Genetic Diversity of Human Immunodeficiency Virus Type 1 in Asymptomatic Blood Donors in Islamabad, Pakistan

Usman Waheed1,2, Farooq Ahmed Noor3, Noor e Saba4, Akhlaaq Wazeer3, Zahida Qasim3, Muhammad Arshad5, Saira Karimi6, Ahmad Farooq5, Javaid Usman7, Hasan Abbas Zaheer1,8

1Department of Pathology and Transfusion Medicine, Shaheed Zulfiqar Ali Bhutto Medical University, Islamabad, Pakistan
2Islamabad Blood Transfusion Authority, Ministry of National Health Services, Government of Pakistan, Islamabad, Pakistan
3Department of Pathology and Transfusion Medicine, Divisional Headquarters Teaching Hospital, Mirpur, Azad Jammu and Kashmir, Pakistan
4Department of Health, Peshawar Regional Blood Centre, Khyber Pakhtunkhwa, Pakistan
5Department of Biological Sciences, International Islamic University, Islamabad, Pakistan
6Department of Biosciences, COMSATS University, Islamabad, Pakistan
7Department of Pathology, Army Medical College, National University of Medical Sciences, Rawalpindi, Pakistan
8Safe Blood Transfusion Programme, Ministry of National Health Services, Government of Pakistan, Islamabad, Pakistan

Address for correspondence Usman Waheed, PhD, Department of Pathology and Transfusion Medicine, Shaheed Zulfiqar Ali Bhutto Medical University, Islamabad 44000, Pakistan (e-mail: drusman.waheed1@gmail.com).

Introduction

Objective The serological testing of human immunodeficiency virus (HIV) is mandatory under the blood safety legislation of Pakistan; hence, data exist on the prevalence of HIV in blood donors. However, little is known about the molecular epidemiology of HIV in the blood donor population. Therefore, the current study was designed to study the genetic diversity of HIV-1 infection in a population of apparently healthy treatment-naive blood donors in Islamabad, Pakistan.

Material and Methods A total of 85,736 blood donors were tested for HIV by the chemiluminescence immunoassay. All positive donor samples were analyzed for the presence of various HIV genotypes (types and subtypes). Viral ribonucleic acid was extracted from blood samples of HIV positive donors and reverse transcribed into complementary deoxyribonucleic acid (cDNA). The cDNA of all positive donors was then analyzed for the presence of various HIV genotypes (types and subtypes) by employing subtype-specific primers in a nested polymerase chain reaction. The amplified products were run on ethidium bromide-stained 2% agarose gel and visualized using an ultraviolet transilluminator. A particular subtype was assigned to a sample if the subtype-specific reaction made a band 20% highly intense compared with the band made by the subtype-independent reaction.

Results A total of 85,736 blood donors were screened for the presence of antibodies to HIV. Out of them, 114 were initially found reactive for HIV. The repeat testing resulted in 112 (0.13%) positive donors, 95% confidence interval 0.0014 (0.0011–0.0018). These 112 samples were analyzed for molecular typing of HIV-1. The predominant HIV-1 subtype was A (n = 101) (90.1%) followed by subtype B (n = 11) (9.9%).

Conclusion These findings are key to understand the diversified HIV epidemic at the molecular level and should assist public health workers in implementing measures to lessen the further dissemination of these viruses in the country.
The human immunodeficiency virus (HIV) is the causative agent of acquired immunodeficiency syndrome (AIDS) in humans which leads to progressive failure of the immune system allowing life-threatening opportunistic infections to flourish. The epidemiologic and phylogenetic studies suggest that HIV was introduced to humans from 1920 to 1940. HIV is a single-stranded ribonucleic acid (RNA) retrovirus surrounded by an envelope with glycoproteins (gp) 120 and gp41 responsible for viral attachment to the host. Classified as HIV-1 and HIV-2 on the genetic differences and other genetic characteristics, both types have many similarities such as mode of transmission and basic gene arrangement, among others. HIV-1 has evolved from nonhuman primate immunodeficiency virus from Central African chimpanzees (SIVcpz) whereas HIV-2 evolved from the West African sooty mangabeys (SIVsm). HIV-1 incidence is global while HIV-2 is largely limited to West Africa.

The AIDS pandemic has had a devastating multifaceted effect on the lives of people globally. According to the World Health Organization (WHO) estimates, HIV has infected approximately 70 million people since 1981 and is responsible for the death of more than 32 million people so far. By the end of 2018, around 37.9 million population were found to be living with HIV worldwide with 1.7 million people becoming newly infected in 2018.

