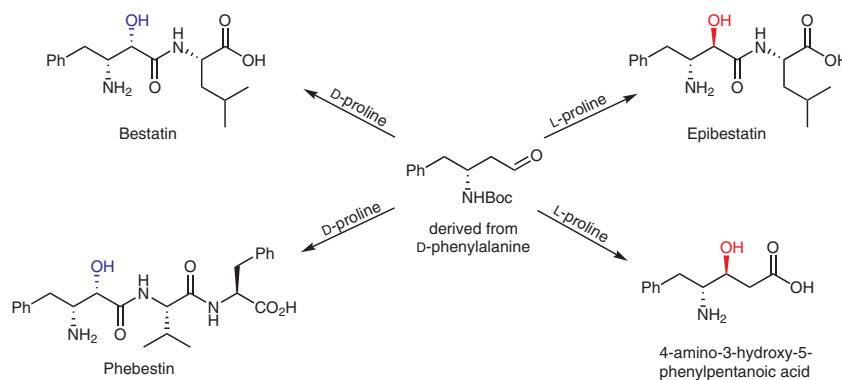


Diastereoselective Synthesis of (–)-Bestatin, Epibestatin, Phebestin and (3*S*,4*R*)-4-Amino-3-hydroxy-5-phenylpentanoic Acid from an Aldehyde Derived from D-Phenylalanine

Vipin Kumar Jain*

Department of Chemistry, Indian Institute of Technology Kanpur, Kanpur-208016, India
jain91vipin@gmail.com



Received: 27.09.2019

Accepted after revision: 09.10.2019

Published online: 25.10.2019

DOI: 10.1055/s-0039-1690223; Art ID: so-2019-h0027-op

License terms:

© 2019. The Author(s). This is an open access article published by Thieme under the terms of the Creative Commons Attribution-NonDerivative-NonCommercial License, permitting copying and reproduction so long as the original work is given appropriate credit. Contents may not be used for commercial purposes or adapted, remixed, transformed or built upon. (<https://creativecommons.org/licenses/by-nc-nd/4.0/>)

Abstract A convenient and efficient method for the synthesis of (–)-bestatin, epibestatin, phebestin, and (3*S*,4*R*)-4-amino-3-hydroxy-5-phenylpentanoic acid is reported. The key step is a proline-catalyzed α -hydroxylation of an aldehyde derived from D-phenylalanine, which leads to incorporation of a hydroxyl group at the α -position of that aldehyde with good yield and very high diastereoselectivity. Bestatin and its diastereomer epibestatin are synthesized from the same starting material using the same sequence of reactions, except for proline as the catalyst. An *O*-MOM and Boc-protected amino acid, a common intermediate for bestatin, was coupled with a dipeptide, H-Val-Phe-OMe followed by global deprotection to yield phebestin. (3*S*,4*R*)-4-Amino-3-hydroxy-5-phenylpentanoic acid was also synthesized in eight steps from the same starting material. The reported synthetic route offers a general method for the synthesis of such types of compounds and their analogues by changing the proline catalyst and/or the starting material from D- to L-phenylalanine.

Key words asymmetric hydroxylation, organocatalysis, reductive cleavage

Structure modification studies of bestatin and similar molecules such as phebestin, a tripeptide, indicate that biological activities of these molecules are significantly influenced by the (2*S*)-*syn*-stereochemistry of the hydroxyl group.^{9,10}

Various stereoselective methods for the synthesis of bestatin, phebestin^{11–27} and epibestatin^{28,29} are available and most of them utilized D-phenylalanine as a chiral starting material. Reported herein is an alternative and short method for the synthesis of bestatin, epibestatin, phebestin and (3*S*,4*R*)-4-amino-3-hydroxy-5-phenylpentanoic acid using proline-catalyzed asymmetric α -hydroxylation of an aldehyde derived from D-phenylalanine. The structures of these compounds are shown in Figure 1.

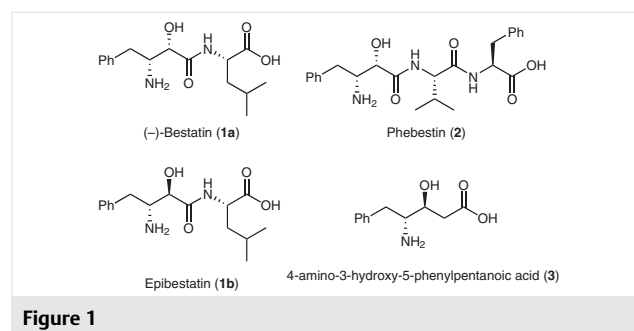


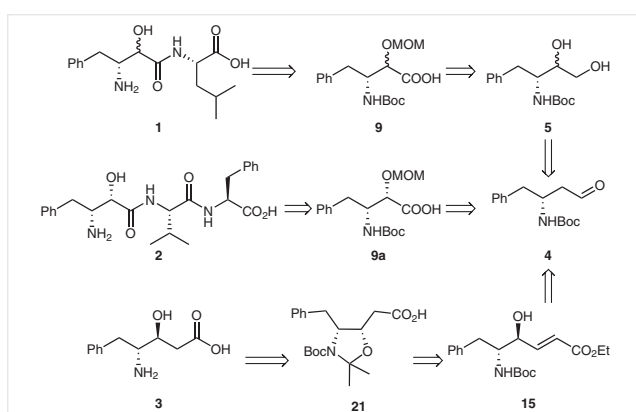
Figure 1

(–)-Bestatin (Ubenimex) is a dipeptide containing an α -hydroxy- β -amino amide subunit that was first isolated from *Streptomyces olivoreticulithe* by Umezawa et al. in 1976.^{1,2} It is an aminopeptidase inhibitor that exhibits immunostimulatory activity as well as cytotoxic activity.^{3,4} It is used clinically for the treatment of cancer, HIV, hypertension, and shows potential as an anti-inflammatory agent.^{5–8}

Proline-catalyzed α -hydroxylation of an aldehyde using nitrosobenzene followed by reduction of the N–O bond is an attractive method to introduce a hydroxyl group stereoselectively.^{30–32} The aldehyde functional group can be further reduced to an alcohol or converted into an alkene through Wittig reaction in order to avoid racemization at the α -position. As the part of our studies towards the synthesis of various bioactive and naturally occurring mole-

cules,^{32–41} we recently reported the synthesis of *D*-threo-sphinganine, *L*-erythro-sphinganine and (–)-spisulosine from an aldehyde derived from aspartic acid.⁴²

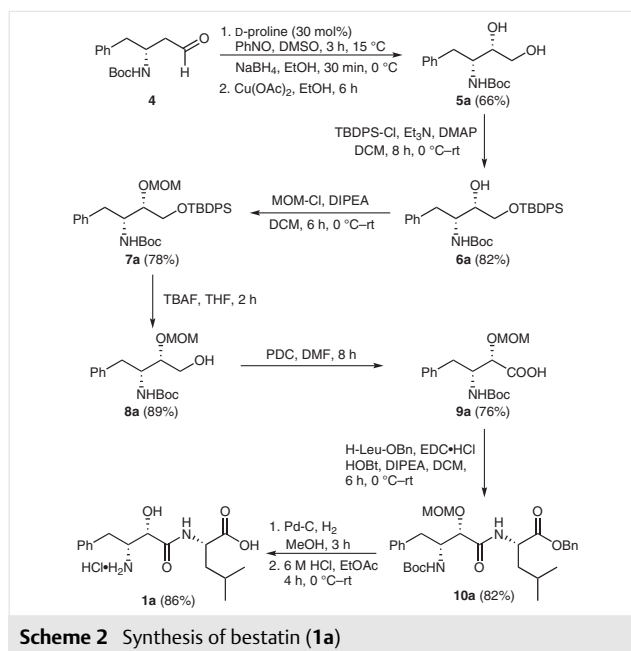
In the retrosynthetic analysis, it was anticipated that both bestatin and epibestatin could be synthesized from acid **9** using peptide coupling followed by deprotection of the Boc and MOM groups. Diol **5** could be obtained from aldehyde **4** using an α -hydroxylation reaction. Compound **9a** could be converted into phebestin. Olefin **15** could be obtained from aldehyde **4** using an α -hydroxylation reaction followed by Wittig reaction and would yield compound **3** as shown in Scheme 1.



Scheme 1 Retrosynthetic analysis of compounds **1**, **2** and **3** from **4**

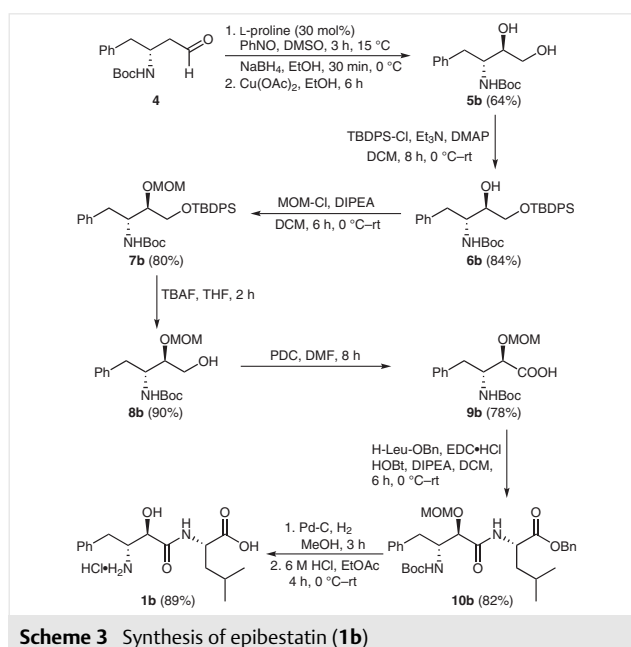
Aldehyde **4** (for preparation see the literature⁴³) was subjected to diastereoselective hydroxylation using nitrosobenzene, and *D*-proline as catalyst and subsequently reduced to the corresponding primary alcohol by NaBH_4 in one pot. The crude product was further subjected to N–O bond cleavage using $\text{Cu}(\text{OAc})_2$ to give diol **5a** in 66% yield overall. It was observed by ^1H NMR spectroscopy that the hydroxylation reaction proceeded with 90:10 diastereoselectivity. The primary and secondary hydroxyl groups of compound **5a** were protected as their TBDPS and MOM derivatives, respectively, to obtain the fully protected compound **7a** in 64% overall yield. TBAF was then used to remove the silyl protecting group in compound **7a** to furnish the primary alcohol **8a** in 89% yield, which was then treated with PDC in DMF to produce the corresponding carboxylic acid **9a** in 76% yield (Scheme 2).

