

# Traps N' Clots: NET-Mediated Thrombosis and Related Diseases

Dimitrios Stakos<sup>1,2,\*</sup> Panagiotis Skendros<sup>2,3,\*</sup> Stavros Konstantinides<sup>1,4</sup> Konstantinos Ritis<sup>2,3</sup>

<sup>1</sup> Department of Cardiology, Democritus University of Thrace, Alexandroupolis, Greece

<sup>2</sup> Laboratory of Molecular Hematology, University Hospital of Alexandroupolis, Democritus University of Thrace, Alexandroupolis, Greece

<sup>3</sup> First Department of Internal Medicine, University Hospital of Alexandroupolis, Democritus University of Thrace, Alexandroupolis, Greece

<sup>4</sup> Center for Thrombosis and Hemostasis, University Medical Center of the Johannes Gutenberg University Mainz, Mainz, Germany

Address for correspondence Konstantinos Ritis, MD, PhD, University Hospital of Alexandroupolis, Alexandroupolis 68100, Greece (e-mail: kritis@med.duth.gr).

Thromb Haemost 2020;120:373–383.

## Insights into Neutrophils and NETs: A Historical Perspective

Vessel wall injury and subsequent blood extravasation activates a series of local biological processes to prevent excess blood loss via the formation of hemostatic plug strictly restricted at the site of vascular injury with minimal or no extension in the vessel lumen.<sup>1</sup> In the vast majority of cases, a catastrophic systemic activation of these processes is contained by specific mechanisms. As opposed to hemostasis, thrombosis is characterized by the deregulated clot formation, various degrees of vessel occlusion, tissue ischemia, and necrosis.<sup>1</sup>

A large body of accumulating experimental and clinical data over the past 20 years has clearly indicated the reciprocal relationship and dynamic interplay between inflammation and thrombosis.<sup>2–5</sup> Today, targeting inflammation to prevent thrombotic events represents a realistic and promising therapeutic approach.<sup>6</sup> Among immune cell subsets that are implicated in multiple molecular pathways during inflammatory response, neutrophils have a crucial role, recruited first to the site of injury following instructive signals from the tissue environment.<sup>7</sup>

The very first observation linking neutrophils with thromboinflammation was reported almost 70 years ago describing granulocytes as a main component of clotted blood in patients suffering from active lupus erythematosus.<sup>8</sup> During the following years, although several studies had described the accumulation of neutrophils at the site of thrombus formation, these cells remained neglected and less studied in many throm-

botic diseases. The traditional aspect of neutrophils as dispensable, passive bystanders was dramatically revised after the milestone discovery that they represent a primary source of blood-borne tissue factor (TF), the main *in vivo* initiator of the extrinsic coagulation cascade, resulting in thrombin generation and ensuing thrombus formation.<sup>9,10</sup> Later on, several studies from our laboratory and others provided evidence for the critical role of neutrophils in thrombosis and inflammation-mediated thrombotic complications.<sup>3,11,12</sup> Intravital microscopy studies in mouse models of venous and arterial thrombosis demonstrated neutrophil recruitment and activation at the site of endothelial damage in the early phase of thrombosis.<sup>13,14</sup> Of note, neutrophils are not only implicated in thrombotic processes, but also seem to be indispensable for thrombosis. Neutropenia induced *in vivo* by anti-Ly6G or GR-1 antibody abrogated venous<sup>13</sup> and arterial<sup>14</sup> thrombosis, respectively. When purified neutrophils from wild-type mice were injected into transgenic mice that express no mouse TF and only minimal (< 1%) amounts of human TF (low TF mice), the defective fibrin generation in these animals was restored, indicating that TF expressing neutrophils represent the main source of TF during thrombus formation.<sup>14</sup> Similarly, when normal TF expressing mice were transplanted with low-human TF bone marrow cells they did not develop deep vein thrombosis (DVT).<sup>13</sup> However, the mechanisms underlying the activation and delivery of active TF by neutrophils remained unknown.

During the last years, advances in molecular biology provided the most exciting update of neutrophil physiology, in particular their capacity to release neutrophil extracellular traps (NETs). NETs are extracellular web-like structures of chromatin fibers lined with various highly active proteases

\* These authors contributed equally to this work.

received

July 1, 2019

accepted after revision

November 26, 2019

© 2020 Georg Thieme Verlag KG  
Stuttgart · New York

DOI <https://doi.org/10.1055/s-0039-3402731>.  
ISSN 0340-6245.

and proteins of nuclear, granular, and cytosolic origin.<sup>15</sup> The release of NETs from activated neutrophils was initially described in 2004 as a novel defense mechanism able to “trap and kill” a wide range of pathogens.<sup>16</sup> However, increasing evidence during the past few years highlighted their fundamental role in the pathogenesis of numerous noninfectious inflammatory disorders, such as systemic lupus erythematosus (SLE), rheumatoid arthritis (RA), autoimmune vasculitis, gout, ulcerative colitis, interleukin (IL)-1 $\beta$ -mediated autoinflammatory syndromes, and thrombosis.<sup>15,17–19</sup>

Activated platelets are able to induce robust NET release within vasculature providing a scaffold for fibrin deposition and stabilization of thrombus.<sup>20–23</sup> Notably, these NETs are decorated with functionally active TF, which explains its extracellular delivery at the site of tissue injury.<sup>13,19,24</sup> Besides TF, NETs were found to deliver several proteins and clot factors involved in thrombosis such as von Willebrand factor (vWF), XII, fibrinogen, and fibronectin.<sup>2,13,25</sup> The thrombogenic potential of NETs was further supported by experimental studies indicating that extracellular histones induce endothelial activation, platelet activation/aggregation, and thrombin generation<sup>26,27</sup> (→Fig. 1). Phylogenetically, the capacity of NETs to activate coagulation serves to trap and eliminate pathogens, resembling primitive defense systems operating several millions years ago, and it is conserved today in insects.<sup>28</sup> In these organisms, coagulation and immunity use common mechanisms to prevent fluid loss and pathogen invasion. These systems are tightly interrelated, since NET-associated antimicrobial proteases are able to trigger several coagulation pathways,<sup>2</sup> while activation of the coagulation system supports several immune responses such as bacterial compartmentalization, immobilization, and elimination especially in microvasculature (immunothrombosis). Probably, much of the NET-mediated antimicrobial effect is due to entrapment, rather than direct killing<sup>29,30</sup> (→Fig. 2).

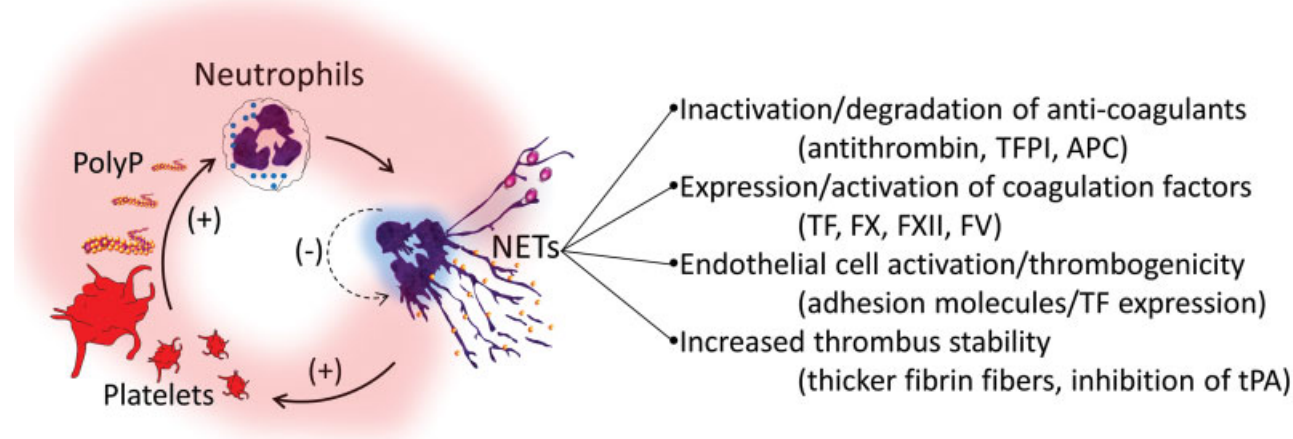
Apart from neutrophils, extracellular traps (ETs) formation has also been described in other types of granulocytes, such as eosinophils and mast cells.<sup>31,32</sup> Very recently data implicate macrophages, mast cells, and eosinophils through

ETs generation in atherosclerotic plaque formation and thrombosis.<sup>33,34</sup> However, ETs formation in macrophages is controversial and remains unclear whether it is distinct from pyroptosis.<sup>35</sup>

