Spinal Epidural Lipomatosis Associated with Intrathecal Flow Voids: Demonstration of Engorged Veins Using Flat Panel Catheter Angiotomography

Nishtha Yadav1, Ketan Hedaoo2, Ambuj Kumar2

1 Department of Neuroradiology, School of Excellence in Neurosurgery, Super Specialty Hospital, Netaji Subhash Chandra Bose Medical College, Jabalpur, Madhya Pradesh, India

2 Department of Neurosurgery, Super Specialty Hospital, Netaji Subhash Chandra Bose Medical College, Jabalpur, Madhya Pradesh, India


Abstract

Keywords

► spinal epidural lipomatosis
► flow voids
► flat panel catheter angiotomography
► spinal angiography

We present a case of a 54-year-old male with spinal epidural lipomatosis who had associated flow voids on magnetic resonance imaging with dilated intrathecal vessels. During spinal angiogram, 20s DynaCT (flat panel catheter angiotomography) was utilized to demonstrate the intrathecal engorged veins. Venous engorgement of epidural venous plexus has been previously described in epidural lipomatosis; however, dilated intrathecal perimedullary veins have not been demonstrated by imaging. We have described the utility of flat panel catheter angiotomography in understanding venous disorders in such patients.

Introduction

Spinal epidural lipomatosis is a rare and complex disorder that can present with progressive neurological deficits and is caused by hypertrophy of adipose tissue in spinal epidural space. We present a rare case of spinal epidural lipomatosis associated with flow voids on magnetic resonance imaging (MRI). In our patient, the flow voids were due to intrathecal perimedullary vein engorgement, which was demonstrated using flat panel catheter angiotomography (FPCA). Although flow voids due to engorged veins in spinal epidural lipomatosis are rarely found, high degree of suspicion of venous pathology in such cases (rather than a spinal vascular malformation) should be considered especially in a setting of normal spinal angiography and FPCA should be utilized in demonstration of the same.

Case Report

A 54-year-old male presented with history of gradually progressive weakness for past 2 years. At the time of presentation, he was using cane for walking. There was no history of trauma, fever, drug intake, or infections. No history of bowel/bladder incontinence was noted. Physical examination revealed body mass index 22 kg/m² (weight 60 kg, height 165 cm) without any signs of endocrinopathies/Cushing’s syndrome. Power in bilateral lower limbs with preserved muscle bulk. Knee and ankle jerks. Bilateral planters were extensor. There was no sensory loss. There was no spinal deformity or tenderness. MRI was done which revealed T1, T2 hyperintense lesion...
in posterior epidural space extending from D2 to D9 levels, causing spinal canal compromise and anterior displacement of cord (► Fig. 1). The lesion showed hypointense signal on fat saturated images and no enhancement was noted. No cord signal changes were noted. Intrathecal flow voids were noted from D9 to D11 levels, with presence of serpiginous enhancement posterior to cord at same location (► Fig. 1). Diagnosis of thoracic epidural lipomatosis was made with spinal canal stenosis and myelopathy. However, due to presence of myelopathy along with intrathecal flow voids, a spinal angiogram was planned to rule out a coexisting spinal vascular malformation.

Spinal angiogram did not reveal evidence of spinal vascular malformation. Injection of left D10 intercostal artery showed opacification of artery of Adamkiewicz. Delayed acquisition revealed venous phase at 15 seconds with visualization of engorged veins at the same level as noted on MRI (D9–D11) (► Fig. 2). Initial FPCA was done after selective injection of left D10 intercostal artery using a 5-second acquisition (DynaCT) on Artis zee, (Siemens, Erlangen, Germany). Continuous hand injection of 10 mL of 50% diluted iodinated contrast agent (300 mgI/mL) was done using 20 cc syringe while acquiring 5s DynaCT. Data was reconstructed using high-resolution matrices (0.1 mm voxel size). There was opacification of anterior spinal artery, with no opacification of early draining veins/fistula (► Fig. 2). After 5 minutes, another FPCA was done after similar selective injection of left D10 intercostal artery using a 20-second acquisition (DynaCT, Siemens). The reconstructed image evaluation revealed presence of engorged anterior median spinal vein, posterior median spinal vein, extrinsic surface anastomosis between anterior and posterior median vein, and few other dilated perimedullary veins (► Fig. 3).
engorged perimedullary veins demonstrated on FPCA were considered the cause of flow voids on MRI and spinal vascular malformation was ruled out. The patient underwent surgical excision of epidural lipomatosis and had improvement in weakness postoperatively.