The fully protected α -hydroxy- β -amino acid **9a** is the precursor for the synthesis of both bestatin and phebestin. To obtain bestatin, compound **9a** was coupled with the benzyl ester of *L*-leucine in the presence of EDC·HCl, HOBT and DIPEA to give the corresponding fully protected dipeptide **10a** in 82% yield. Compound **10a** was further subjected to Pd-catalysed hydrogenolysis followed by acidolysis of the Boc and MOM groups to furnish target molecule **1a** from **10a** in 86% yield (Scheme 2).



Scheme 2 Synthesis of bestatin (**1a**)

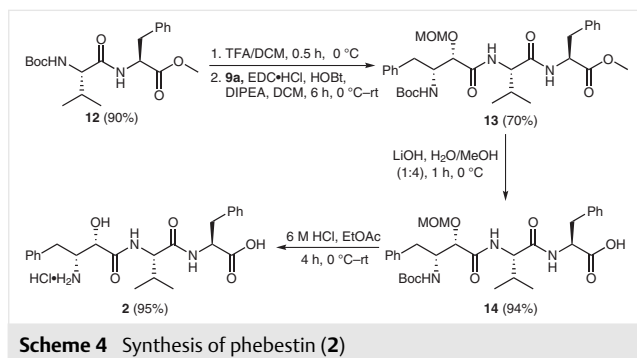
Epibestatin **1b** was obtained in an overall yield of 22% from aldehyde **4** using exactly the same sequence of reactions but using *L*-proline in the asymmetric α -hydroxylation reaction (Scheme 3) leading to a diastereomer ratio of 87:13 as judged by ^1H NMR spectroscopy. Epibestatin is available in very limited quantities commercially and to date only a few synthetic strategies have been reported.^{28,29}



Scheme 3 Synthesis of epibestatin (**1b**)

To synthesize phebestin, compound **9a** was coupled with dipeptide **12**, which was obtained from coupling the methyl ester of *L*-phenylalanine with NH-Boc protected

L-valine, to give the fully protected tripeptide **13** in 70% yield. Hydrolysis of the methyl ester using LiOH followed by acidolysis of the Boc and MOM groups furnished the target molecule **2** in 89% yield over two steps (Scheme 4).

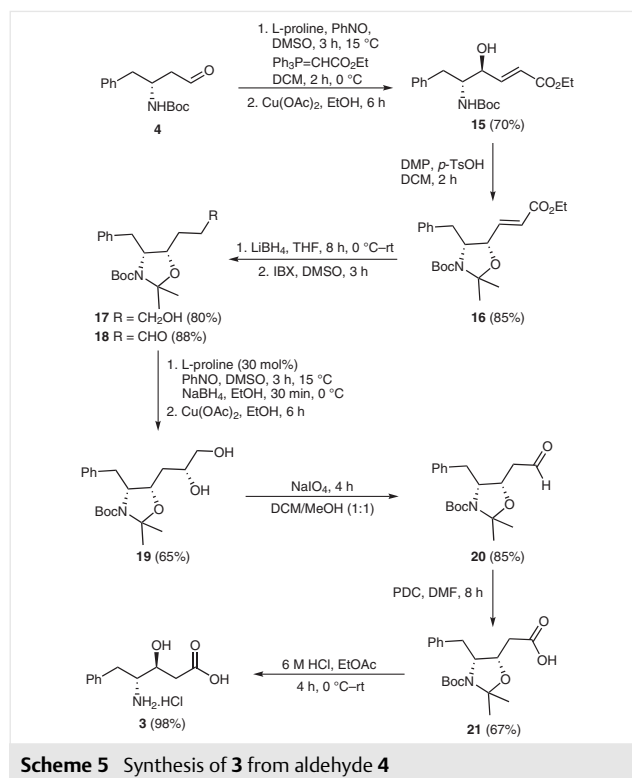


β -Hydroxy- γ -amino acids have been designed for biologically active peptide mimics and for HIV protease inhibitors. Stictamide A, tasiamide B and hapolosin are biologically important compounds that contain 4-amino-3-hydroxy-5-phenylpentanoic acid as a structural fragment. The activities of such compounds depend on the stereochemistries of both the amino- and hydroxyl groups.^{44,45} A variety of stereoselective methods for the synthesis of these acids and their analogues is available.^{46–50} (3*S*,4*R*)-4-Amino-3-hydroxy-5-phenylpentanoic acid (**3**) was also synthesized from the same starting material **4** in eight steps and in an overall yield of 15% (Scheme 5).

Thus, aldehyde **4** was subjected to L-proline-catalysed asymmetric α -hydroxylation and subsequent Wittig reaction in one pot. The crude product was further treated with Cu(OAc)₂ leading to cleavage of the N–O bond to form olefin **15** in 70% overall yield (Scheme 5).

Both the hydroxyl and amino groups in compound **15** were protected as an oxazolidine using 2,2-dimethoxypropane (DMP) and a catalytic amount of *p*-TsOH to **16** in 85% yield. LiBH₄ was used to reduce compound **16** to primary alcohol **17** in 80% yield, and this was then oxidized to aldehyde **18** using 2-iodoxybenzoic acid (IBX) in 88% yield. The aldehyde **18** was subjected to L-proline-catalysed asymmetric α -hydroxylation reaction followed by reduction and N–O bond cleavage using NaBH₄ and Cu(OAc)₂, respectively, to furnish diol **19** in 65% overall yield. NaIO₄ was used to cleave the diol to produce aldehyde **20**, which was further oxidised to an acid **21** using PDC in 57% yield after two steps. Acidolysis of the Boc group and oxazolidine ring in compound **21** furnished **3** in 98% yield (Scheme 5).

In conclusion, we have demonstrated a convenient and efficient route for the synthesis of bestatin, epibestatin, phebestin and (3*S*,4*R*)-4-amino-3-hydroxy-5-phenylpentanoic acid using proline-catalysed α -hydroxylation of an aldehyde derived from D-phenylalanine with high diastereo-



selectivities and in good overall yields. The method described here offers a general method to synthesize several similar molecules using an organocatalytic route.

See the Supporting Information for general information.

Asymmetric α -Hydroxylation of Aldehydes; General Procedure

To a stirred solution of aldehyde **4** (1.00 g, 3.80 mmol) and nitrosobenzene (0.44 g, 4.18 mmol) in anhydrous DMSO (10 mL), D- or L-proline (0.13 g, 1.14 mmol, 30 mol%) was added at 15 °C. The mixture was stirred for 3 h at the same temperature, then cooled to 0 °C and NaBH₄ (0.28 g, 7.60 mmol) in EtOH (15 mL) was added and the mixture was stirred vigorously for 30 min at 0 °C. On complete disappearance of starting material, the reaction was quenched with saturated aqueous NH₄Cl (30 mL) and the mixture was extracted with EtOAc (2 × 30 mL). The combined organic phases were washed with brine (30 mL), dried over Na₂SO₄, filtered, and concentrated. The crude amino-hydroxylated product was taken as such to the next step leading to the cleavage of O–N bond.

Cu(OAc)₂ (0.17 g, 0.96 mmol) was added to a stirred solution of the above product in EtOH (15 mL) and the mixture was stirred vigorously for 6 h at room temperature. On complete disappearance of starting material, the reaction was quenched with saturated aqueous NH₄Cl (20 mL) and extracted with EtOAc (2 × 20 mL). The combined organic phases were washed with brine (30 mL), dried over Na₂SO₄, filtered, concentrated, and purified by column chromatography.

The same procedure was used for the preparation of compound **19**.

tert-Butyl ((2R,3S)-3,4-Dihydroxy-1-phenylbutan-2-yl)carbamate (5a)

Column chromatography (petroleum ether/EtOAc, 60:40).

Yield: 0.70 g (66%); clear oil; $[\alpha]_D^{27} +18.97$ (c 1.22, CHCl₃).

IR (thin film): 3382, 3063, 3028, 2924, 2854, 1682, 1604 cm⁻¹.

¹H NMR (CDCl₃, 400 MHz): δ = 7.28–7.17 (m, 5 H), 4.98 (d, *J* = 8.0 Hz, 1 H), 4.65 (d, *J* = 8.0 Hz, 1 H), 3.91–3.89 (m, 1 H), 3.63–3.38 (m, 4 H), 3.18 (br s, 1 H), 2.88 (d, *J* = 8.0 Hz, 2 H), 1.37 (s, 9 H).