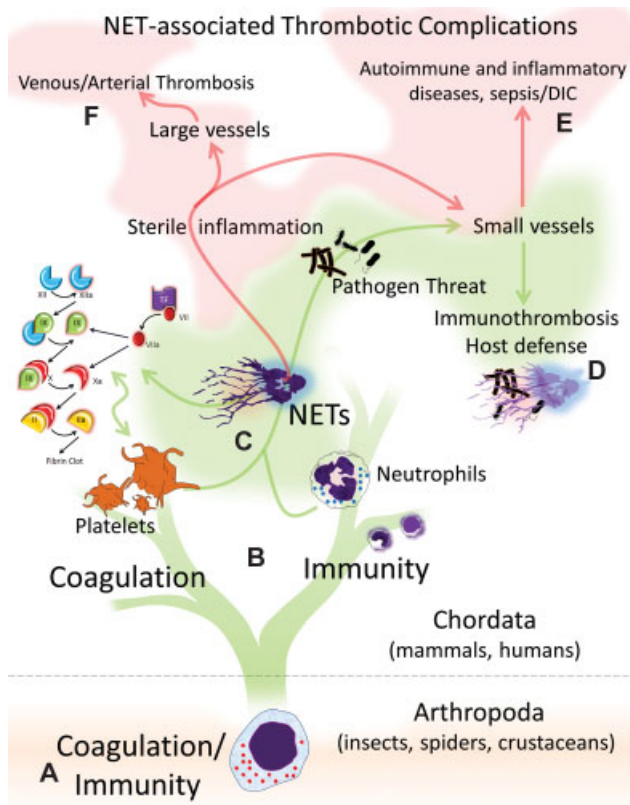
In view of the above, NETs could be perceived as a double-edged sword during disease processes. They may be beneficial by enhancing the antimicrobial potential in numerous infectious diseases or contributing to normal hemostasis and pathogen curbing in neutrophil clots lattice, but also harmful by amplifying systemic or local inflammation leading to tissue damage and thrombosis<sup>15,36</sup> (→Fig. 2). Therefore, NETs and their components emerge today as novel candidate for diagnostic and therapeutic targets of thrombosis in many clinical settings.<sup>12,15,37,38</sup>

## NET Formation and Regulation

The exact molecular mechanisms that drive NET release are not clearly defined and still being characterized.<sup>35</sup> Several lines of evidence indicated that NET formation is regulated via multiple, probably interdependent, pathways that can be triggered in vivo by several microbial and noninfectious stimuli, such as cytokines, chemokines, immune complexes, crystals, and inorganic polyphosphate (polyP) or high mobility group box 1 (HMGB1) expressed from activated platelets.<sup>22,24,35,39–41</sup> Stimuli of NET formation may act also synergistically. Thus, it has been suggested, that platelet-induced NET formation requires synchronous neutrophil stimulation by platelet-derived chemokines CXCL4 and CCL5, and Mac-1 integrin.<sup>23</sup> On the other hand, excessive and disproportionate NET formation that leads to tissue damage is homeostatically regulated by anti-inflammatory mechanisms such as NET degradation by DNase1 and DNase1-like 3, and phagocytic removal of NET remnants by macrophages.<sup>18,42</sup> Interestingly, recent data suggested that an anti-inflammatory action of NETs themselves also exists mainly through the proteolytic modulation of cytokine and chemokine activity by the proteases that decorate aggregated NETs.<sup>43–45</sup>



**Fig. 1** Mechanisms of neutrophil extracellular trap (NET) thrombogenicity. Left: Platelets-PolyP-neutrophils-NETs interaction leads to NET generation. (–) denotes NET autoregulation. Right: NETs can deliver thrombogenic signals through many different mechanisms presented here. APC, activated protein C; FX, FXII, FV, coagulation factors; PolyP: polyphosphate; TF, tissue factor; TFPI, TF pathway inhibitor; tPA, tissue plasminogen activator.



**Fig. 2** Skewing of immunothrombosis (tissue protection) to pathological thrombosis (tissue damage). In primitive organisms (bottom, A) immunity and coagulation use a common, hemocyte-based system to prevent fluid loss and pathogen invasion. In mammals including humans (B) hemostasis and immunity are operated by distinct systems (platelets and coagulation, left; and white blood cells such as neutrophils, right, respectively). Interaction between platelets/coagulation and neutrophils leads to neutrophil extracellular trap (NET) formation (C). In cases of a pathogen invasion, NETs are involved in pathogen entrapment and elimination in microcirculation (green path, host defense, D). On the contrary, inappropriate NET formation in cases of sterile inflammation (autoimmune or inflammatory environment) leads to thrombotic complications (red path, tissue damage) in microcirculation (E) or large vessels (F). DIC, disseminated intravascular coagulation.

Recent clinical and experimental studies suggested that, in the context of different diseases, neutrophils are able to release NETs that are qualitatively different and coated with disease-specific bioactive proteins determined by the disease inflammatory environment such as TF, IL-1 $\beta$ , IL-17, and LL-37.<sup>17,19,46–52</sup> It has been suggested that the systemic inflammatory environment of each disease leads to transcriptional reprogramming in circulating neutrophils inducing the de novo expression of disease-related proteins (first-hit), and an additional trigger (second-hit) enables NET formation leading to the extracellular delivery of these bioactive proteins via NETs.<sup>19,49</sup>

## NETs and Local Thrombotic Manifestations

### Experimental Arterial Thrombosis

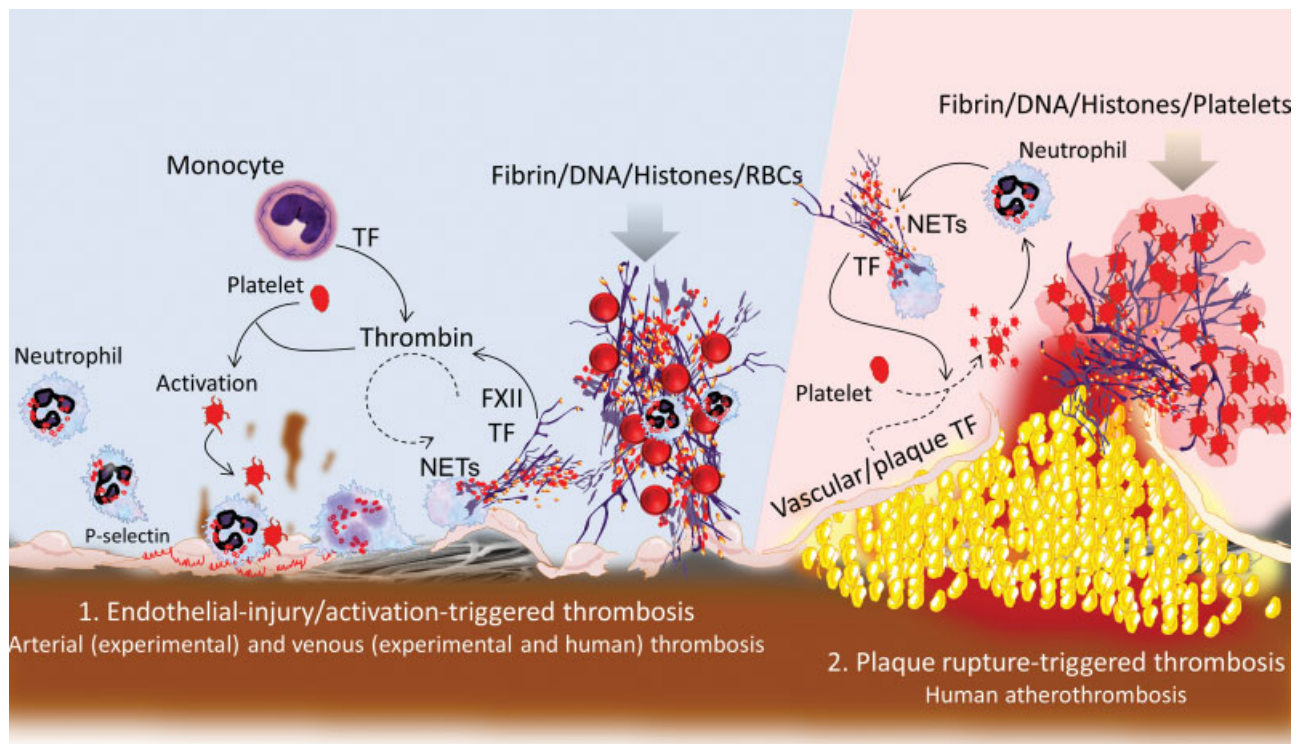
In mice, following endothelial injury and activation, neutrophils are recruited first and promote thrombosis by forming

NETs and by delivering active TF and neutrophil proteases such as elastase (NE) and cathepsin G (CG).<sup>2,14,53</sup> In mice lacking NE and CG (Elane<sup>-/-</sup>; Ctsg<sup>-/-</sup>), FeCl<sub>3</sub> carotid injury resulted in lower fibrin formation, smaller and unstable thrombi leading to rapid recanalization, possibly due to defective TF pathway inhibitor degradation from NET-bound NE and CG.<sup>2</sup> Extracellular histones, an essential NET component, also aggravated ischemic injury in a mouse model of transient occlusion and reperfusion of middle cerebral artery when injected during the reperfusion phase, resulting in larger infarcts and worse functional outcomes. Significant attenuation of these effects was observed with administration of histone-neutralizing antibody or recombinant human DNase1.<sup>54</sup> NETs also operate in myocardial reperfusion injury. In a left anterior descending artery ligation and reperfusion model in rat, treatment with DNase1 and recombinant tissue plasminogen activator (tPA) reduced ischemic injury, no reflow and infarct size, and improved long-term left ventricular remodeling and systolic function compared with control and to either treatment alone.<sup>55</sup> Recently, in models of experimental atherosclerosis it has been also demonstrated that NET-derived extracellular histone H4 induces smooth muscle cells (SMCs) lysis leading to plaque destabilization, while neutralization of histone H4 prevents cell death of SMCs and stabilizes atherosclerotic lesions.<sup>56</sup>