Pakistan, a developing nation of 220 million inhabitants, is witnessing an increase in the number of HIV-infected individuals. The epidemic is now well established and intensifying in injection drug users and the sex workers (including male and transgender sex workers), forming the core of the epidemic. According to the National AIDS Control Program, 180,000 individuals are living with HIV/AIDS in the country, 39,529 are registered while 22,947 are receiving antiretroviral therapy. The number of infected individuals will be much higher if systematic mass screening is performed nationally.

It is imperative to strengthen knowledge and evidence about HIV/AIDS in responding effectively to the global epidemic. Countries around the globe have invested in clinical and molecular research that tries to inhibit, treat, or remedy HIV/AIDS, as well as vital research about the characteristics of HIV as a communicable agent and AIDS as the syndrome instigated by HIV. But still more research is essential to support the millions of individuals whose health remains at risk by the global HIV/AIDS pandemic. Testing for molecular epidemiology of HIV infection is of paramount importance to develop sensitive diagnostic tools, to manage individual infections, and also to keep track of transmission patterns, prevalence, and evolution of the pandemic.

In Pakistan, molecular epidemiology investigations are currently at infancy and have yet to contribute to the formulation of a well-versed HIV prevention policy and programming. The earlier reported data on HIV-1 genotypes showed the epidemic is vastly diverse involving cocirculation of genotypes mainly “A” followed by genotypes “B,” CRF_AE, CRF_AG. Despite this molecular characterization of the diverse genetic complexity of HIV-1 among various high-risk groups in Pakistan, the genotypic prevalence of HIV-1 among the blood donor population has yet to be explored. According to WHO, an estimated 117.4 million blood donations take place each year and transfusion of contaminated blood and blood components carries a significant risk of transmitting blood-borne infections including HIV, and hepatitis B and C to the patients. Transmission of HIV infection due to contaminated blood and blood components transfusion has been reported repeatedly since 1982. In Pakistan, the incidence of HIV infection in healthy donors ranged from 0.008% in Peshawar city to 1.0% in Gwadar city. Assessing the pattern of HIV in blood donors is crucial to guarantee the safety of blood and also to sensitize the policymakers on the level of the disease in the apparently healthy population. Hence, this study aimed to characterize the genetic diversity of HIV-1 infection among a population of apparently healthy treatment-naive blood donors in Islamabad from 2016 to 2018.

Material and Methods

This was a cross-sectional, prospective single-center study conducted at the Department of Pathology and Transfusion Medicine, Shaheed Zulfiqar Ali Bhutto Medical University, Islamabad, Pakistan, from January 2016 to December 2018. The study protocol was endorsed by the Ethics Committee of the Shaheed Zulfiqar Ali Bhutto Medical University, Islamabad.

All the data and samples were kept confidential and anonymous. HIV analyses were performed according to the conditions of “5-Cs”: comprising of informed consent, be confidential, involve counseling, deliver correct test results, and connections to prevention, treatment, and care services. Informed written consent was received from all study participants enrolled in this study. Clinical and epidemiological data were maintained only for therapeutic and research purposes. No monetary incentive was offered.

The blood donors were selected after careful examination in the donor management department of the blood bank following the national guidelines. According to general inclusion criteria, a donor aged 18 to 60 years, weight > 50 kg, and hemoglobin level of > 12.5 g/dL were selected. Pulse (60–100 beats per minute) and blood pressure (systolic not more than 160 and diastolic not more than 100 mm of Hg without medication) were checked as well. Any potential donor with pregnancy, lactation, recent blood donation, and vaccination were temporarily deferred. In addition, the donors answered a detailed history questionnaire about their health to determine any risk factor that can temporarily or permanently defer a donor.

Serological Analyses

A total of 85,736 blood donors were screened for HIV along with other transfusion-transmitted infections (TTIs) during the study period. HIV screening was performed by the chemiluminescence immunoassay (CLIA) using a fully automated Abbott Architect i2000SR system. The CLIA HIV is a combo immunoassay which is a two-step procedure to detect the presence of HIV-1 p24 antigen and antibodies to
HIV-1 (groups M and O) and HIV-2. The system measures the cutoff by utilizing the mean chemiluminescent signal (relative light units) from three copies of the Calibrator 1 and records the result. The samples tested positive were retested using Abbott’s CLIA system. The plasma samples were stored at –80°C for further molecular analyses.