¹³C NMR (CDCl₃, 125 MHz): δ = 156.9, 138.0, 129.5, 129.3, 128.6, 126.5, 80.5, 80.1, 73.2, 71.6, 64.0, 59.6, 52.6, 38.2, 31.3, 29.8, 28.4.

HRMS (ESI–TOF): *m/z* [M + Na]⁺ calcd for C₁₅H₂₃NNaO₄: 304.1525; found: 304.1523.

tert-Butyl ((2R,3R)-3,4-Dihydroxy-1-phenylbutan-2-yl)carbamate (5b)

Column chromatography (petroleum ether/EtOAc, 60:40).

Yield: 0.69 g (64%); clear oil; $[\alpha]_D^{27} -8.59$ (c 0.74, CHCl₃).

IR (thin film): 3360, 2978, 2928, 1686, 1524 cm⁻¹.

¹H NMR (CDCl₃, 500 MHz): δ = 7.31–7.22 (m, 5 H), 4.82 (d, *J* = 5.0 Hz, 1 H), 4.56 (d, *J* = 5.0 Hz, 1 H), 3.86–3.81 (m, 1 H), 3.68–3.36 (m, 4 H), 3.11–3.08 (m, 1 H), 2.92–2.88 (m, 2 H), 1.38 (s, 9 H).

¹³C NMR (CDCl₃, 125 MHz): δ = 157.2, 137.4, 129.5, 128.8, 126.8, 80.6, 73.2, 63.0, 52.4, 36.6, 31.7, 29.8, 28.3.

HRMS (ESI–TOF): *m/z* [M + Na]⁺ calcd for C₁₅H₂₃NNaO₄: 304.1525; found: 304.1528.

tert-Butyl (4R,5S)-4-Benzyl-5-((R)-2,3-dihydroxypropyl)-2,2-dimethyloxazolidine-3-carboxylate (19)

Column chromatography (petroleum ether/EtOAc, 50:50).

Yield: 0.68 g (65%); clear oil; $[\alpha]_D^{27} +11.94$ (c 0.92, CHCl₃).

IR (thin film): 3418, 3063, 3029, 2924, 2855, 1694, 1682, 1604 cm⁻¹.

¹H NMR (CDCl₃, 500 MHz): δ = 7.27–7.18 (m, 5 H), 4.29–4.10 (m, 2 H), 3.66 (br s, 1 H), 3.46–3.23 (m, 2 H), 2.97–2.82 (m, 2 H), 1.82–1.67 (m, 3 H), 1.57–1.53 (m, 6 H), 1.44, 1.34 (s, 9 H).

¹³C NMR (CDCl₃, 125 MHz): δ = 151.9, 151.6, 138.7, 129.4, 129.3, 128.6, 128.4, 126.4, 126.2, 93.6, 92.9, 80.4, 80.0, 76.1, 71.2, 71.1, 66.3, 61.1, 60.9, 36.7, 36.0, 32.9, 29.8, 28.4, 28.1, 27.5, 26.8, 25.2, 24.0.

HRMS (ESI–TOF): *m/z* [M + Na]⁺ calcd for C₂₀H₃₁NNaO₅: 388.2100; found: 388.2100.

Silyl Protection; General Procedure

Compound **5** (1.00 g, 3.55 mmol) was dissolved in anhydrous DCM (20 mL) and the solution cooled to 0 °C. TBDPSCI (1.07 mL, 3.91 mmol), DMAP (0.08 g, 0.71 mmol) and triethylamine (0.74 mL, 5.32 mmol) were added and the reaction mixture was stirred at r.t. for 8 h. On complete disappearance of starting material, the reaction was quenched with saturated aqueous citric acid (20 mL), the crude product was extracted with DCM (2 × 30 mL) and the combined organic phases containing crude product were dried over Na₂SO₄, filtered, concentrated under vacuum, and purified by column chromatography.

tert-butyl ((2R,3S)-4-((tert-Butyldiphenylsilyloxy)-3-hydroxy-1-phenylbutan-2-yl)carbamate (6a)

Column chromatography (petroleum ether/EtOAc, 80:20).

Yield: 1.51 g (82%); clear oil; $[\alpha]_D^{27} +17.28$ (c 0.96, CHCl₃).

IR (thin film): 3434, 3070, 3027, 2927, 2856, 1689 cm⁻¹.

¹H NMR (CDCl₃, 400 MHz): δ = 7.62–7.57 (m, 4 H), 7.42–7.21 (m, 11 H), 4.93 (br s, 1 H), 3.76–3.69 (m, 2 H), 3.61–3.60 (m, 2 H), 2.96–2.85 (m, 2 H), 2.67 (br s, 1 H), 1.35 (s, 9 H), 1.04 (s, 9 H).

¹³C NMR (CDCl₃, 100 MHz): δ = 155.9, 138.4, 135.6, 133.1, 130.0, 129.5, 128.5, 127.9, 126.4, 79.4, 71.1, 65.7, 52.7, 38.6, 29.8, 28.4, 27.0, 19.3.

HRMS (ESI–TOF): *m/z* [M + Na]⁺ calcd for C₃₁H₄₁NNaO₄Si: 542.2703; found: 542.2700.

tert-Butyl ((2R,3R)-4-((tert-Butyldiphenylsilyloxy)-3-hydroxy-1-phenylbutan-2-yl)carbamate (6b)

Column chromatography (petroleum ether/EtOAc, 80:20).

Yield: 1.55 g (84%); clear oil; $[\alpha]_D^{27} +2.26$ (c 1.45, CHCl₃).

IR (thin film): 3417, 2930, 2857, 1692, 1497 cm⁻¹.

¹H NMR (CDCl₃, 500 MHz): δ = 7.70–7.68 (m, 3 H), 7.46–7.39 (m, 6 H), 7.29–7.17 (m, 6 H), 4.98 (br s, 1 H), 3.99 (br s, 1 H), 3.76–3.62 (m, 3 H), 3.08 (br s, 1 H), 2.96–2.85 (m, 2 H), 1.36 (s, 9 H), 1.11 (s, 9 H).

¹³C NMR (CDCl₃, 125 MHz): δ = 156.0, 138.0, 135.7, 132.9, 132.8, 130.0, 129.5, 128.5, 128.0, 127.9, 126.4, 79.4, 72.6, 65.4, 54.3, 36.6, 29.8, 28.4, 27.0, 19.3.

HRMS (ESI–TOF): *m/z* [M + Na]⁺ calcd for C₃₁H₄₁NNaO₄Si: 542.2703; found: 542.2705.

MOM Protection; General Procedure

MOM chloride (0.58 mL, 7.68 mmol) followed by Hunig's base, DIPEA (1.68 mL, 9.62 mmol) were added to a stirred solution of compound **6** (1.00 g, 1.92 mmol) in DCM (25 mL) at 0 °C, and the mixture was stirred vigorously at r.t. for 6 h. On complete disappearance of starting material, the reaction was quenched with water (20 mL), and the mixture was extracted with DCM (2 × 30 mL) and the combined organic phases were washed with 2% HCl (2 × 20 mL), dried over Na₂SO₄, filtered, concentrated and purified through column chromatography.

tert-Butyl ((2R,3S)-4-((tert-Butyldiphenylsilyloxy)-3-(methoxy-methoxy)-1-phenylbutan-2-yl)carbamate (7a)

Column chromatography (petroleum ether/EtOAc, 85:15).

Yield: 0.84 g (78%); clear oil; $[\alpha]_D^{27} +1.65$ (c 0.48, CHCl₃).

IR (thin film): 2928, 2856, 1715, 1494 cm⁻¹.

¹H NMR (CDCl₃, 400 MHz): δ = 7.53–7.47 (m, 4 H), 7.35–7.15 (m, 11 H), 4.93 (d, *J* = 8.0 Hz, 1 H), 4.58–4.43 (m, 2 H), 4.09–4.04 (m, 1 H), 3.56–3.51 (m, 3 H), 3.28 (s, 3 H), 2.88–2.71 (m, 2 H), 1.33 (s, 9 H), 0.91 (s, 9 H).

¹³C NMR (CDCl₃, 100 MHz): δ = 155.5, 135.6, 133.2, 129.7, 129.6, 128.5, 127.8, 126.3, 97.1, 79.1, 63.6, 55.9, 52.4, 38.7, 28.5, 26.8, 19.2.

HRMS (ESI–TOF): *m/z* [M + H]⁺ calcd for C₃₃H₄₆NO₅Si: 564.3145; found: 564.3141.

tert-Butyl ((2R,3R)-4-((tert-Butyldiphenylsilyloxy)-3-(methoxy-methoxy)-1-phenylbutan-2-yl)carbamate (7b)

Column chromatography (petroleum ether/EtOAc, 85:15).

Yield: 0.86 g (80%); clear oil; $[\alpha]_D^{27} +11.05$ (c 2.63, CHCl₃).

IR (thin film): 3070, 3027, 2930, 2891, 2857, 1713, 1603, 1589 cm⁻¹.

^1H NMR (CDCl_3 , 400 MHz): δ = 7.70–7.68 (m, 4 H), 7.45–7.38 (m, 6 H), 7.25–7.16 (m, 5 H), 5.38 (d, J = 12.0 Hz, 1 H), 4.65 (br s, 2 H), 4.17 (d, J = 8.0 Hz, 1 H), 3.84–3.80 (m, 1 H), 3.71–3.61 (m, 2 H), 3.32 (s, 3 H), 2.82 (d, J = 8.0 Hz, 2 H), 1.35 (s, 9 H), 1.08 (s, 9 H).