### Human Arterial Thrombosis

Atherothrombosis is triggered by strong platelet activation following plaque erosion or rupture end exposure of thrombogenic material to the blood stream. Initial platelet-rich thrombus is potentiated by activation of blood coagulation through vascular and blood-borne TF (**Fig. 3**). Neutrophils and NETs have been implicated in coronary thrombosis induced by endothelial activation,<sup>57</sup> plaque erosion,<sup>58,59</sup> and rupture.<sup>33,60</sup> Neutrophils and NETs are abundantly present in thrombi derived from patients with ST-elevation myocardial infarction (STEMI)<sup>19,22,61</sup> and thrombosed stents.<sup>62</sup> Coronary thrombus-resident NETs express IL-17A/F and TF, both of which promote platelet and coagulation activation.<sup>19,63,64</sup> Mechanistically, TF-bearing NETs are induced by polyP secreted from thrombin-activated platelets, while IL-29/interferon  $\lambda$ 1 attenuates NET formation by inhibiting polyP/mammalian target of rapamycin (mTOR)/autophagy pathway.<sup>24</sup>

The presence of NETs was associated with more stable coronary thrombi, sustained ischemia, and increased enzymatic and cardiovascular magnetic resonance-measured infarct size in 111 patients with STEMI.<sup>61</sup> In a prospective, cross-sectional study of 282 individuals with clinically suspected coronary artery disease, plasma markers of NETosis citH4 and myeloperoxidase (MPO)-deoxyribonucleic acid (DNA) complexes, were positively associated with increase thrombin-antithrombin complex and vWF levels as well as revascularization, acute coronary syndrome, and cardiac death after a median follow-up period of 545 days.<sup>65</sup> Significantly elevated concentrations of DNA, nucleosomes, and citH3 as markers of enhanced NETotic burden were also measured in stroke patients.<sup>66–68</sup> In patients with acute ischemic stroke, NETotic markers



**Fig. 3** Left (1): In case of endothelial damage/activation, neutrophils adhere through P-selectin and release neutrophil extracellular traps (NETs) upon interaction with neighboring activated platelets. NETs support tissue factor (TF) and FXII activation and thrombin generation (together with TF derived by monocytes). The result is the formation of a thrombus composed of fibrin, deoxyribonucleic acid (DNA), histones, red blood cells (RBCs), and some platelets. Right (2): Following plaque rupture, vascular TF activates locally recruited platelets which interact with TF-loaded neutrophils to release TF-bearing NETs. Active NET-bound TF supports thrombin generation which further activates more platelets in a vicious cycle. The result is a robust platelet activation and the formation of a thrombus composed of fibrin, DNA, histones, and platelets.

were positively correlated with clot stability and resistance to endovascular therapy,<sup>69</sup> and with stroke severity scores and all-cause mortality at 1 year.<sup>68</sup> MPO, cell-free DNA, and MPO-DNA complexes were also detected in intraplaque hemorrhage segments of carotid atherosclerosis specimens,<sup>70</sup> and in intraluminal thrombi derived from patients with abdominal aortic aneurysms.<sup>53,71</sup>

### Experimental and Human venous Thrombosis

A massive platelet aggregation is not essential for venous thrombosis (→Fig. 3). Low shear venous thrombosis is triggered by stasis-induced endothelial activation, rapid neutrophil recruitment, and adhesion via endothelial P-selectin. Both neutrophils and NETs were found in large amounts in venous thrombi in a mouse model of inferior vena cava flow restriction-induced thrombosis. Adherent neutrophils interact with platelets locally and expose thrombogenic elements such as TF and FXII through NET release.<sup>13,72</sup> Elevated NETosis markers in plasma were measured in experimental iliac vein thrombosis in baboons (extracellular DNA) and humans (circulating nucleosomes and NE/ $\alpha$ 1-antitrypsin complexes).<sup>27,73–75</sup> In a case-control study with 345 participants, high levels of NETotic markers increased the risk of DVT by threefold even after adjustment for known confounders (malignancy, smoking, recent immobilization, recent hospitalization) in a dose-dependent manner.<sup>73</sup>

### Thrombus Stabilization and Lysis

Beyond their contribution to thrombus propagation, extracellular DNA and NETs affect also thrombus structure and stability with clinical implications related to thrombus lysis. Inhibition of leukocyte infiltration in a baboon model of DVT resulted in unstable thrombi suggesting a role for neutrophils/NETs in thrombus stabilization.<sup>76</sup> NETs can directly degrade fibrin via NET-bound NE and CG while histone H2B can serve as a receptor for plasminogen, recruiting plasminogen from the plasma.<sup>77</sup> However, neutralization of NET-associated proteases by plasma antiproteases *in vivo* results in a net inhibitory effect of NETs on clot lysis in the blood.<sup>78</sup> Addition of DNA, histones, or both, on clotting plasma, resulted in thicker fibrin fibers, while NETs incorporated into plasma clots significantly delayed clot lysis *in vitro* by reducing tPA activity. In quantitative terms, the addition of histone and DNA to clots produced 50% thicker fibers and doubled the critical shear stress for loss of fibrin viscosity.<sup>79</sup> Moreover, we have observed absence of NETotic neutrophils and NETs in thrombus remnants aspirated from STEMI patients with spontaneous thrombus resolution.<sup>19</sup> These observations suggest that the formation of NETs is imperative for thrombus stability and lytic resistance.

The characteristic of chronic thromboembolic pulmonary hypertension (CTEPH) is extreme persistence and lytic resistance of pulmonary thromboemboli leading to chronic obstruction of pulmonary arteries and in small vessel arteriopathy.<sup>80</sup>

Since the presence of NETs and DNA has been associated with thrombus stability, it is possible that NETotic infiltration of pulmonary thrombi (inflamed thrombi) causes resistance to endogenous lytic mechanism and thrombus persistence. Indeed, in two independent cohorts of CTEPH patients a 10-fold increase of MPO plasma levels, 7-fold increase in plasma NE, and more than 2-fold increase in circulating MPO/DNA levels compared with healthy volunteers were measured.<sup>81</sup> Moreover, analysis of endarterectomy specimens obtained from several independent cohorts of CTEPH patients consistently revealed abundant CitH3-positive cells, protein arginine deiminase-4 (PAD4)+ NETotic neutrophils, MPO/DNA colocalization, and NETs in the organizing stage of thrombus development and maturation.<sup>75,81</sup>

## NETs and Systemic Inflammation-Mediated Thrombosis

Apart from locally occurring thrombosis in the context of cardiovascular diseases, several lines of evidence demonstrate that NETs mediate thrombosis in various systemic thromboinflammatory conditions, such as sepsis, autoimmunity, and malignancy. These disorders are characterized by the presence of activated neutrophils and NETs fragments in the bloodstream and affected tissues, and a high propensity for serious, life-threatening thrombotic events.

### Autoimmune and Inflammatory Diseases

It is well established that NET formation is implicated in the induction of autoimmunity and correlated with the severity of related diseases.<sup>17,38,82</sup> Accordingly, NET-dependent coagulation has been suggested as a common mechanism that might explain the increased incidence of thromboembolic events in patients suffering from systemic inflammatory diseases, especially during periods of exacerbation.<sup>11,12</sup>

The key role of NETs in the pathogenesis of autoimmune-related thrombosis was first demonstrated in antineutrophil cytoplasmic antibodies (ANCA)-associated vasculitis (AAV). Excessive NETosis has been observed in pulmonary and venous thrombi from autopsy specimens in severe AVV.<sup>83,84</sup> In line with these, TF-bearing NETs were released by circulating and bronchoalveolar lavage neutrophils, and were found in nasal and renal biopsies of active AVV patients. Additionally, *in vitro* stimulation of neutrophils with serum or ANCA derived from these patients induced the expression of TF in neutrophils and its extracellular delivery via NETs. Interestingly, disease remission or ANCA depletion attenuated these effects suggesting the pathogenic role of ANCA in AVV.<sup>85,86</sup> The presence of thrombogenic TF on NETs has also been demonstrated in peripheral blood and inflamed tissues from active SLE and ulcerative colitis patients in a suggestive mechanism that is triggered by disease inflammatory microenvironment and is regulated by mTOR/autophagy pathway.<sup>48,52</sup> Besides thrombosis, signaling through protease-activated receptors that results in the activation of endothelial, epithelial, and other tissue cells could be an additional pathogenic pathway for the involvement of neutrophils/NETs-expressed TF in organ damage of several chronic inflammatory diseases.<sup>87,88</sup>

Thirteen years after the discovery that purified immunoglobulin G from patients with antiphospholipid antibody syndrome (APS) is able to induce TF expression in human neutrophils,<sup>9</sup> recent experimental and clinical data demonstrate that NET formation and regulation constitutes a new pathogenic mechanism in typical (auto)immune-mediated thrombotic diseases associated with a severe prothrombotic state, such as APS, heparin-induced thrombocytopenia/thrombosis, and thrombotic thrombocytopenic purpura, further supporting the crucial role of neutrophils in immunothrombosis.<sup>89–94</sup>