**Molecular Analyses**

Viral RNA was extracted from blood samples of HIV positive donors using ExiPrepDx Viral RNA extraction kit. Extraction was performed according to the manufacturer’s protocol and included sample preparation, extraction process setup, pipetting samples into the specimen loading tubes, and executing the extraction by operating the ExiPrep16 Dx. The extracted viral RNA was automatically loaded into the elution tubes. After the RNA extraction, reverse transcription procedure was followed where the RNA was reverse transcribed using a commercially available kit (Thermo Fisher Scientific) into complementary deoxyribonucleic acid (cDNA) through utilizing total viral RNA, an enzyme reverse transcriptase, a primer, deoxyribonucleoside triphosphates, and an RNase inhibitor. The synthesized cDNA was quantified by Nanodrop-1000 spectrophotometer (Thermal Scientific, Wilmington, Massachusetts, United States). HIV cDNA of all positive donors was then analyzed for the presence of various HIV genotypes (types and subtypes) by employing subtype-specific primers in a nested polymerase chain reaction (PCR) as described below.

**Polymerase Chain Reaction**

**Standardization of PCR**

The β-globin was used as a control for DNA extraction, and PCR was performed with PC03 and PC04 primers (►Table 1).

**HIV-1 Subtyping**

Nested PCR was done to ensure HIV-1 infection as conditions described by Kato et al,19 to differentiate subtypes A, B, C, and CRF01AE in the env C2V3C3 region.

**env V3 viral DNA Sequences were Amplified by Nested PCR as Follows**

For both first and second round of PCR, the total 25 µL reaction mixture was made.

**First Round of PCR**

The first round of PCR was done using 0.5 µL primer sets JA9AE, JA9B, JA12A, and JA12B. The condition is presented in ►Table 2.

**Second Round of PCR**

Note that 1.5 µL of the amplified product from the first round of PCR was taken to perform the second round of PCR.

**Upstream Primers**

The upstream primers were mixture of three primers (03 µL each). The upstream primers are shown in ►Table 3.

**Downstream Primers**

For subtype-independent amplification, a three-primer mixture was used (0.3 µL). The downstream primers are shown in ►Table 4.

---

**Table 1** β-globin amplification, PCR conditions, and primers used

<table>
<thead>
<tr>
<th>Primer</th>
<th>Sequence</th>
</tr>
</thead>
<tbody>
<tr>
<td>PC03</td>
<td>ACACAACTGTGTTCACTAGC</td>
</tr>
<tr>
<td>PC04</td>
<td>CAACTGCTACACGTTCAACC</td>
</tr>
</tbody>
</table>

**Thermocycling conditions**

| 1× PCR buffer, 1 mM MgCl₂, 200 µM of dNTPs, 0.2 pmol of each primer, 0.2 U of Taq polymerase | 5 min at 95°C |
| 39 cycles: 30 s at 95°C | 30 s at 53°C |
| 30 s at 72°C | 5 min at 72°C |

**Abbreviations:** dNTP, deoxynucleoside triphosphate; HIV, human immunodeficiency virus; PCR, polymerase chain reaction.

**Table 2** HIV-1 subtyping first round PCR conditions and primers used

<table>
<thead>
<tr>
<th>Primer</th>
<th>Sequence</th>
</tr>
</thead>
<tbody>
<tr>
<td>JA9AE</td>
<td>CACAGTACAATGCACACATG</td>
</tr>
<tr>
<td>JA9B</td>
<td>CACAGTACAATGTACACATG</td>
</tr>
<tr>
<td>JA12A</td>
<td>GCAAATAGAAAAATTCTCCTC</td>
</tr>
<tr>
<td>JA12B</td>
<td>ACAGTAGAAAAATTCCCCCTC</td>
</tr>
</tbody>
</table>

**Abbreviations:** dNTP, deoxynucleoside triphosphate; HIV, human immunodeficiency virus; PCR, polymerase chain reaction.