**Discussion**

Spinal epidural lipomatosis can present with back pain, limb weakness, sensory changes, radicular pain, or rarely bowel/bladder incontinence. The neurological symptoms are dependent on the degree and level of canal compromise (cervical/thoracic/lumbar leading to cord/conus/cauda equine compression). Myelopathy is attributed to direct effects of mechanical compression due to spinal canal stenosis as well as indirectly due to compression of the epidural blood vessels with subsequent venous engorgement. Intraoperative epidural venous engorgement has been noted during excision of epidural lipomatosis. Engorgement of epidural venous with epidural lipomatosis has been described by Park et al on MRI. However, dilatation of intrathecal perimedullary veins has rarely been described previously.

Presence of myelopathy along with flow voids on MRI requires further imaging in form of spinal angiogram to rule out spinal vascular malformation. In a study by Alhilali et al, they aimed to determine prevalence of spinal dural arteriovenous fistulae (SDAVF) in patients presenting with prominent vascular flow voids on imaging without other imaging findings suggestive of SDAVF. They studied 18 patients with flow voids on MRI, without any other imaging evidence of spinal vascular malformation (such as cord signal change/venous infarct), and found only 17% patients had SDAVF on angiography, and all of them had myelopathy symptoms. They concluded that prominent flow voids without other imaging findings suggestive of SDAVF are poorly predictive of the presence of a SDAVF, unless myelopathy is present clinically. They also noted a negative spinal angiogram in one of their patients who had associated epidural lipomatosis, similar to our patient. In our case, we were further able to demonstrate the engorged veins in venous phase on spinal angiogram using FPCA.

Demonstration of venous phase in spinal digital subtraction angiography (DSA) is difficult because it requires the patient to hold the breath for 15 to 20 seconds. Additionally, bowel peristalsis and poor bowel preparation also make it difficult to evaluate venous phase on spinal angiogram. Even though it is generally considered that demonstration of venous phase (within 18 seconds) usually indicates absence of spinal vascular malformation this is not always true. In a study by Eckart Sorte et al, they concluded that the mere observation of a normal venous phase cannot be used to exclude the presence of a vascular malformation or justify interrupting a diagnostic spinal angiogram. However, demonstration of venous phase/perimedullary veins is very useful and improves the understanding of venous system anatomy and hemodynamics.

Previous studies have evaluated the utility of FPCA as an adjunct to spinal angiography and have found it to be particularly valuable in detection of spinal venous anomalies. FPCA has many advantages for the demonstration of spinal venous phase. FPCA provides high-resolution images of the spinal vasculature, particularly, of the spinal veins, in any desired plane. Moreover, there is no need for breath hold because breathing and bowel peristalsis artifacts do not affect the quality of reconstructed images in FPCA. The rich information provided by FPCA helps in the treatment planning for vascular malformations. Additionally, FPCA can transform a “negative spinal angiogram” into a positive angiographic diagnosis, thus eliminating the need of repeat studies/additional investigations for such patients that are usually performed due to the fear of an overlooked lesion after a negative spinal angiogram. Our case exemplifies this situation. While presence of myelopathy along with intrathecal flow voids on MRI had raised the suspicion of a

**Fig. 3** (A) Sagittal, (B) axial, and (C) coronal reconstruction of 20s DynaCT (flat panel catheter angiotomography) show visualization of anterior median spinal vein (black arrow in [A]), posterior median spinal vein (white arrow in [A]), extrinsic surface anastomosis between anterior and posterior median vein (arrowhead in [B]), and engorged perimedullary veins (curved arrow in [C]).
vascular malformation in our patient, the spinal DSA was normal. However, the findings on FPCA showing engorged veins are correlated with the abnormalities on MRI (flow voids and serpiginous enhancement) along the dorsal surface of the spinal cord in the same location. Epidural lipomatosis has been known to cause compression of epidural veins with venous engorgement. Compacted epidural venous plexus may lead to transmission of increased pressure to perimedullary veins with resultant intrathecal venous engorgement. This case further demonstrates the usefulness of FPCA technique in improving spinal venous imaging. FPCA can help in better understanding of poorly understood conditions involving the spinal venous system and in future can possibly demonstrate venous disorders that are not as yet appreciated.

**Conclusion**

We report a case of spinal epidural lipomatosis with associated intrathecal flow voids. Venous engorgement of epidural venous plexus has been previously described in epidural lipomatosis; however, dilated intrathecal perimedullary veins have not been demonstrated by imaging. We have described the utility of FPCA in understanding venous disorders in such patients.

**Ethical Approval**

The study has been approved by the appropriate ethics committee and has therefore been performed in accordance with the ethical standards laid down in the 1964 Declaration of Helsinki and its later amendments.

**Funding**

None.

**Conflict of Interest**

None declared.

**References**