Apart from TF, the contribution of histones and other neutrophils/NETs-derived components, such as phosphatidylserine positive microparticles (MPs) or autoantigens, in the hypercoagulability of clinical active autoimmune/inflammatory diseases have also been recognized.<sup>85,95,96</sup> Recently, the role of platelet/neutrophil interaction in autoimmune vascular damage is further reinforced. HMGB1 on platelets-derived MPs was found to be key inducer of autophagy-mediated NET release in patients suffering from systemic sclerosis suggesting that neutrophil autophagy and NETs are tightly implicated in disease vasculopathy and microthrombosis.<sup>97</sup>

Besides their direct prothrombotic action, NETs are major contributors to endothelial dysfunction and atherosclerotic plaque formation.<sup>98</sup> NET formation provides a reasonable pathophysiological link between inflammation and atherothrombosis in patients with chronic rheumatic diseases such as SLE and RA.<sup>99–101</sup> Consistent with this, blocking of NET formation using PAD4 inhibition in a murine model of lupus that is characterized by accelerated vascular damage and prothrombotic tendency, significantly improved endothelial dysfunction and thrombosis risk supporting an important role for NETs in the atherosclerosis and hypercoagulability of systemic autoimmunity.<sup>102</sup>

Taken together, functionally diverse neutrophil subsets are apparently attractive targets for “omics” studies that may/will unveil novel anti-inflammatory strategies in several devastating, difficult-to-treat autoimmune thromboinflammatory conditions.<sup>103–106</sup> For example, RNAseq analysis of whole blood from patients with AAV has previously linked low-density granulocytes (LDGs), a population functionally characterized by high propensity to form NETs, with disease activity and decreased response to treatment. This probably implies that more aggressive treatment and close follow-up are needed in AAV patients with increased frequency of LDGs.<sup>103</sup> Similarly, transcriptome analysis of neutrophils revealed P-selectin glycoprotein ligand-1 (PSGL-1) adhesion molecule as a potential therapeutic target in primary APS. Interestingly, PSGL-1 deficient mice were characterized by reduced NET formation.<sup>104</sup> Moreover, genome-wide DNA methylation analysis of neutrophils showed an entirely distinct DNA methylation profile between primary APS and SLE patients.<sup>105</sup> Whether this heterogeneity is related with NETosis or/and therapeutic response in different phenotypes of APS remains to be studied.

### NETs and Cancer-Associated Thrombosis

Malignancy is characterized by neutrophil accumulation in tumor environment and high thrombotic risk, whereas thrombosis is a main cause of morbidity and mortality in

**Table 1** Important findings on NET-associated thrombosis in different diseases bearing therapeutic potential

Disease		Reference
STEMI	NETs abundance in coronary thrombi of patients NETs are present in coronary stent thrombus NETs promote thrombus organization Platelet-neutrophil interaction through polyP triggers NET generation Functionally active TF on NETs of culprit coronary artery	19,22,61 62 63 24 19
Stroke	NETs in thrombi are positively correlated with resistance to endovascular therapy	69
DVT/PE	TF positive neutrophils adhere to activated vein endothelium in vivo NETs promote thrombus organization and maturation Activated neutrophils and plasma nucleosomes/DNA as DVT risk factors NETotic neutrophils in endarterectomy specimens from VTE patients	13 72,75 73,74 75
CTEPH	NETs activate pulmonary arterial endothelial cells and proliferation of pulmonary smooth muscle cells NETs in plasma and intrapulmonary thrombi of patients	81 81
AAV	LDGs may be associated with disease activity and response to treatment NETs in pulmonary and venous thrombi in severe disease Functionally active TF on peripheral and tissue NETs	103 83,84 85,86
SLE	Functionally active TF on peripheral and tissue NETs REDD1/mTOR/autophagy pathway is related to thrombogenic NETs	52 52
APS	Decreased degradation of NETs Antiphospholipid antibodies induce NETs Neutrophil PSGL-1 as a putative therapeutic target Adenosine receptor agonism protects against NETosis and thrombosis	89 90 104 106
HIT	Neutrophil activation leads to NETs-induced thrombosis	92,93
TTP	Impaired DNase1-mediated degradation of NETs	94
UC	NETs enhance procoagulant activity Functionally active TF on peripheral and intestinal NETs REDD1/mTOR/autophagy pathway is related to thrombogenic NETs	48,96 48 48
Sepsis	Functionally active TF on peripheral NETs NETs are associated with DIC, VTE, and impaired fibrinolysis	47 123-127
Cancer	NETs as prediction marker of cancer-associated VTE Tumor environment primes neutrophils to release procoagulant NETs NETs-bound TF in colon cancer and metastatic lymph nodes NETs promote cancer-associated venous and arterial thrombosis	37 109,111 118 115-117 119,120

Abbreviations: AAV, anti-neutrophil cytoplasmic antibodies (ANCA)-associated vasculitis; APS, antiphospholipid antibody syndrome; CTEPH, chronic thromboembolic pulmonary hypertension; DIC, disseminated intravascular coagulation; DNA, deoxyribonucleic acid; DVT/PE, deep venous thrombosis/pulmonary embolism; HIT, heparin-induced thrombocytopenia/thrombosis; LDGs, low-density granulocytes; mTOR, mammalian target of rapamycin; NET, neutrophil extracellular trap; PSGL-1, P-selectin glycoprotein ligand-1; REDD1, regulated in development and DNA damage responses 1; STEMI, ST-elevation myocardial infarction; TF, tissue factor; TTP, thrombotic thrombocytopenic purpura; UC, ulcerative colitis; VTE, venous thromboembolism.

cancer patients.<sup>107,108</sup> First, Demers et al using various animal models of cancer showed an increase in neutrophils count and their propensity for NET formation in combination with the appearance of venous thrombi in tumor affected tissues.<sup>109</sup> This effect is probably through a systemic effect on the host that involves tumoral granulocyte colony-stimulating factor and other tumor microenvironment such as IL-8 and exosomes.<sup>109-111</sup>

Over the last years, many studies in tumor experimental models and patients suffering from hematological or solid malignancies strongly implicate NETs in cancer-associated venous and arterial thrombosis, as well as tumor growth and metastasis.<sup>112-117</sup> NETs were found in colon tumor biopsies and the respective metastatic lymph nodes representing a prominent source of TF in cancer microenvironment.<sup>118</sup> Mechanistically, intratumor NET-mediated TF expression may promote tumor thrombosis and necrosis, or contrariwise, may enhance neoangiogenesis offering meta-

bolic support to rapidly growing malignant cells. Furthermore, tumor-associated NETs could be an important source of TF that contributes to systemic thrombosis. Tumor cells can also promote platelet aggregation by stimulating endothelial production of vWF, another coagulation factor that is able to bind NET scaffold contributing to thrombosis.<sup>119</sup> In another experimental study, tumorigenesis was associated with coagulation induced by low-density neutrophils that displayed spontaneous NET formation in a complement-dependent manner.<sup>120</sup> Consistent with the abovementioned data, in a prospective observational study that comprises almost 1,000 patients with newly diagnosed cancer or cancer progression after remission, levels of CitH3, a NET formation biomarker, are independently associated with the occurrence of venous thromboembolism (VTE).<sup>37</sup> The same group has currently reported that NET biomarkers in patients with cancer were not associated with arterial thromboembolism, but with higher mortality.<sup>121</sup>

### Systemic Inflammatory Response Syndrome

Besides noninfectious inflammatory conditions, systemic inflammatory response syndrome in the context of severe infection/sepsis is able to trigger excessive NET release in the vasculature causing endothelial tissue damage, intravascular coagulation, and organ dysfunction.<sup>18,122–124</sup> Septic patients spontaneously release NETs carrying functionally active TF in an autophagy-dependent manner, whereas the inflammatory environment of sepsis induces the *in vitro* formation of thrombogenic TF-bearing NETs.<sup>47</sup> In line with this, several studies have indicated that increased levels of NET releasing in septic patients were significantly correlated with disseminated intravascular coagulation development, VTE risk, and impaired fibrinolysis.<sup>123,125–127</sup>