^{13}C NMR (CDCl_3 , 125 MHz): δ = 155.6, 138.4, 135.7, 135.7, 133.0, 129.9, 129.9, 129.2, 128.3, 127.8, 127.8, 126.2, 96.6, 93.6, 78.7, 64.4, 63.5, 55.7, 52.9, 38.6, 36.9, 28.4, 26.9, 19.2.

HRMS (ESI–TOF): m/z [$M + \text{H}$] $^+$ calcd for $\text{C}_{33}\text{H}_{46}\text{NO}_5\text{Si}$: 564.3145; found: 564.3149.

Silyl Deprotection; General Procedure

TBAF (1 M in THF, 1.94 mL, 1.94 mmol) was added to a stirred solution of compound **7** (1.00 g, 1.77 mmol) in anhydrous THF (15 mL) at 0 °C and the solution was stirred at r.t. for 2 h. On complete disappearance of starting material, the reaction was quenched with saturated aqueous NH_4Cl (30 mL) and the mixture was extracted with EtOAc (2 \times 30 mL). The combined organic phases were dried over Na_2SO_4 , filtered, concentrated under vacuum, and purified by column chromatography.

tert-Butyl ((2*R*,3*S*)-4-Hydroxy-3-(methoxymethoxy)-1-phenylbutan-2-yl)carbamate (**8a**)

Column chromatography (petroleum ether/EtOAc, 70:30).

Yield: 0.51 g (89%); clear oil; $[\alpha]_{\text{D}}^{27}$ +42.55 (c 0.79, CHCl_3).

IR (thin film): 3444, 3063, 3028, 2927, 2854, 1693, 1604 cm^{-1} .

^1H NMR (CDCl_3 , 500 MHz): δ = 7.23–7.12 (m, 5 H), 4.71 (d, J = 10.0 Hz, 1 H), 4.67–4.53 (m, 2 H), 4.05 (dd, J = 15.0, 10.0 Hz, 1 H), 3.61 (dd, J = 10.0, 5.0 Hz, 1 H), 3.46–3.44 (m, 1 H), 3.41–3.37 (m, 1 H), 3.34 (s, 3 H), 2.84–2.74 (m, 2 H), 1.33 (s, 9 H).

^{13}C NMR (CDCl_3 , 125 MHz): δ = 156.6, 137.9, 129.1, 128.6, 126.6, 97.7, 80.7, 80.0, 62.6, 55.9, 52.1, 38.3, 29.8, 28.4.

HRMS (ESI–TOF): m/z [$M + \text{Na}$] $^+$ calcd for $\text{C}_{17}\text{H}_{27}\text{NNaO}_5$: 348.1787; found: 348.1788.

tert-Butyl ((2*R*,3*R*)-4-Hydroxy-3-(methoxymethoxy)-1-phenylbutan-2-yl)carbamate (**8b**)

Column chromatography (petroleum ether/EtOAc, 70:30).

Yield: 0.52 g (90%); clear oil; $[\alpha]_{\text{D}}^{27}$ +0.47 (c 1.02, CHCl_3).

IR (thin film): 3471, 3368, 3021, 2964, 2929, 1692, 1523 cm^{-1} .

^1H NMR (CDCl_3 , 500 MHz): δ = 7.28–7.18 (m, 5 H), 4.80–4.70 (m, 3 H), 4.04 (br s, 1 H), 3.70–3.66 (m, 2 H), 3.50 (br s, 1 H), 3.45 (s, 3 H), 3.03–3.00 (m, 1 H), 2.75–2.70 (m, 1 H), 1.89 (br s, 1 H), 1.32 (s, 9 H).

^{13}C NMR (CDCl_3 , 125 MHz): δ = 156.0, 137.9, 129.3, 128.5, 126.5, 97.0, 82.4, 79.7, 62.3, 56.0, 52.0, 36.7, 29.7, 28.3.

HRMS (ESI–TOF): m/z [$M + \text{H}$] $^+$ calcd for $\text{C}_{17}\text{H}_{28}\text{NO}_5$: 326.1967; found: 326.1968.

Oxidation of Primary Alcohols; General Procedure

Pyridinium dichromate (11.56 g, 30.75 mmol) was added to the stirred solution of alcohol **8** (1.00 g, 3.07 mmol) in DMF (30 mL) and stirring was continued at r.t. for 8 h. On complete disappearance of the starting material, the reaction was quenched with water (300 mL) and extracted with Et_2O (2 \times 50 mL). The combined organic phases were washed with saturated aqueous NaHCO_3 (2 \times 30 mL) and the aqueous extracts containing the carboxylate salts were combined and acidified with saturated aqueous KHSO_4 (2 \times 50 mL) and this was ex-

tracted with Et_2O (2 \times 50 mL). The ether layers were combined, dried over Na_2SO_4 , filtered, concentrated, and purified by column chromatography.

(2*S*,3*R*)-3-((*tert*-Butoxycarbonyl)amino)-2-(methoxymethoxy)-4-phenylbutanoic Acid (**9a**)

Column chromatography (DCM/MeOH, 95:5).

Yield: 0.79 g (76%); clear oil; $[\alpha]_{\text{D}}^{27}$ +9.18 (c 0.29, CHCl_3).

IR (thin film): 3334, 2924, 2853, 1715, 1497 cm^{-1} .

^1H NMR (CDCl_3 , 500 MHz): δ = 8.03 (s, 1 H), 7.30–7.19 (m, 5 H), 5.09 (d, J = 8.0 Hz, 1 H), 4.77–4.70 (m, 2 H), 4.37 (d, J = 4.0 Hz, 1 H), 4.17 (s, 1 H), 3.46 (s, 3 H), 2.90–2.88 (m, 2 H), 1.34 (s, 9 H).

^{13}C NMR (CDCl_3 , 125 MHz): δ = 173.1, 163.2, 155.6, 137.5, 129.4, 128.7, 126.7, 96.8, 80.1, 75.1, 56.7, 54.0, 38.5, 29.8, 28.3.

HRMS (ESI–TOF): m/z [$M + \text{Na}$] $^+$ calcd for $\text{C}_{17}\text{H}_{25}\text{NNaO}_6$: 362.1580; found: 362.1558.

(2*R*,3*R*)-3-((*tert*-Butoxycarbonyl)amino)-2-(methoxymethoxy)-4-phenylbutanoic Acid (**9b**)

Column chromatography (DCM/MeOH, 95:5).

Yield: 0.80 g (78%); clear oil; $[\alpha]_{\text{D}}^{27}$ +48.55 (c 0.41, CHCl_3).

IR (thin film): 3395, 2924, 2853, 1692, 1603, 1497 cm^{-1} .

^1H NMR (CDCl_3 , 400 MHz): δ = 7.30–7.19 (m, 5 H), 5.07 (d, J = 8.0 Hz, 1 H), 4.76–4.69 (m, 2 H), 4.37 (d, J = 8.0 Hz, 1 H), 4.16 (br s, 1 H), 3.45 (s, 3 H), 2.88 (d, J = 8.0 Hz, 2 H), 1.33 (s, 9 H).

^{13}C NMR (CDCl_3 , 100 MHz): δ = 173.4, 163.4, 155.3, 137.5, 129.3, 128.3, 126.4, 96.6, 79.6, 56.2, 53.3, 36.9, 36.0, 31.8, 29.7, 28.2, 28.0.

HRMS (ESI–TOF): m/z [$M + \text{Na}$] $^+$ calcd for $\text{C}_{17}\text{H}_{25}\text{NNaO}_6$: 362.1580; found: 362.1584.

Peptide Coupling of **9**

Compound **9** (0.19 g, 0.55 mmol) was dissolved in anhydrous DCM (10 mL) and the solution was cooled in an ice bath followed by addition of EDC-HCl (0.21 g, 1.12 mmol) and HOBt (0.15 g, 1.12 mmol) and then stirred for 20 min. H-Leu-OBn (0.17 g, 0.55 mmol) was added to the reaction mixture followed by DIPEA (0.20 mL, 1.23 mmol) and the mixture was stirred at r.t. for 6 h. On complete disappearance of starting material, the organic layer was washed with aqueous citric acid (3 \times 15 mL) and 2 M aqueous NaHCO_3 (3 \times 15 mL). The organic layers were combined, dried over Na_2SO_4 , filtered, concentrated, and purified by column chromatography.

Benzyl ((2*S*,3*R*)-3-((*tert*-Butoxycarbonyl)amino)-2-(methoxymethoxy)-4-phenylbutanoyl)-L-leucinate (**10a**)

Column chromatography (petroleum ether/EtOAc, 70:30).

Yield: 0.24 g (82%); white solid; $[\alpha]_{\text{D}}^{27}$ +32.23 (c 0.69, CHCl_3); mp 99–101 °C.

IR (thin film): 3333, 3277, 3063, 3030, 2961, 2929, 2873, 1748, 1688, 1650, 1547, 1524 cm^{-1} .

^1H NMR (CDCl_3 , 400 MHz): δ = 7.31–7.09 (m, 10 H), 6.96 (d, J = 8.7 Hz, 1 H), 5.19 (d, J = 9.9 Hz, 1 H), 5.11–5.04 (m, 2 H), 4.69–4.62 (m, 3 H), 4.17 (br s, 1 H), 4.06 (m, 1 H), 3.35 (s, 3 H), 2.82 (dd, J = 13.7, 5.4 Hz, 1 H), 2.59–2.54 (m, 1 H), 1.62–1.50 (m, 3 H), 1.23 (s, 9 H), 0.86 (d, J = 4.3 Hz, 6 H).