**Table 3** The upstream primers used for HIV-1 subtyping

<table>
<thead>
<tr>
<th>Primer</th>
<th>Sequence</th>
</tr>
</thead>
<tbody>
<tr>
<td>JA10UB</td>
<td>CTGTTAAATGGCAGTCTAGC</td>
</tr>
<tr>
<td>JA10UC</td>
<td>CTGTTAAATGGTAGTCTAGC</td>
</tr>
<tr>
<td>JA10UG</td>
<td>CTGTTAAATGGCAGTCTAGC</td>
</tr>
</tbody>
</table>

**Abbreviation:** HIV, human immunodeficiency virus.

**Table 4** The downstream primers used for HIV-1 subtyping

<table>
<thead>
<tr>
<th>Primer</th>
<th>Sequence</th>
</tr>
</thead>
<tbody>
<tr>
<td>JA11LA</td>
<td>AATTTCTAGATCCCCCTCG</td>
</tr>
<tr>
<td>JA11LB</td>
<td>AATTTCTAGATCCCCCTCG</td>
</tr>
<tr>
<td>JA11LC</td>
<td>AATTTCTAGATCCCCCTCG</td>
</tr>
</tbody>
</table>

**Abbreviation:** HIV, human immunodeficiency virus.
For subtype A: 1 µM of JA11QA was used (CCCCCTCTGAGGAGTACGCA).
For subtype B: 1 µM of JA11VB was used (CACAATTAAACTGTCATTACAA).
For subtype C: 1 µM of JA11XC was used (TTGGTTTATAGGGAAGTGTTC).
For subtype CRF01-AE: 1 µM of JA11YE was used (AAATTCCCCCTCAATTAAATGA).

The thermocycling conditions for second round of PCR are presented in Table 5.

For subtype A: 1 µM of JA11QA was used (CCCCCTCTGAGGAGTACGCA).
For subtype B: 1 µM of JA11VB was used (CACAATTAAACTGTCATTACAA).
For subtype C: 1 µM of JA11XC was used (TTGGTTTATAGGGAAGTGTTC).
For subtype CRF01-AE: 1 µM of JA11YE was used (AAATTCCCCCTCAATTAAATGA).

The thermocycling conditions for second round of PCR are presented in Table 5.

<table>
<thead>
<tr>
<th>Conditions</th>
<th>PCR Cycle</th>
</tr>
</thead>
<tbody>
<tr>
<td>5 min at 95°C; 39 cycles: 20 s at 95°C</td>
<td>30 s at 58°C</td>
</tr>
<tr>
<td>50 s at 72°C</td>
<td>5 min at 72°C</td>
</tr>
</tbody>
</table>

Abbreviation: PCR, polymerase chain reaction.

Results

A total of 85,736 blood donors were screened for the presence of antibodies to HIV during the period 2016 through 2018. Of these donors, 85,199 (99.38%) were males and 537 (0.62%) were females. Out of them, 114 were found initially reactive for HIV. The repeat testing resulted in 112 (0.13%) positive donors, 95% CI 0.0014 (0.0011–0.0018). All positive donors were males.

The number of samples analyzed for HIV-1 genotypic analysis was 112. The technique applied for analysis is the subtype-specific PCR. From the analysis of 112 HIV positive individuals, the predominant HIV-1 subtype was A (n = 101) (90.1%) followed by subtype B (n = 11) (9.9%).

Discussion

HIV epidemic is still ongoing and evolving. The dynamics of the HIV epidemic are diverse in Pakistan and are growing. This is the time to implement effective prevention and care programs to stop the spread of HIV/AIDS. The window of opportunity is the incidence rate of HIV in the general population (<1%) which clearly offers a prospect to act decisively.

Unfortunately, the cases reported so far, only represents the “tip of the iceberg.” The behaviors encouraging the spread of HIV infection to young people include increased interest in sex and drugs, negative peer pressure, and economic frustration in Pakistan. These vulnerabilities and patterns of risky behaviors signal the need to take action now before it is too late. The HIV epidemic is growing at an alarming rate, but unfortunately it is overlooked as the prevalence is mainly confined to high-risk groups who are traditionally marginalized.

Poor awareness about HIV/AIDS with lack of information, persistent scarcity of financial resources, weak regulatory oversight, and common misunderstanding that HIV only infects “individuals with bad characters” are some aspects affecting the occurrence of HIV/AIDS.

The spread of HIV infection via blood transfusions is one of the major concerns and often reports of such cases appear in the print and electronic media. According to blood safety legislation of Pakistan, screening for five markers (called TTIs) is mandatory on every blood unit collected.