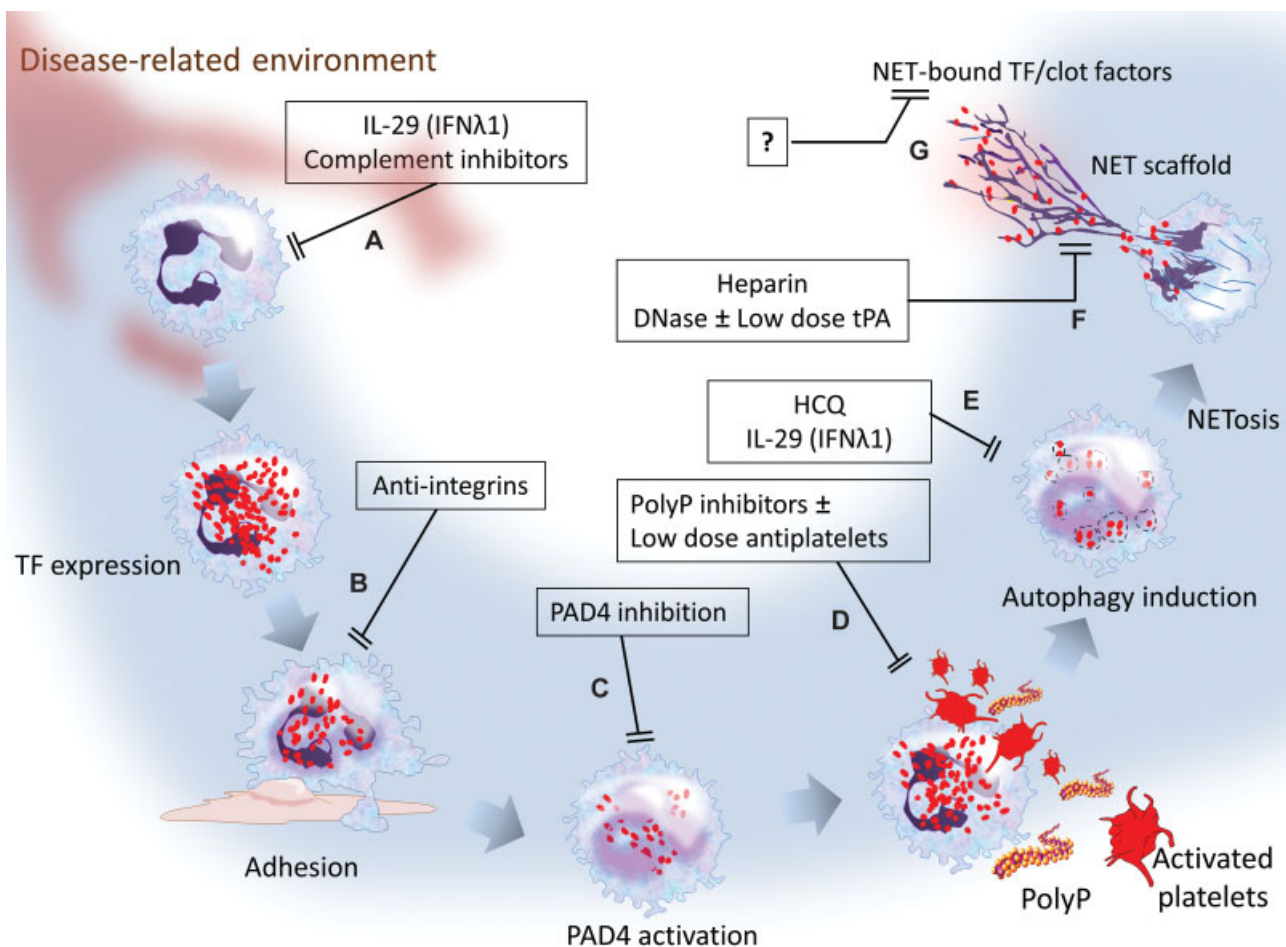
### Translational Impact and Future Perspectives

Although effective antithrombotic approaches have been developed to combat this cardiovascular epidemic, current regimens target both pathological thrombosis and physiological hemostasis, and therefore almost inevitably increase

the risk of bleeding. Therapeutic approaches aimed at inflammatory mechanisms of thrombosis induction rather than directly in the coagulation system may be used in the future alternatively or synergistically with conventional treatments, enhancing therapeutic benefit and potentially reducing the risk of hemorrhagic complications.

The above described findings regarding the important role of NETs in the generation and stabilization of thrombus, offers the potential for new therapeutic options targeting neutrophil-driven thromboinflammation in several diseases (–Table 1). For example, therapies against: (1) inductive/regulatory mechanisms of NET formation (e.g., complement, autophagy, anti-polyP), (2) NETs themselves (chromatin scaffold integrity, histones), and (3) specific NET-bound proteins (e.g., TF/clot factors, IL-1 $\beta$ /cytokines), without affecting NET release per se (–Fig. 4), may prove more effective and/or safer antithrombotic agents provided that they will not interfere with normal hemostasis or immunocompetence.<sup>24,128–136</sup>

Taken in consideration the experimental and human data obtained since today, and without ignoring traditional risk factors and classical antithrombotic regimens, clinical trials that will investigate the anti-NETotic effect on thrombotic



**Fig. 4** Possible targets of neutrophil/neutrophil extracellular trap (NET)-driven thromboinflammation. Inhibition of the intracellular expression of disease-related proteins in circulating neutrophils (A); inhibition of adhesion and early NETotic steps (B, C); inhibition of late NETotic steps and autophagy (D, E); dismasting of NET scaffold (F); blocking of specific NET-associated proteins (G). HCQ, hydroxychloroquine; IFNλ, interferon-λ; IL-29, interleukin-29; PAD4, protein arginine deiminase-4; PolyP, inorganic polyphosphate; tPA, tissue plasminogen activator.

diseases are needed for the development of novel therapeutics. Recently, the CANTOS (Canakinumab Anti-Inflammatory Thrombosis Outcome Study) trial demonstrated that anti-inflammatory therapy targeting IL-1 $\beta$  significantly reduced cardiovascular events in patients with previous myocardial infarction and residual inflammatory risk, without affecting lipid levels. This large randomized, double-blind trial, represents the first "proof-of-principle" of the inflammation hypothesis of atherothrombosis paving the way for further research into this field.<sup>6</sup> In a similar way, large international trial HIBISCUS has been launched to examine the efficacy of hydroxychloroquine (HCQ) in the secondary prevention of thrombotic events of primary APS.<sup>131</sup> Interestingly, HCQ inhibits autophagy<sup>129</sup> and several human studies have linked IL-1 $\beta$  and autophagy with NET formation and activity.<sup>22,24,43,47-49,52,97,132,137</sup>

Further investigation of the neutrophil/NET/coagulation axis is expected to unveil novel candidate diagnostic, prognostic, and therapeutic targets in inflammatory and thrombotic conditions, but also to redefine the clinical indications of old drugs through repositioning.<sup>12,37,68,121,130-132</sup> In parallel, new, NETosis-based, coagulation assays are on the way.<sup>138,139</sup>

#### Funding

This work was supported by Externally Sponsored Scientific Research Grant number ESR-16-12098 from AstraZeneca, and Research Grant number 80895 from the Scientific Committee of Democritus University of Thrace.

#### Conflict of Interest

D.S. reports grants from Externally Sponsored Scientific Research Grant ESR-16-12098 from AstraZeneca, outside the submitted work. S.K. reports grants and personal fees from null, grants and personal fees from null, personal fees from null, grants and personal fees from null, grants from null, personal fees from null, personal fees from null, outside the submitted work.

#### References

- Jackson SP. Arterial thrombosis—insidious, unpredictable and deadly. *Nat Med* 2011;17(11):1423–1436
- Massberg S, Grahnl L, von Bruehl M-L, et al. Reciprocal coupling of coagulation and innate immunity via neutrophil serine proteases. *Nat Med* 2010;16(08):887–896
- Kambas K, Mitroulis I, Ritis K. The emerging role of neutrophils in thrombosis—the journey of TF through NETs. *Front Immunol* 2012;3:385
- Jackson SP, Darbousset R, Schoenwaelder SM. Thromboinflammation: challenges of therapeutically targeting coagulation and other host defense mechanisms. *Blood* 2019;133(09):906–918
- Burzynski LC, Humphry M, Pyriellou K, et al. The coagulation and immune systems are directly linked through the activation of interleukin-1 $\alpha$  by thrombin. *Immunity* 2019;50(04):1033–1042
- Ridker PM, Everett BM, Thuren T, et al; CANTOS Trial Group. Antiinflammatory therapy with canakinumab for atherosclerotic disease. *N Engl J Med* 2017;377(12):1119–1131
- Mócsai A. Diverse novel functions of neutrophils in immunity, inflammation, and beyond. *J Exp Med* 2013;210(07):1283–1299
- Gonyea LM, Kallsen RA, Marlow AA. The occurrence of the "L. E." cell in clotted blood. *J Invest Dermatol* 1950;15(01):11–12
- Ritis K, Dumas M, Mastellos D, et al. A novel C5a receptor-tissue factor cross-talk in neutrophils links innate immunity to coagulation pathways. *J Immunol* 2006;177(07):4794–4802
- Maugeri N, Brambilla M, Camera M, et al. Human polymorphonuclear leukocytes produce and express functional tissue factor upon stimulation. *J Thromb Haemost* 2006;4(06):1323–1330
- Kapoor S, Opneja A, Nayak L. The role of neutrophils in thrombosis. *Thromb Res* 2018;170:87–96
- Bonaventura A, Montecucco F, Dallegri F, et al. Novel findings in neutrophil biology and their impact on cardiovascular disease. *Cardiovasc Res* 2019;115(08):1266–1285
- von Brühl M-L, Stark K, Steinhart A, et al. Monocytes, neutrophils, and platelets cooperate to initiate and propagate venous thrombosis in mice in vivo. *J Exp Med* 2012;209(04):819–835
- Darbousset R, Thomas GM, Mezouar S, et al. Tissue factor-positive neutrophils bind to injured endothelial wall and initiate thrombus formation. *Blood* 2012;120(10):2133–2143
- Mitsios A, Arampatzioglou A, Arelaki S, Mitroulis I, Ritis K. NETopathies? Unraveling the dark side of old diseases through neutrophils. *Front Immunol* 2017;7:678
- Brinkmann V, Reichard U, Goosmann C, et al. Neutrophil extracellular traps kill bacteria. *Science* 2004;303(5663):1532–1535
- Jorch SK, Kubes P. An emerging role for neutrophil extracellular traps in noninfectious disease. *Nat Med* 2017;23(03):279–287
- Jiménez-Alcázar M, Rangaswamy C, Panda R, et al. Host DNases prevent vascular occlusion by neutrophil extracellular traps. *Science* 2017;358(6367):1202–1206
- Stakos DA, Kambas K, Konstantinidis T, et al. Expression of functional tissue factor by neutrophil extracellular traps in culprit artery of acute myocardial infarction. *Eur Heart J* 2015;36(22):1405–1414
- Caudrillier A, Kessenbrock K, Gilliss BM, et al. Platelets induce neutrophil extracellular traps in transfusion-related acute lung injury. *J Clin Invest* 2012;122(07):2661–2671
- Clark SR, Ma AC, Tavener SA, et al. Platelet TLR4 activates neutrophil extracellular traps to ensnare bacteria in septic blood. *Nat Med* 2007;13(04):463–469
- Maugeri N, Campana L, Gavina M, et al. Activated platelets present high mobility group box 1 to neutrophils, inducing autophagy and promoting the extrusion of neutrophil extracellular traps. *J Thromb Haemost* 2014;12(12):2074–2088
- Rossaint J, Herter JM, Van Aken H, et al. Synchronized integrin engagement and chemokine activation is crucial in neutrophil extracellular trap-mediated sterile inflammation. *Blood* 2014;123(16):2573–2584
- Chrysanthopoulou A, Kambas K, Stakos D, et al. Interferon lambda1/IL-29 and inorganic polyphosphate are novel regulators of neutrophil-driven thromboinflammation. *J Pathol* 2017;243(01):111–122
- Martinod K, Wagner DD. Thrombosis: tangled up in NETs. *Blood* 2014;123(18):2768–2776
- Xu J, Zhang X, Pelayo R, et al. Extracellular histones are major mediators of death in sepsis. *Nat Med* 2009;15(11):1318–1321
- Fuchs TA, Brill A, Duerschmied D, et al. Extracellular DNA traps promote thrombosis. *Proc Natl Acad Sci U S A* 2010;107(36):15880–15885
- Iwanaga S, Lee BL. Recent advances in the innate immunity of invertebrate animals. *J Biochem Mol Biol* 2005;38(02):128–150
- Engelmann B, Massberg S. Thrombosis as an intravascular effector of innate immunity. *Nat Rev Immunol* 2013;13(01):34–45
- Loof TG, Mörgelin M, Johansson L, et al. Coagulation, an ancestral serine protease cascade, exerts a novel function in early immune defense. *Blood* 2011;118(09):2589–2598
- Yousefi S, Gold JA, Andina N, et al. Catapult-like release of mitochondrial DNA by eosinophils contributes to antibacterial defense. *Nat Med* 2008;14(09):949–953
- von Köckritz-Blickwede M, Goldmann O, Thulin P, et al. Phagocytosis-independent antimicrobial activity of mast cells by