^{13}C NMR (CDCl_3 , 125 MHz): δ = 172.5, 170.5, 155.0, 137.8, 135.3, 129.4, 128.7, 128.6, 128.4, 128.4, 126.5, 115.5, 96.9, 79.2, 78.1, 67.2, 56.7, 53.3, 50.4, 41.5, 37.5, 29.8, 28.3, 24.9, 22.9, 21.8.

HRMS (ESI-TOF): m/z [M + H]⁺ calcd for C₃₀H₄₃N₂O₇: 543.3070; found: 543.3079.

Benzyl ((2R,3R)-3-((tert-Butoxycarbonyl)amino)-2-(methoxymethoxy)-4-phenylbutanoyl)-L-leucinate (10b)

Column chromatography (petroleum ether/EtOAc, 70:30).

Yield: 0.24 g (82%); white solid; [α]_D²⁷ +14.68 (c 0.68, CHCl₃); mp 98–99 °C.

IR (thin film): 3348, 3306, 3030, 2957, 2929, 1738, 1693, 1654, 1524 cm⁻¹.

¹H NMR (CDCl₃, 500 MHz): δ = 7.33–7.17 (m, 10 H), 6.87 (d, J = 5.0 Hz, 1 H), 5.22–5.13 (m, 3 H), 4.71–4.61 (m, 3 H), 4.34–4.23 (m, 2 H), 3.37 (s, 3 H), 2.93–2.55 (m, 2 H), 1.73–1.56 (m, 3 H), 1.34 (s, 9 H), 0.93 (d, J = 5.0 Hz, 6 H).

¹³C NMR (CDCl₃, 125 MHz): δ = 172.6, 169.9, 155.6, 138.0, 135.4, 129.5, 128.7, 128.5, 128.4, 126.4, 96.9, 79.3, 78.8, 67.3, 56.4, 54.1, 50.7, 40.8, 36.8, 29.8, 28.4, 25.1, 22.9, 21.8.

HRMS (ESI-TOF): m/z [M + H]⁺ calcd for C₃₀H₄₃N₂O₇: 543.3070; found: 543.3079.

Procedure for Hydrogenolysis of 10

To a stirred solution of **10** (0.13 g, 0.24 mmol) in anhydrous MeOH (10 mL), Pd/C (10 mol%) was added and the mixture was stirred vigorously for 3 h at r.t. under H₂. On complete disappearance of starting material, the reaction mixture was filtered through a Celite® pad, solvent was removed under vacuum and the residue was purified by column chromatography.

((2S,3R)-3-((tert-Butoxycarbonyl)amino)-2-(methoxymethoxy)-4-phenylbutanoyl)-L-leucine (11a)

Column chromatography (CH₂Cl₂/MeOH, 95:5).

Yield: 0.10 g (92%); clear oil; [α]_D²⁷ +21.11 (c 0.36, CHCl₃).

IR (thin film): 3333, 2925, 2854, 1714, 1529, 1454 cm⁻¹.

¹H NMR (CDCl₃, 500 MHz): δ = 7.73 (d, J = 10.0 Hz, 1 H), 7.30–7.22 (m, 5 H), 6.84 (d, J = 5.0 Hz, 1 H), 4.88–4.75 (m, 2 H), 4.56 (d, J = 5.2 Hz, 1 H), 4.14–4.08 (m, 2 H), 3.47 (s, 3 H), 2.98–2.86 (m, 2 H), 1.73–1.54 (m, 3 H), 1.29 (s, 9 H), 0.92–0.89 (m, 6 H).

¹³C NMR (CDCl₃, 125 MHz): δ = 176.4, 169.6, 157.5, 137.8, 129.6, 128.8, 126.8, 97.1, 81.8, 78.0, 57.1, 56.1, 50.5, 43.3, 39.7, 29.8, 28.1, 25.0, 22.8, 22.4.

HRMS (ESI-TOF): m/z [M + Na]⁺ calcd for C₂₃H₃₆N₂NaO₇: 475.2420; found: 475.2452.

((2R,3R)-3-((tert-Butoxycarbonyl)amino)-2-(methoxymethoxy)-4-phenylbutanoyl)-L-leucine (11b)

Column chromatography (DCM/MeOH, 95:5).

Yield: 0.10 g (92%); clear oil; [α]_D²⁷ +25.37 (c 0.66, CHCl₃).

IR (thin film): 3300, 2954, 2740, 1730, 1520 cm⁻¹.

¹H NMR (CDCl₃, 400 MHz): δ = 7.25–7.10 (m, 5 H), 5.24 (br s, 1 H), 4.63 (m, 2 H), 4.31–4.14 (m, 2 H), 3.35 (s, 3 H), 2.94–2.73 (m, 2 H), 1.79–1.53 (m, 3 H), 1.24 (s, 9 H), 0.93–0.83 (m, 6 H).

¹³C NMR (CDCl₃, 100 MHz): δ = 175.7, 175.4, 170.6, 169.6, 157.3, 155.8, 138.3, 137.9, 129.6, 129.4, 128.4, 126.4, 96.6, 96.4, 81.1, 79.5, 79.0, 78.4, 56.4, 55.8, 54.2, 50.7, 50.4, 41.2, 37.4, 36.7, 32.0, 29.8, 28.3, 28.0, 25.1, 23.1, 22.8, 21.7, 14.2.

HRMS (ESI-TOF): m/z [M + Na]⁺ calcd for C₂₃H₃₆N₂NaO₇: 475.2420; found: 475.2452.

Acidolysis Reaction; General Procedure

HCl (6 M in EtOAc, 0.50 mL) was added to **11** (0.083 g, 0.18 mmol), **14** (0.092 g, 0.15 mmol) or **21** (0.050 g, 0.14 mmol) at 0 °C and the mixture was stirred at r.t. for 4 h. On complete disappearance of starting material, solvent was removed under vacuum and the white residual solid was triturated 3 to 4 times with cold EtOAc (5 mL).

((2S,3R)-3-Amino-2-hydroxy-4-phenylbutanoyl)-L-leucine (1a)

Yield: 0.058 g (94%); white solid; [α]_D²⁷ –15.83 (c 0.24, CH₃OH); mp 212–215 °C {lit.¹⁹ [α]_D²⁰ –15.2 (c 0.83, 1 M HCl); mp 210–214 °C}.

IR (thin film): 3737, 2953, 1725, 1660, 1555, 1518, 1492 cm⁻¹.

¹H NMR (D₂O, 500 MHz): δ = 7.48–7.36 (m, 5 H), 4.41 (dd, J = 10.0, 5.1 Hz, 1 H), 4.33 (d, J = 5.0 Hz, 1 H), 3.89–3.85 (m, 1 H), 3.19 (dd, J = 15.0, 5.0 Hz, 1 H), 2.97 (dd, J = 15.0, 10.0 Hz, 1 H), 1.79–1.67 (m, 3 H), 0.97 (d, J = 5.0 Hz, 3 H), 0.94 (d, J = 5.0 Hz, 3 H).

¹³C NMR (D₂O, 125 MHz): δ = 176.3, 172.7, 134.9, 129.4, 129.2, 127.7, 69.5, 54.9, 51.6, 39.1, 34.8, 24.5, 22.0, 20.7.

HRMS (ESI-TOF): m/z [M + H]⁺ calcd for C₁₆H₂₅N₂O₄: 309.1814; found: 309.1811.

((2R,3R)-3-Amino-2-hydroxy-4-phenylbutanoyl)-L-leucine (1b)

Yield: 0.060 g (97%); white solid; [α]_D²⁷ +5.82 (c 0.38, H₂O); mp 226–228 °C {lit.²⁸ [α]_D²⁰ +5.90 (c 0.38 H₂O); mp 228–230 °C}.

IR (thin film): 3394, 2926, 1739, 1651, 1454 cm⁻¹.

¹H NMR (CD₃OD, 500 MHz): δ = 7.26–7.18 (m, 5 H), 4.38 (br s, 2 H), 4.08 (d, J = 5.0 Hz, 2 H), 3.72 (d, J = 10.0 Hz, 1 H), 3.02–3.00 (m, 1 H), 2.83–2.79 (m, 1 H), 1.65–1.61 (m, 3 H), 0.90 (d, J = 5.0 Hz, 3 H), 0.86 (d, J = 5.0 Hz, 3 H).

¹³C NMR (CD₃OD, 125 MHz): δ = 173.6, 173.2, 137.0, 130.4, 130.0, 128.4, 72.0, 62.5, 56.9, 52.0, 41.0, 34.3, 25.9, 23.2, 21.7, 14.4.

HRMS (ESI-TOF): m/z [M + H]⁺ calcd for C₁₆H₂₅N₂O₄: 309.1814; found: 309.1812.

((2S,3R)-3-Amino-2-hydroxy-4-phenylbutanoyl)-L-valyl-L-phenylalanine (2)

Yield: 0.068 g (95%); white solid; [α]_D²⁷ –12.20 (c 1.02, H₂O); mp 187–189 °C {lit.¹⁹ [α]_D²⁰ –11.9 (c 1.00, HOAc); 188–191 °C}.

IR (thin film): 2924, 2853, 1732, 1647, 1456 cm⁻¹.

