of various commercially available assays, we have used the MoMs. This may allow other laboratories to use our specific MoM thresholds to their centers once they have established their own first trimester-specific medians. Further research is needed to validate the extrapolation of our specific MoM values to other centers.

**Conclusion**

In this study, GRIs of serum TSH and fT4 were determined for singleton pregnant Indian women with anti-TPO less than 60 kIU/L and adequate level of iodine. The GRIs determined in our study are different from that of ATA and the GRIs determined by other studies (Table 5). Our laboratory-specific reference ranges may be used for the pregnant women in Chhattisgarh. Once first-trimester median values are obtained by a laboratory, it will also be able to use our MoM values as cutoffs to obviate differences in the assay methods. This can aid accurate detection of thyroid disorders during pregnancy in a specific population.

**Authors’ Disclosure Statement**

No competing financial or personal interest.

**Authorship Participation**

R.N. was involved in concept and design of work, sample analysis, and wrote first draft of the manuscript. P.K.N. was involved in design of work, acquisition, and analysis of data. S.P. was involved in interpretation of data and drafting of manuscript. E.M. was involved in sample analysis and revising the work critically, and S.A. was involved in acquisition and interpretation of data and revising the work critically. All authors have reviewed and approved of the manuscript prior to submission. This manuscript has been submitted solely to this journal and is not published in press or submitted elsewhere.
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Conflict of Interest
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