- means of extracellular trap formation. *Blood* 2008;111(06):3070–3080
- 33 Pertiwi KR, de Boer OJ, Mackaaij C, et al. Extracellular traps derived from macrophages, mast cells, eosinophils and neutrophils are generated in a time-dependent manner during atherosclerosis. *J Pathol* 2019;247(04):505–512
  - 34 Marx C, Novotny J, Salbeck D, et al. Eosinophil-platelet interactions promote atherosclerosis and stabilize thrombosis with eosinophil extracellular traps. *Blood* 2019;134(21):1859–1872
  - 35 Boeltz S, Amini P, Anders H-J, et al. To NET or not to NET: current opinions and state of the science regarding the formation of neutrophil extracellular traps. *Cell Death Differ* 2019;26(03):395–408
  - 36 Papayannopoulos V. Neutrophil extracellular traps in immunity and disease. *Nat Rev Immunol* 2018;18(02):134–147
  - 37 Mauracher L-M, Posch F, Martinod K, et al. Citrullinated histone H3, a biomarker of neutrophil extracellular trap formation, predicts the risk of venous thromboembolism in cancer patients. *J Thromb Haemost* 2018;16(03):508–518
  - 38 Barnado A, Crofford LJ, Oates JC. At the Bedside: neutrophil extracellular traps (NETs) as targets for biomarkers and therapies in autoimmune diseases. *J Leukoc Biol* 2016;99(02):265–278
  - 39 Vogel S, Bodenstern R, Chen Q, et al. Platelet-derived HMGB1 is a critical mediator of thrombosis. *J Clin Invest* 2015;125(12):4638–4654
  - 40 Sollberger G, Tilley DO, Zychlinsky A. Neutrophil extracellular traps: the biology of chromatin externalization. *Dev Cell* 2018;44(05):542–553
  - 41 Chen KW, Monteleone M, Boucher D, et al. Noncanonical inflammatory signaling elicits gasdermin D-dependent neutrophil extracellular traps. *Sci Immunol* 2018;3(26):3
  - 42 Farrera C, Fadeel B. Macrophage clearance of neutrophil extracellular traps is a silent process. *J Immunol* 2013;191(05):2647–2656
  - 43 Apostolidou E, Skendros P, Kambas K, et al. Neutrophil extracellular traps regulate IL-1 $\beta$ -mediated inflammation in familial Mediterranean fever. *Ann Rheum Dis* 2016;75(01):269–277
  - 44 Schauer C, Janko C, Munoz LE, et al. Aggregated neutrophil extracellular traps limit inflammation by degrading cytokines and chemokines. *Nat Med* 2014;20(05):511–517
  - 45 Kolaczowska E, Jenne CN, Surewaard BGJ, et al. Molecular mechanisms of NET formation and degradation revealed by intravital imaging in the liver vasculature. *Nat Commun* 2015;6:6673
  - 46 Chrysanthopoulou A, Mitroulis I, Apostolidou E, et al. Neutrophil extracellular traps promote differentiation and function of fibroblasts. *J Pathol* 2014;233(03):294–307
  - 47 Kambas K, Mitroulis I, Apostolidou E, et al. Autophagy mediates the delivery of thrombogenic tissue factor to neutrophil extracellular traps in human sepsis. *PLoS One* 2012;7(09):e45427
  - 48 Angelidou I, Chrysanthopoulou A, Mitsios A, et al. REDD1/autophagy pathway is associated with neutrophil-driven IL-1 $\beta$  inflammatory response in active ulcerative colitis. *J Immunol* 2018;200(12):3950–3961
  - 49 Skendros P, Chrysanthopoulou A, Rousset F, et al. Regulated in development and DNA damage responses 1 (REDD1) links stress with IL-1 $\beta$ -mediated familial Mediterranean fever attack through autophagy-driven neutrophil extracellular traps. *J Allergy Clin Immunol* 2017;140(05):1378–1387
  - 50 Lande R, Ganguly D, Facchinetti V, et al. Neutrophils activate plasmacytoid dendritic cells by releasing self-DNA-peptide complexes in systemic lupus erythematosus. *Sci Transl Med* 2011;3(73):73ra19
  - 51 Arampatzioglou A, Papazoglou D, Konstantinidis T, et al. Clarithromycin enhances the antibacterial activity and wound healing capacity in type 2 diabetes mellitus by increasing LL-37 load on neutrophil extracellular traps. *Front Immunol* 2018;9:2064
  - 52 Frangou E, Chrysanthopoulou A, Mitsios A, et al. REDD1/autophagy pathway promotes thromboinflammation and fibrosis in human systemic lupus erythematosus (SLE) through NETs decorated with tissue factor (TF) and interleukin-17A (IL-17A). *Ann Rheum Dis* 2019;78(02):238–248
  - 53 Oklu R, Albadawi H, Watkins MT, Monestier M, Sillesen M, Wicky S. Detection of extracellular genomic DNA scaffold in human thrombus: implications for the use of deoxyribonuclease enzymes in thrombolysis. *J Vasc Interv Radiol* 2012;23(05):712–718
  - 54 De Meyer SF, Suidan GL, Fuchs TA, Monestier M, Wagner DD. Extracellular chromatin is an important mediator of ischemic stroke in mice. *Arterioscler Thromb Vasc Biol* 2012;32(08):1884–1891
  - 55 Ge L, Zhou X, Ji W-J, et al. Neutrophil extracellular traps in ischemia-reperfusion injury-induced myocardial no-reflow: therapeutic potential of DNase-based reperfusion strategy. *Am J Physiol Heart Circ Physiol* 2015;308(05):H500–H509
  - 56 Silvestre-Roig C, Braster Q, Wichapong K, et al. Externalized histone H4 orchestrates chronic inflammation by inducing lytic cell death. *Nature* 2019;569(7755):236–240
  - 57 Folco EJ, Mawson TL, Vromman A, et al. Neutrophil extracellular traps induce endothelial cell activation and tissue factor production through interleukin-1 $\alpha$  and cathepsin G. *Arterioscler Thromb Vasc Biol* 2018;38(08):1901–1912
  - 58 Quillard T, Araújo HA, Franck G, Shvartz E, Sukhova G, Libby P. TLR2 and neutrophils potentiate endothelial stress, apoptosis and detachment: implications for superficial erosion. *Eur Heart J* 2015;36(22):1394–1404
  - 59 Franck G, Mawson T, Sausen G, et al. Flow perturbation mediates neutrophil recruitment and potentiates endothelial injury via TLR2 in mice: implications for superficial erosion. *Circ Res* 2017;121(01):31–42
  - 60 Megens RTA, Vijayan S, Lievens D, et al. Presence of luminal neutrophil extracellular traps in atherosclerosis. *Thromb Haemost* 2012;107(03):597–598
  - 61 Mangold A, Alias S, Scherz T, et al. Coronary neutrophil extracellular trap burden and deoxyribonuclease activity in ST-elevation acute coronary syndrome are predictors of ST-segment resolution and infarct size. *Circ Res* 2015;116(07):1182–1192
  - 62 Riegger J, Byrne RA, Joner M, et al; Prevention of Late Stent Thrombosis by an Interdisciplinary Global European Effort (PRESTIGE) Investigators. Histopathological evaluation of thrombus in patients presenting with stent thrombosis. A multicenter European study: a report of the prevention of late stent thrombosis by an interdisciplinary global European effort consortium. *Eur Heart J* 2016;37(19):1538–1549
  - 63 de Boer OJ, Li X, Teeling P, et al. Neutrophils, neutrophil extracellular traps and interleukin-17 associate with the organisation of thrombi in acute myocardial infarction. *Thromb Haemost* 2013;109(02):290–297
  - 64 Maione F, Cicala C, Liverani E, Mascolo N, Perretti M, D'Acquisto F. IL-17A increases ADP-induced platelet aggregation. *Biochem Biophys Res Commun* 2011;408(04):658–662
  - 65 Borissoff JJ, Joosen IA, Versteylen MO, et al. Elevated levels of circulating DNA and chromatin are independently associated with severe coronary atherosclerosis and a prothrombotic state. *Arterioscler Thromb Vasc Biol* 2013;33(08):2032–2040
  - 66 Geiger S, Holdenrieder S, Stieber P, et al. Nucleosomes in serum of patients with early cerebral stroke. *Cerebrovasc Dis* 2006;21(1–2):32–37
  - 67 Lam NY-L, Rainer TH, Wong LK-S, Lam W, Lo YM. Plasma DNA as a prognostic marker for stroke patients with negative neuroimaging within the first 24 h of symptom onset. *Resuscitation* 2006;68(01):71–78
  - 68 Vallés J, Lago A, Santos MT, et al. Neutrophil extracellular traps are increased in patients with acute ischemic stroke: prognostic significance. *Thromb Haemost* 2017;117(10):1919–1929