The available data from blood donors show a rise in the number of HIV cases detected. The monitoring of HIV patterns in blood donors is critical to ensure the safety of blood and blood components and in addition to sensitizing the decision-makers on the magnitude of the HIV epidemic in blood donors which represent the general healthy population. In developed countries, donor behavioral screening, use of highly sensitive screening tests, and more rational use of blood and blood components have brought about a dramatic decline in the spread of HIV through blood transfusions.

The incidence of HIV reported in the present study (0.13%) was matched with the mean HIV incidence from 1988 to 2016 which was 0.13%. We also compared the findings of our study with those reported by other countries. Our incidence rate of 0.13% was relatively lower than those reported by Northeast Ethiopia 5.1%, Nepal 0.21%, China 0.31%, South Sudan 7.9%, Nigeria 2.8%, Northwest Ethiopia 3.8%, and Burkina Faso 2.21% but higher than those reported from India 0.1%, Iran 0.001%, Italy 0.00019%, and Australia 0.0003%. There is considerable diversity in the incidence of HIV in different countries and indicate variations in the awareness programs, blood screening techniques, and other preventive measures in individual countries.

The blood safety can be enhanced through a comprehensive donor recruitment scheme with standardized behavioral and serological screening. According to previous research studies, the incidence of HIV/AIDS can be reduced to 2 in 1,000,000 or even lesser through behavioral evaluation of blood donor and quality assured serological TTI screening of blood bags.

Molecular epidemiology testing for HIV is a key tool to track patterns of transmission and the evolution of the HIV epidemic in the general public. HIV-1 has a high genetic variability and comprised of four groups (M, N, O, and P). For the HIV
pandemic, M is the major group responsible, being further subclassified into numerous subclasses (A to K, excluding E) and several mosaic strains termed as circulating recombinant forms. Recently, researchers from the United States have also reported a group M subtype in samples from the Democratic Republic of Congo.

The prevalence of HIV-1 subtypes in a population follows the evolution of the epidemic. Hemelaar et al. investigated the country-specific HIV-1 molecular epidemiology data of 65,913 specimens from 109 countries. The analyses of the worldwide pattern of HIV-1 subtypes and circulating recombinant forms specified a largely steady distribution with a noteworthy upsurge in the percentage of circulating recombinant forms, a decline in the unique recombinant forms, and a general rise in recombinants.

Our findings confirmed that “A” subtype is predominant HIV subtype present in our country followed by subtype “B.” This outcome is consistent with earlier available reports showing the subtype A as the most common subtype in Pakistan. In the study by Khan et al., the subtype A was 100% while it was 69.73% as reported by Khanani et al. The study by Chen et al. reported the prevalence of HIV subtype A as 47% with an increasing number of recombinant forms. According to the Los Alamos HIV databases, the national prevalence shows a high peak for subtype A (73.3%). This is in contrast to the global prevalence trends where the prevalent subtype is B (55.8%).

Although this trend may shift during the course of the next few years and more subtypes may become prevalent in the country, HIV is persistently advancing ever since the pioneer case of HIV in Pakistan diagnosed in 1987, and our data suggested that the B subtype was not as common in earlier reports as in our findings. This is largely because of the migrant workers who are infected and introduce new subtypes of HIV. These include laborers and commercial sex workers of bordering countries such as Afghanistan, Iran, or the Gulf States. A significant number of Pakistani nationals are working in the Gulf States including the Kingdom of Saudi Arabia, Kuwait, and the United Arab Emirates. These HIV-infected persons then infect their spouses and kids and ultimately placing the general public at danger. Because of weak regulatory oversight by the government, this practice will lead us to a rapid rise in the incidence of HIV infection and will also alter the pattern of different subtypes in Pakistan.

In conclusion, the results establish HIV-1 molecular complexity among blood donors with subtypes A and B. The findings of this study may be used for further genetic analysis and development of highly advanced assays for blood screening to ensure blood safety. Continuous efforts must be directed toward improving effective treatment and prevention strategies.

Conflict of Interest
The authors declare no potential conflict of interest.

Acknowledgments
We would like to thank Inam Ullah Marwat (IU), Bilal Ahmed Khan Tareen (PIMS), Muhammad Umair (QUA), Saifdar Abbas (QUA), Shahan Irshad (SMS), Muhammad Nisar Khan (PIMS), Mahmood Awan (SMS), and Khawaja Junaid Shokat (SMS) for assistance and support.

References

37 Los Alamos National Laboratory. HIV databases. Available at: https://www.hiv.lanl.gov/content/index. Accessed November 27, 2019