¹H NMR (DMSO-*d*₆, 500 MHz): δ = 8.41 (d, J = 10.0 Hz, 1 H), 8.06 (br s, 1 H), 7.81 (d, J = 10.0 Hz, 1 H), 7.34–7.11 (m, 10 H), 6.83 (br s, 1 H), 4.43–4.39 (m, 1 H), 4.19–4.16 (m, 1 H), 4.02 (s, 1 H), 3.55 (s, 1 H), 3.06–2.90 (m, 4 H), 2.02–1.98 (m, 1 H), 0.84–0.82 (m, 6 H).

¹³C NMR (DMSO-*d*₆, 125 MHz): δ = 172.6, 170.5, 137.5, 136.4, 129.5, 129.1, 128.6, 128.1, 126.9, 126.4, 68.2, 57.2, 54.2, 53.5, 36.5, 34.6, 30.7, 29.0, 19.0, 18.0.

HRMS (ESI-TOF): m/z [M + H]⁺ calcd for C₂₄H₃₂N₃O₅: 442.2342; found: 442.2343.

(3S,4R)-4-Amino-3-hydroxy-5-phenylpentanoic Acid (3)

Yield: 0.032 g (98%); clear oil; [α]_D²⁷ –1.50 (c 0.13, MeOH).

IR (thin film): 3405, 2925, 2854, 1737, 1458 cm⁻¹.

¹H NMR (D₂O, 500 MHz): δ = 7.44–7.33 (m, 5 H), 4.18–4.14 (m, 1 H), 3.62–3.58 (m, 1 H), 3.15 (dd, J = 15.0, 5.0 Hz, 1 H), 2.89 (dd, J = 15.0, 10.0 Hz, 1 H), 2.82–2.78 (m, 1 H), 2.64 (dd, J = 15.0, 10.0 Hz, 1 H).

¹³C NMR (D₂O, 125 MHz): δ = 174.8, 135.1, 129.4, 129.2, 127.7, 66.8, 56.6, 38.7, 35.3.

HRMS (ESI-TOF): m/z [M + H]⁺ calcd for C₁₁H₁₆NO₃: 210.1130; found: 210.1129.

Synthesis of Dipeptide Boc-Val-Phe-OMe

Boc-Val-OH (0.20 g, 0.92 mmol) was dissolved in anhydrous DCM (10 mL) and the solution was cooled in an ice bath followed by addition of EDC-HCl (0.35 g, 1.84 mmol) and HOBt (0.24 g, 1.84 mmol) and the mixture was stirred for 20 min. HCl-H₂N-Phe-OMe (0.19 g, 0.92 mmol) was added to the reaction mixture followed by DIPEA (0.35 mL, 2.03 mmol) and the mixture was stirred at r.t. for 6 h. On complete disappearance of starting material, the organic layer was washed with aqueous citric acid (3 × 15 mL) and 2 M aqueous NaHCO₃ (3 × 15 mL). The organic layers were combined, dried over Na₂SO₄, filtered, concentrated under vacuum, and purified by column chromatography.

Methyl (*tert*-Butoxycarbonyl)-L-valyl-L-phenylalaninate (12)

Column chromatography (petroleum ether/EtOAc, 70:30).

Yield: 0.31 g (90%); white solid; [α]_D²⁷ +30.38 (c 0.88, CHCl₃); mp 101–103 °C.

IR (thin film): 3361, 3287, 3094, 2958, 2929, 2871, 1746, 1691, 1656, 1567, 1514 cm⁻¹.

¹H NMR (CDCl₃, 500 MHz): δ = 7.28–7.20 (m, 3 H), 7.09 (d, *J* = 7.3 Hz, 2 H), 6.40 (br s, 1 H), 5.05 (br s, 1 H), 4.85 (dd, *J* = 15.0, 5.0 Hz, 1 H), 3.90 (m, 1 H), 3.68 (d, *J* = 1.4 Hz, 3 H), 3.10–3.07 (m, 2 H), 2.08–2.04 (m, 1 H), 1.43 (s, 9 H), 0.90 (d, *J* = 5.0 Hz, 3 H), 0.84 (d, *J* = 5.0 Hz, 3 H).

¹³C NMR (CDCl₃, 125 MHz): δ = 171.8, 171.3, 155.8, 135.8, 129.3, 128.7, 127.2, 79.9, 59.9, 53.23, 52.3, 38.0, 30.9, 28.4, 19.2, 17.7.

HRMS (ESI-TOF): m/z [M + Na]⁺ calcd for C₂₀H₃₀N₂NaO₅: 401.2052; found: 401.2052.

Synthesis of Tripeptide 13

TFA (1.00 mL) was added to a stirred solution of Boc-Val-Phe-OMe (0.22 g, 0.58 mmol) in anhydrous DCM (4 mL) at 0 °C and the mixture was stirred for 30 min. After completion of the reaction as observed in TLC, the solvent was removed under vacuum with addition of DCM (5 mL, 3 to 4 times). The residue (0.20 g, 0.58 mmol) was dissolved in anhydrous DCM (10 mL) in an ice bath, followed by addition of EDC-HCl (0.22 g, 1.18 mmol) and HOBt (0.15 g, 1.18 mmol) and stirred for 20 min. Boc deprotected dipeptide **12** (0.22 g, 0.58 mmol) was added to the reaction mixture followed by DIPEA (0.20 mL, 1.30 mmol). The reaction mixture was stirred at r.t. for a further 6 h. On complete disappearance of starting material, the organic layer was washed with aqueous citric acid (3 × 15 mL) and 2 M aqueous NaHCO₃ (3 × 15 mL). The organic layers were combined, dried over Na₂SO₄, filtered, concentrated, and purified by column chromatography.

Methyl ((2*S*,3*R*)-3-((*tert*-Butoxycarbonyl)amino)-2-(methoxymethoxy)-4-phenylbutanoyl)-L-valyl-L-phenylalaninate (13)

Column chromatography (petroleum ether/EtOAc, 70:30).

Yield: 0.24 g (70%); white solid; [α]_D²⁷ +18.26 (c 0.56, CHCl₃); mp 135–137 °C.

IR (thin film): 3295, 3064, 3029, 2923, 2854, 1747, 1692, 1650, 1531, 1455 cm⁻¹.

¹H NMR (CDCl₃, 500 MHz): δ = 7.22–7.13 (m, 9 H), 7.05–7.01 (m, 3 H), 6.28 (d, *J* = 5.1 Hz, 1 H), 5.10 (d, *J* = 9.9 Hz, 1 H), 4.76 (dd, *J* = 10.0, 5.0 Hz, 1 H), 4.64–4.60 (m, 2 H), 4.21 (dd, *J* = 10.0, 5.0 Hz, 2 H), 4.00 (d, *J* = 2.0 Hz, 1 H), 3.65 (s, 3 H), 3.34 (s, 3 H), 3.06 (dd, *J* = 10.0, 5.0 Hz,

1 H), 2.97 (dd, *J* = 10.0, 5.0 Hz, 1 H), 2.82 (dd, *J* = 10.0, 5.0 Hz, 1 H), 2.63 (dd, *J* = 13.6, 10.0 Hz, 1 H), 2.06–2.04 (m, 1 H), 1.24 (s, 9 H), 0.86 (d, *J* = 6.6 Hz, 3 H), 0.81 (d, *J* = 6.5 Hz, 3 H).

¹³C NMR (CDCl₃, 125 MHz): δ = 171.7, 170.5, 154.9, 137.8, 135.7, 129.4, 129.3, 128.7, 128.5, 127.3, 126.5, 97.4, 79.3, 78.8, 58.0, 56.8, 53.5, 53.3, 52.4, 38.1, 37.9, 31.1, 29.8, 28.4, 19.3, 17.8.

HRMS (ESI-TOF): m/z [M + Na]⁺ calcd for C₃₂H₄₆N₃O₈: 600.3285; found: 600.3282.

Procedure for Hydrolysis of 13

LiOH (0.030 g, 0.48 mmol) was added to a stirred solution of **13** (0.24 g, 0.40 mmol) in MeOH/H₂O (4:1, 10 mL) at 0 °C and the reaction mixture was stirred for 1 h at the same temperature. After the disappearance of starting material as observed in TLC, the reaction was quenched with saturated aqueous KHSO₄ (10 mL) and the free acid was extracted with EtOAc (2 × 40 mL). The organic layers were combined, dried over Na₂SO₄, filtered, concentrated under vacuum, and purified by column chromatography.

((2*S*,3*R*)-3-((*tert*-Butoxycarbonyl)amino)-2-(methoxymethoxy)-4-phenylbutanoyl)-L-valyl-L-phenylalanine (14)

Column chromatography (DCM/MeOH, 95:5).

Yield: 0.22 g (94%); white solid; [α]_D²⁷ +15.69 (c 0.86, CHCl₃); mp 109–110 °C.

IR (thin film): 3312, 3064, 3029, 2962, 2925, 2854, 1716, 1650, 1524 cm⁻¹.

¹H NMR (CDCl₃, 500 MHz): δ = 7.47 (br s, 1 H), 7.27–7.13 (m, 10 H), 6.07 (br s, 1 H), 5.07 (d, *J* = 5.0 Hz, 1 H), 4.81 (br s, 1 H), 4.61–4.59 (m, 2 H), 4.34–4.07 (m, 3 H), 3.39 (s, 3 H), 2.90–2.71 (m, 3 H), 2.04 (br s, 1 H), 1.76–1.61 (m, 1 H), 1.25 (s, 9 H), 0.89–0.84 (m, 6 H).