- 69 Ducroux C, Di Meglio L, Loyau S, et al. Thrombus neutrophil extracellular traps content impair tPA-induced thrombolysis in acute ischemic stroke. *Stroke* 2018;49(03):754–757
- 70 Rangé H, Labreuche J, Louedec L, et al. Periodontal bacteria in human carotid atherosclerosis as a potential trigger for neutrophil activation. *Atherosclerosis* 2014;236(02):448–455
- 71 Delbosc S, Alsac J-M, Journe C, et al. Porphyromonas gingivalis participates in pathogenesis of human abdominal aortic aneurysm by neutrophil activation. Proof of concept in rats. *PLoS One* 2011;6(04):e18679
- 72 Brill A, Fuchs TA, Savchenko AS, et al. Neutrophil extracellular traps promote deep vein thrombosis in mice. *J Thromb Haemost* 2012;10(01):136–144
- 73 van Montfoort ML, Stephan F, Lauw MN, et al. Circulating nucleosomes and neutrophil activation as risk factors for deep vein thrombosis. *Arterioscler Thromb Vasc Biol* 2013;33(01):147–151
- 74 Diaz JA, Fuchs TA, Jackson TO, et al; for the Michigan Research Venous Group\*. Plasma DNA is elevated in patients with deep vein thrombosis. *J Vasc Surg Venous Lymphat Disord* 2013;1(04):1
- 75 Savchenko AS, Martinod K, Seidman MA, et al. Neutrophil extracellular traps form predominantly during the organizing stage of human venous thromboembolism development. *J Thromb Haemost* 2014;12(06):860–870
- 76 Meier TR, Myers DD Jr, Wroblewski SK, et al. Prophylactic P-selectin inhibition with PSI-421 promotes resolution of venous thrombosis without anticoagulation. *Thromb Haemost* 2008;99(02):343–351
- 77 Das R, Burke T, Plow EF. Histone H2B as a functionally important plasminogen receptor on macrophages. *Blood* 2007;110(10):3763–3772
- 78 Varjú I, Longstaff C, Szabó L, et al. DNA, histones and neutrophil extracellular traps exert anti-fibrinolytic effects in a plasma environment. *Thromb Haemost* 2015;113(06):1289–1298
- 79 Longstaff C, Varjú I, Sótönyi P, et al. Mechanical stability and fibrinolytic resistance of clots containing fibrin, DNA, and histones. *J Biol Chem* 2013;288(10):6946–6956
- 80 Lang IM, Madani M. Update on chronic thromboembolic pulmonary hypertension. *Circulation* 2014;130(06):508–518
- 81 Aldabbous L, Abdul-Salam V, McKinnon T, et al. Neutrophil extracellular traps promote angiogenesis: evidence from vascular pathology in pulmonary hypertension. *Arterioscler Thromb Vasc Biol* 2016;36(10):2078–2087
- 82 Knight JS, Carmona-Rivera C, Kaplan MJ. Proteins derived from neutrophil extracellular traps may serve as self-antigens and mediate organ damage in autoimmune diseases. *Front Immunol* 2012;3:380
- 83 Nakazawa D, Tomaru U, Yamamoto C, Jodo S, Ishizu A. Abundant neutrophil extracellular traps in thrombus of patient with microscopic polyangiitis. *Front Immunol* 2012;3:333
- 84 Imamoto T, Nakazawa D, Shida H, et al. Possible linkage between microscopic polyangiitis and thrombosis via neutrophil extracellular traps. *Clin Exp Rheumatol* 2014;32(01):149–150
- 85 Kambas K, Chrysanthopoulou A, Vassilopoulos D, et al. Tissue factor expression in neutrophil extracellular traps and neutrophil derived microparticles in antineutrophil cytoplasmic antibody associated vasculitis may promote thromboinflammation and the thrombophilic state associated with the disease. *Ann Rheum Dis* 2014;73(10):1854–1863
- 86 Huang Y-M, Wang H, Wang C, Chen M, Zhao MH. Promotion of hypercoagulability in antineutrophil cytoplasmic antibody-associated vasculitis by C5a-induced tissue factor-expressing microparticles and neutrophil extracellular traps. *Arthritis Rheumatol* 2015;67(10):2780–2790
- 87 Mitroulis I, Kambas K, Anyfanti P, Doumas M, Ritis K. The multivalent activity of the tissue factor-thrombin pathway in thrombotic and non-thrombotic disorders as a target for therapeutic intervention. *Expert Opin Ther Targets* 2011;15(01):75–89
- 88 Posma JJ, Grover SP, Hisada Y, et al. Roles of coagulation proteases and PARs (protease-activated receptors) in mouse models of inflammatory diseases. *Arterioscler Thromb Vasc Biol* 2019;39(01):13–24
- 89 Leffler J, Stojanovich L, Shoenfeld Y, Bogdanovic G, Hesselstrand R, Blom AM. Degradation of neutrophil extracellular traps is decreased in patients with antiphospholipid syndrome. *Clin Exp Rheumatol* 2014;32(01):66–70
- 90 Yalavarthi S, Gould TJ, Rao AN, et al. Release of neutrophil extracellular traps by neutrophils stimulated with antiphospholipid antibodies: a newly identified mechanism of thrombosis in the antiphospholipid syndrome. *Arthritis Rheumatol* 2015;67(11):2990–3003
- 91 Meng H, Yalavarthi S, Kanthi Y, et al. In vivo role of neutrophil extracellular traps in antiphospholipid antibody-mediated venous thrombosis. *Arthritis Rheumatol* 2017;69(03):655–667
- 92 Gollomp K, Kim M, Johnston I, et al. Neutrophil accumulation and NET release contribute to thrombosis in HIT. *JCI Insight* 2018;3(18):3
- 93 Perdomo J, Leung HHL, Ahmadi Z, et al. Neutrophil activation and NETosis are the major drivers of thrombosis in heparin-induced thrombocytopenia. *Nat Commun* 2019;10(01):1322
- 94 Jiménez-Alcázar M, Napirei M, Panda R, et al. Impaired DNase1-mediated degradation of neutrophil extracellular traps is associated with acute thrombotic microangiopathies. *J Thromb Haemost* 2015;13(05):732–742
- 95 Wang Y, Luo L, Braun OÖ, et al. Neutrophil extracellular trap-microparticle complexes enhance thrombin generation via the intrinsic pathway of coagulation in mice. *Sci Rep* 2018;8(01):4020
- 96 He Z, Si Y, Jiang T, et al. Phosphatidylserine exposure and neutrophil extracellular traps enhance procoagulant activity in patients with inflammatory bowel disease. *Thromb Haemost* 2016;115(04):738–751
- 97 Maugeri N, Capobianco A, Rovere-Querini P, et al. Platelet microparticles sustain autophagy-associated activation of neutrophils in systemic sclerosis. *Sci Transl Med* 2018;10(451):10
- 98 Qi H, Yang S, Zhang L. Neutrophil extracellular traps and endothelial dysfunction in atherosclerosis and thrombosis. *Front Immunol* 2017;8:928
- 99 Wiseman SJ, Ralston SH, Wardlaw JM. Cerebrovascular disease in rheumatic diseases: a systematic review and meta-analysis. *Stroke* 2016;47(04):943–950
- 100 Agca R, Heslinga SC, Rollefstad S, et al. EULAR recommendations for cardiovascular disease risk management in patients with rheumatoid arthritis and other forms of inflammatory joint disorders: 2015/2016 update. *Ann Rheum Dis* 2017;76(01):17–28
- 101 Gu M-M, Wang X-P, Cheng Q-Y, et al. A meta-analysis of cardiovascular events in systemic lupus erythematosus. *Immunol Invest* 2019;48(05):505–520
- 102 Knight JS, Zhao W, Luo W, et al. Peptidylarginine deiminase inhibition is immunomodulatory and vasculoprotective in murine lupus. *J Clin Invest* 2013;123(07):2981–2993
- 103 Grayson PC, Carmona-Rivera C, Xu L, et al; Rituximab in ANCA-Associated Vasculitis-Immune Tolerance Network Research Group. Neutrophil-related gene expression and low-density granulocytes associated with disease activity and response to treatment in antineutrophil cytoplasmic antibody-associated vasculitis. *Arthritis Rheumatol* 2015;67(07):1922–1932
- 104 Knight JS, Meng H, Coit P, et al. Activated signature of antiphospholipid syndrome neutrophils reveals potential therapeutic target. *JCI Insight* 2017;2(18):2
- 105 Weeding E, Coit P, Yalavarthi S, Kaplan MJ, Knight JS, Sawalha AH. Genome-wide DNA methylation analysis in primary antiphospholipid syndrome neutrophils. *Clin Immunol* 2018;196:110–116
- 106 Ali RA, Gandhi AA, Meng H, et al. Adenosine receptor agonism protects against NETosis and thrombosis in antiphospholipid syndrome. *Nat Commun* 2019;10(01):1916

- 107 White C, Noble SIR, Watson M, et al. Prevalence, symptom burden, and natural history of deep vein thrombosis in people with advanced cancer in specialist palliative care units (HIDDEN): a prospective longitudinal observational study. *Lancet Haematol* 2019;6(02):e79–e88
- 108 Spek CA, Versteeg HH, Borensztajn KS. Anticoagulant therapy of cancer patients: Will patient selection increase overall survival? *Thromb Haemost* 2015;114(03):530–536
- 109 Demers M, Krause DS, Schatzberg D, et al. Cancers predispose neutrophils to release extracellular DNA traps that contribute to cancer-associated thrombosis. *Proc Natl Acad Sci U S A* 2012;109(32):13076–13081
- 110 Alfaro C, Teijeira A, Oñate C, et al. Tumor-produced interleukin-8 attracts human myeloid-derived suppressor cells and elicits extrusion of neutrophil extracellular traps (NETs). *Clin Cancer Res* 2016;22(15):3924–3936
- 111 Leal AC, Mizurini DM, Gomes T, et al. Tumor-derived exosomes induce the formation of neutrophil extracellular traps: implications for the establishment of cancer-associated thrombosis. *Sci Rep* 2017;7(01):6438
- 112 Cools-Lartigue J, Spicer J, McDonald B, et al. Neutrophil extracellular traps sequester circulating tumor cells and promote metastasis. *J Clin Invest* 2013;67484
- 113 Cedervall J, Zhang Y, Huang H, et al. Neutrophil extracellular traps accumulate in peripheral blood vessels and compromise organ function in tumor-bearing animals. *Cancer Res* 2015;75(13):2653–2662
- 114 Park J, Wysocki RW, Amoozgar Z, et al. Cancer cells induce metastasis-supporting neutrophil extracellular DNA traps. *Sci Transl Med* 2016;8(361):361ra138
- 115 Wolach O, Sellar RS, Martinod K, et al. Increased neutrophil extracellular trap formation promotes thrombosis in myeloproliferative neoplasms. *Sci Transl Med* 2018;10(436):10
- 116 Thälén C, Demers M, Blomgren B, et al. NETosis promotes cancer-associated arterial microthrombosis presenting as ischemic stroke with troponin elevation. *Thromb Res* 2016;139:56–64
- 117 Hisada Y, Grover SP, Maqsood A, et al. Neutrophils and neutrophil extracellular traps enhance venous thrombosis in mice bearing human pancreatic tumors. *Haematologica* 2019;haematol.2019.217083
- 118 Arelaki S, Arampatzioglou A, Kambas K, et al. Gradient infiltration of neutrophil extracellular traps in colon cancer and evidence for their involvement in tumour growth. *PLoS One* 2016;11(05):e0154484
- 119 Bauer AT, Suckau J, Frank K, et al. von Willebrand factor fibers promote cancer-associated platelet aggregation in malignant melanoma of mice and humans. *Blood* 2015;125(20):3153–3163
- 120 Guglietta S, Chiavelli A, Zagato E, et al. Coagulation induced by C3aR-dependent NETosis drives protumorigenic neutrophils during small intestinal tumorigenesis. *Nat Commun* 2016;7:11037
- 121 Grilz E, Mauracher L-M, Posch F, et al. Citrullinated histone H3, a biomarker for neutrophil extracellular trap formation, predicts the risk of mortality in patients with cancer. *Br J Haematol* 2019;186(02):311–320
- 122 McDonald B, Davis RP, Kim S-J, et al. Platelets and neutrophil extracellular traps collaborate to promote intravascular coagulation during sepsis in mice. *Blood* 2017;129(10):1357–1367
- 123 Iba T, Miki T, Hashiguchi N, Tabe Y, Nagaoka I. Is the neutrophil a 'prima donna' in the procoagulant process during sepsis? *Crit Care* 2014;18(04):230
- 124 Liaw PC, Ito T, Iba T, Thachil J, Zeerleder S. DAMP and DIC: the role of extracellular DNA and DNA-binding proteins in the pathogenesis of DIC. *Blood Rev* 2016;30(04):257–261
- 125 Gould TJ, Vu TT, Stafford AR, et al. Cell-free DNA modulates clot structure and impairs fibrinolysis in sepsis. *Arterioscler Thromb Vasc Biol* 2015;35(12):2544–2553
- 126 Yang S, Qi H, Kan K, et al. Neutrophil extracellular traps promote hypercoagulability in patients with sepsis. *Shock* 2017;47(02):132–139
- 127 Delabranche X, Stiel L, Severac F, et al. Evidence of netosis in septic shock-induced disseminated intravascular coagulation. *Shock* 2017;47(03):313–317
- 128 Sharma A, McCann K, Tripathi JK, et al. Tamoxifen restores extracellular trap formation in neutrophils from patients with chronic granulomatous disease in a reactive oxygen species-independent manner. *J Allergy Clin Immunol* 2019;144(02):597–600.e3
- 129 Rockel JS, Kapoor M. Autophagy: controlling cell fate in rheumatic diseases. *Nat Rev Rheumatol* 2016;12(09):517–531
- 130 Papagoras C, Chrysanthopoulou A, Mitsios A, Arampatzioglou A, Ritis K, Skendros P. Autophagy inhibition in adult-onset Still's disease: still more space for hydroxychloroquine? *Clin Exp Rheumatol* 2017;35(06, Suppl 108):133–134
- 131 Belizna C, Pregnolato F, Abad S, et al. HIBISCUS: hydroxychloroquine for the secondary prevention of thrombotic and obstetrical events in primary antiphospholipid syndrome. *Autoimmun Rev* 2018;17(12):1153–1168
- 132 Manfredi AA, Rovere-Querini P, D'Angelo A, Maugeri N. Low molecular weight heparins prevent the induction of autophagy of activated neutrophils and the formation of neutrophil extracellular traps. *Pharmacol Res* 2017;123:146–156
- 133 Van Avondt K, Maegdefessel L, Soehnlein O. Therapeutic targeting of neutrophil extracellular traps in atherogenic inflammation. *Thromb Haemost* 2019;119(04):542–552
- 134 Dubois AV, Gauthier A, Bréa D, et al. Influence of DNA on the activities and inhibition of neutrophil serine proteases in cystic fibrosis sputum. *Am J Respir Cell Mol Biol* 2012;47(01):80–86
- 135 Mastellos DC, Reis ES, Ricklin D, Smith RJ, Lambris JD. Complement C3-targeted therapy: replacing long-held assertions with evidence-based discovery. *Trends Immunol* 2017;38(06):383–394
- 136 Boone BA, Murthy P, Miller-Ocuin J, et al. Chloroquine reduces hypercoagulability in pancreatic cancer through inhibition of neutrophil extracellular traps. *BMC Cancer* 2018;18(01):678
- 137 Mitroulis I, Kambas K, Chrysanthopoulou A, et al. Neutrophil extracellular trap formation is associated with IL-1 $\beta$  and autophagy-related signaling in gout. *PLoS One* 2011;6(12):e29318
- 138 Healy LD, Puy C, Itakura A, et al. Colocalization of neutrophils, extracellular DNA and coagulation factors during NETosis: development and utility of an immunofluorescence-based microscopy platform. *J Immunol Methods* 2016;435:77–84
- 139 Abrams ST, Morton B, Alhamdi Y, et al. A novel assay for neutrophil extracellular traps (NETs) formation independently predicts disseminated intravascular coagulation and mortality in critically ill patients. *Am J Respir Crit Care Med* 2019;200(07):869–880