¹³C NMR (CDCl₃, 125 MHz): δ = 175.7, 174.2, 173.7, 171.0, 170.7, 156.6, 155.1, 137.9, 137.6, 136.1, 129.6, 128.6, 127.1, 126.7, 115.5, 97.3, 96.9, 81.1, 79.6, 78.5, 58.4, 58.2, 56.9, 55.7, 53.7, 53.3, 39.7, 38.5, 37.8, 31.8, 31.1, 29.8, 28.3, 27.9, 20.8, 19.4, 18.3.

HRMS (ESI-TOF): m/z [M + H]⁺ calcd for C₃₁H₄₄N₃O₈: 586.3128; found: 586.3121.

Asymmetric α-Hydroxylation of Aldehyde 4

L-Proline (0.13 g, 1.14 mmol, 30 mol%) and nitrosobenzene (0.44 g, 4.18 mmol) were added to a stirred solution of **4** (1.00 g, 3.80 mmol) in anhydrous DMSO (10 mL) at 15 °C and the mixture was stirred for 3 h at the same temperature. After 3 h the reaction was cooled to 0 °C and phosphorane Ph₃P=CHCO₂Et (2.65 g, 7.60 mmol) in DCM (10 mL) was added and the reaction mixture was stirred for a further 2 h at 0 °C. On complete disappearance of starting material, the reaction was quenched with saturated aqueous NH₄Cl (30 mL) and the mixture was extracted with DCM (2 × 30 mL). The combined organic phases were washed with brine (30 mL), dried over Na₂SO₄, filtered, and concentrated under vacuum. The crude aminohydroxylated product was taken as such to the next step, leading to the cleavage of O–N bond.

Cu(OAc)₂ (0.17 g, 0.96 mmol) was added to a stirred solution of the above product (1.43 g, 3.24 mmol) in EtOH (10 mL) and the mixture stirred at r.t. for 6 h. On complete disappearance of starting material, the reaction was quenched with saturated aqueous NH₄Cl (20 mL) and the mixture was extracted with DCM (2 × 20 mL). The combined organic phases were washed with brine (30 mL), dried over Na₂SO₄, filtered, concentrated under vacuum, and purified by column chromatography.

Ethyl (4*S*,5*R*,*E*)-5-((*tert*-Butoxycarbonyl)amino)-4-hydroxy-6-phenylhex-2-enoate (15)

Column chromatography (petroleum ether/EtOAc, 80:20).

Yield: 0.80 g (70%); clear oil; $[\alpha]_D^{27} -3.91$ (c 0.23, CHCl₃).

IR (thin film): 3355, 2926, 1729, 1683, 1524 cm⁻¹.

¹H NMR (CDCl₃, 500 MHz): δ = 7.31–7.17 (m, 5 H), 6.98 (dd, J = 15.0, 5.0 Hz, 1 H), 6.15 (d, J = 15.0, 5.0 Hz, 1 H), 4.62 (d, J = 10.0 Hz, 1 H), 4.43 (br s, 1 H), 4.21 (q, J = 5.0 Hz, 2 H), 4.02 (s, 1 H), 3.81 (s, 1 H), 2.84–2.77 (m, 2 H), 1.36 (s, 9 H), 1.29 (t, J = 5.0 Hz, 3 H).

¹³C NMR (CDCl₃, 125 MHz): δ = 166.4, 157.0, 146.0, 137.4, 129.2, 128.8, 126.9, 122.8, 80.5, 73.6, 60.6, 57.0, 36.2, 29.8, 28.3, 14.3.

HRMS (ESI-TOF): m/z [M + Na]⁺ calcd for C₁₉H₂₇NNaO₅: 372.1787; found: 372.1772.

Procedure for Oxazolidine Protection of 15

A catalytic amount of *p*-TsOH (0.09 g, 0.57 mmol) and dimethoxypropane (1.11 mL, 8.59 mmol) were added to a stirred solution of **15** (1.00 g, 2.86 mmol) in anhydrous DCM (20 mL) and the mixture was stirred at r.t. for 2 h. On complete disappearance of starting material, the reaction was quenched with saturated aqueous NaHCO₃ (20 mL) and the crude product was extracted with DCM (2 × 30 mL). The combined organic phases were dried over Na₂SO₄, filtered, concentrated under vacuum, and purified by column chromatography.

***tert*-Butyl (4*R*,5*S*)-4-Benzyl-5-((*E*)-3-ethoxy-3-oxoprop-1-en-1-yl)-2,2-dimethyloxazolidine-3-carboxylate (16)**

Column chromatography (petroleum ether/EtOAc, 85:15).

Yield: 0.95 g (85%); clear oil; $[\alpha]_D^{27} -13.77$ (c 0.80, CHCl₃).

IR (thin film): 2978, 2930, 1723, 1701, 1604 cm⁻¹.

¹H NMR (CDCl₃, 400 MHz): δ = 7.29–7.10 (m, 5 H), 6.61–6.54 (m, 1 H), 6.16–6.12 (m, 1 H), 4.69 (br s, 1 H), 4.47–4.24 (m, 1 H), 4.19–4.10 (m, 2 H), 3.22 (dd J = 12.0, 4.0 Hz, 1 H), 2.91–2.79 (m, 1 H), 2.71–2.66 (m, 1 H), 1.56 (s, 3 H), 1.52 (s, 3 H), 1.46 (s, 3 H), 1.37 (s, 3 H), 1.28–1.22 (m, 6 H).

¹³C NMR (CDCl₃, 125 MHz): δ = 165.8, 151.9, 151.5, 141.7, 141.6, 138.2, 137.1, 130.0, 129.9, 128.4, 128.2, 126.3, 126.2, 122.6, 93.7, 93.0, 80.4, 80.1, 75.7, 75.4, 63.2, 61.7, 61.3, 60.6, 60.5, 37.4, 36.6, 28.6, 28.5, 28.0, 27.4, 27.0, 25.2, 24.0, 14.3, 14.2.

HRMS (ESI-TOF): m/z [M + Na]⁺ calcd for C₂₂H₃₁NNaO₅: 412.2100; found: 412.2105.

LiBH₄ Reduction of 16

LiBH₄ (0.17 g, 7.70 mmol) was added to a stirred solution of **16** (1.00 g, 2.57 mmol) in anhydrous THF (20 mL) at 0 °C and the mixture was stirred at r.t. for 8 h. On complete disappearance of starting material, the reaction was quenched with saturated aqueous NaHCO₃ (20 mL). The crude product was extracted with EtOAc (2 × 30 mL) and the combined organic phases were dried over Na₂SO₄, filtered, concentrated under vacuum, and purified by column chromatography.

***tert*-Butyl (4*R*,5*S*)-4-Benzyl-5-(3-hydroxypropyl)-2,2-dimethyloxazolidine-3-carboxylate (17)**

Column chromatography (petroleum ether/EtOAc, 70:30).

Yield: 0.72 g (80%); clear oil; $[\alpha]_D^{27} +21.04$ (c 0.51, CHCl₃).

IR (thin film): 3445, 3062, 3027, 2928, 2856, 1696, 1604 cm⁻¹.

¹H NMR (CDCl₃, 400 MHz): δ = 7.29–7.14 (m, 5 H), 4.26–4.11 (m, 1 H), 4.05–3.86 (m, 1 H), 3.52–3.50 (m, 2 H), 3.22–3.18 (dd, J = 12.0, 4.0 Hz, 1 H), 2.92–2.81 (m, 2 H), 1.66–1.49 (m, 9 H), 1.43 (s, 4 H), 1.32 (s, 4 H), 1.24 (s, 2 H).

¹³C NMR (CDCl₃, 100 MHz): δ = 152.0, 151.7, 139.1, 139.1, 129.5, 129.3, 128.5, 128.3, 126.2, 126.1, 93.0, 92.4, 80.1, 79.7, 62.4, 60.9, 60.8, 36.5, 35.9, 29.9, 29.8, 28.5, 28.4, 28.1, 27.5, 26.2, 25.0, 23.8.

HRMS (ESI-TOF): m/z [M + Na]⁺ calcd for C₂₀H₃₁NNaO₄: 372.2151; found: 372.2151.

Oxidation of Primary Alcohols

IBX (0.69 g, 2.47 mmol) was added to a solution of **17** (0.72 g, 2.06 mmol) in DMSO (10 mL) at r.t. and the mixture was stirred for 3 h. On complete disappearance of starting material, the reaction was quenched with saturated aqueous NaHCO₃ (50 mL) and the mixture was extracted with EtOAc (2 × 30 mL). The combined organic phases washed with brine (30 mL) and dried over Na₂SO₄, filtered, concentrated under reduced pressure, and purified by column chromatography.

***tert*-Butyl (4*R*,5*S*)-4-Benzyl-2,2-dimethyl-5-(3-oxopropyl)oxazolidine-3-carboxylate (18)**

Column chromatography (petroleum ether/EtOAc, 80:20).

Yield: 0.63 g (88%); clear oil; $[\alpha]_D^{27} +15.25$ (c 0.72, CHCl₃).

IR (thin film): 2927, 2854, 2719, 1727, 1696, 1604 cm⁻¹.

¹H NMR (CDCl₃, 400 MHz): δ = 9.60, 9.57 (s, 1 H), 7.30–7.15 (m, 5 H), 4.25–4.11 (m, 1 H), 3.99–3.96 (m, 1 H), 2.96–2.79 (m, 2 H), 2.44–2.29 (m, 1 H), 2.23–2.08 (m, 1 H), 1.95–1.79 (m, 1 H), 1.69–1.63 (m, 3 H), 1.54–1.50 (m, 4 H), 1.46–1.44 (m, 5 H), 1.35 (s, 4 H).

¹³C NMR (CDCl₃, 100 MHz): δ = 201.3, 152.0, 151.6, 138.9, 129.4, 129.3, 128.6, 128.4, 126.3, 126.2, 92.9, 92.4, 80.1, 79.8, 76.4, 76.2, 60.8, 60.6, 40.7, 36.5, 35.9, 29.8, 28.5, 28.4, 28.1, 27.4, 25.0, 23.8, 22.1.

HRMS (ESI-TOF): m/z [M + Na]⁺ calcd for C₂₀H₂₉NNaO₄: 370.1994; found: 370.1992.

Synthesis of 20 from Diol 19

NaIO₄ (0.56 g, 2.62 mmol) was added to a stirred solution of diol **19** (0.48 g, 1.31 mmol) in DCM/MeOH (1:1, 10 mL) and the mixture was stirred at r.t. for 4 h. On complete disappearance of starting material, the reaction mixture was filtered and washed with brine (20 mL). The crude product was extracted with EtOAc (2 × 30 mL) and dried over Na₂SO₄, filtered, concentrated under reduced pressure, and purified by column chromatography.

***tert*-Butyl (4*R*,5*S*)-4-Benzyl-2,2-dimethyl-5-(2-oxoethyl)oxazolidine-3-carboxylate (20)**

Column chromatography (petroleum ether/EtOAc, 80:20).

Yield: 0.38 g (85%); clear oil; $[\alpha]_D^{27} -2.82$ (c 0.49, CHCl₃).

IR (thin film): 3439, 2975, 2931, 1728, 1697, 1495, 1455 cm⁻¹.

¹H NMR (CDCl₃, 500 MHz): δ = 9.55 (s, 1 H), 7.30–7.19 (m, 5 H), 4.53–4.40 (m, 1 H), 3.84 (br s, 1 H), 3.32 (d, J = 5.0 Hz, 1 H), 2.76–2.71 (m, 1 H), 2.52–2.46 (m, 1 H), 2.21–2.17 (m, 1 H), 1.39 (s, 15 H).

¹³C NMR (CDCl₃, 100 MHz): δ = 200.0, 152.2, 151.7, 138.2, 137.3, 129.7, 129.4, 128.7, 126.9, 95.1, 94.4, 80.4, 74.3, 73.5, 63.3, 48.7, 48.2, 43.7, 39.8, 37.7, 29.8, 28.6, 26.9.

HRMS (ESI-TOF): m/z [M + Na]⁺ calcd for C₁₉H₂₇NNaO₄: 356.1838; found: 356.1841.

Synthesis of **21** from **20**

Pyridinium dichromate (0.45 g, 1.20 mmol) was added to a stirred solution of **20** (0.10 g, 0.30 mmol) in DMF (10 mL) and stirring was continued at r.t. for 8 h. On complete disappearance of the starting material, the reaction was quenched with water (100 mL), the crude product was extracted with Et₂O (2 × 40 mL) and the combined organic phases were further extracted with saturated aqueous NaHCO₃ (2 × 30 mL). The aqueous extracts containing the carboxylate salt were combined and acidified with saturated aqueous KHSO₄ (2 × 40 mL) and extracted with Et₂O (2 × 50 mL). The combined organic phases were dried over Na₂SO₄, filtered, concentrated under reduced pressure, and purified by column chromatography.

2-((4*R*,5*S*)-4-Benzyl-3-(*tert*-butoxycarbonyl)-2,2-dimethyloxazolidin-5-yl)acetic Acid (**21**)

Column chromatography (DCM/MeOH, 95:05).

Yield: 0.07 g (67%); clear oil; [α]_D²⁷ -5.77 (c 0.48, CHCl₃).

IR (thin film): 3478, 2976, 2927, 2854, 1698, 1495, 1455 cm⁻¹.

¹H NMR (CDCl₃, 500 MHz): δ = 7.29–7.25 (m, 2 H), 7.21–7.20 (m, 3 H), 4.46–4.34 (m, 1 H), 3.89–3.83 (m, 1 H), 3.26 (br s, 1 H), 2.88–2.66 (m, 1 H), 2.49–2.44 (m, 1 H), 2.23–2.19 (m, 1 H), 1.37 (s, 15 H).

¹³C NMR (CDCl₃, 100 MHz): δ = 175.6, 152.3, 138.2, 137.5, 129.4, 128.7, 126.8, 95.1, 94.5, 80.5, 75.7, 75.0, 72.8, 63.4, 59.9, 40.0, 38.0, 28.7, 28.5, 27.3.

HRMS (ESI–TOF): *m/z* [M + Na]⁺ calcd for C₁₉H₂₇NNaO₅: 372.1787; found: 372.1789.

Synthesis of Aldehyde **4**

To a stirred solution of methoxymethyltriphenyl-phosphonium chloride (2.05 g, 6.02 mmol) and *t*-BuOK (0.58 g, 5.21 mmol) in anhydrous THF (10 mL), HN-Boc-*D*-phenyl-alaninal (1.00 g, 4.01 mmol) in anhydrous THF (10 mL) was added slowly at -10 °C and the mixture was stirred vigorously for 2 h at the same temperature. On complete disappearance of starting material, the reaction was quenched with saturated aqueous NH₄Cl (30 mL) and the mixture was extracted with EtOAc (2 × 30 mL). The combined organic phases were dried over Na₂SO₄, filtered, concentrated under vacuum, and purified by column chromatography using 90:10 petroleum ether/EtOAc as eluent.

HCl (2 M, 5 mL) was added to the above compound (1.02 g, 3.68 mmol) in THF (5 mL) at 0 °C and the mixture was stirred vigorously at 0 °C for 1 h. On complete disappearance of starting material, the reaction was quenched with saturated aqueous NaHCO₃ (10 mL), and the mixture was extracted with EtOAc (2 × 20 mL) and the combined organic phases containing crude product were dried over Na₂SO₄, filtered, concentrated, and purified by column chromatography.

tert-Butyl (*R*)-(4-Oxo-1-phenylbutan-2-yl)carbamate (**4**)

Column chromatography (petroleum ether/EtOAc, 80:20).

Yield: 0.85 g (81%); clear oil; [α]_D²⁷ +12.04 (c 0.51, CHCl₃).

IR (thin film): 2928, 2856, 2718, 1727, 1696, 1604 cm⁻¹.

¹H NMR (CDCl₃, 400 MHz): δ = 9.69 (s, 1 H), 7.31–7.14 (m, 5 H), 4.76 (br s, 1 H), 4.25 (br s, 1 H), 2.95–2.61 (m, 2 H), 2.58–2.48 (m, 2 H), 1.39 (s, 9 H).

¹³C NMR (CDCl₃, 100 MHz): δ = 201.1, 155.2, 137.5, 129.4, 128.7, 126.9, 79.7, 47.7, 47.5, 40.8, 31.3, 29.8, 28.4.

HRMS (ESI–TOF): *m/z* [M + Na]⁺ calcd for C₁₅H₂₁NNaO₃: 286.1419; found: 286.1421.

Funding Information

The author thanks CSIR for a Senior Research Fellowship

Acknowledgment

The author thanks his Ph.D Thesis supervisor Dr. Ramesh Ramapanicker for allowing him to work in his laboratory and for providing assistance, IIT Kanpur for providing instrumental facilities.

Supporting Information

Supporting information for this article is available online at <https://doi.org/10.1055/s-0039-1690223>.

References and Notes

- Umezawa, H.; Aoyagi, T.; Suda, H.; Hamada, M.; Takeuchi, T. *J. Antibiot.* **1976**, *29*, 97.
- Nishino, N.; Powers, J. C. *Biochemistry* **1979**, *18*, 4340.
- Suda, H.; Takita, T.; Aoyagi, T.; Umezawa, H. *J. Antibiot.* **1976**, *29*, 100.
- Nakamura, H.; Suda, H.; Takita, T.; Aoyagi, T.; Umezawa, H.; Iitaka, Y. *J. Antibiot.* **1976**, *29*, 102.
- Pulido-Cejudo, G.; Conway, B.; Proulx, P.; Brown, R.; Izaguirre, C. A. *Antiviral Res.* **1997**, *36*, 167.
- Bourinbaiar, A. S.; Lee-Huang, S.; Krasinski, K.; Borkowsky, W. *Biomed. Pharmacother.* **1994**, *48*, 55.
- Inoi, K.; Goto, S.; Nomura, S.; Isobe, K.; Nawa, A.; Okamoto, T.; Tomoda, Y. *Anticancer Res.* **1995**, *15*, 2081.
- Dzolic, E.; Varagic, V. M. *Fundam. Clin. Pharmacol.* **1987**, *1*, 307.
- Harbeson, S. L.; Rich, D. H. *Biochemistry* **1988**, *27*, 7301.
- Nagai, M.; Kojima, F.; Naganawa, H.; Hamada, M.; Aoyagi, T.; Takeuchi, T. *J. Antibiot.* **1997**, *50*, 82.
- Gogoi, N.; Boruwa, J.; Barua, N. C. *Tetrahedron Lett.* **2005**, *46*, 7581.
- Jung, D. Y.; Kang, S.; Chang, S.; Kim, Y. H. *Synlett* **2006**, 86.
- Gogoi, N.; Borah, J. C.; Boruwa, J.; Barua, N. C. *Lett. Org. Chem.* **2007**, *4*, 234.
- Feske, B. D. *Curr. Org. Chem.* **2007**, *11*, 483; and therein.
- (a) George, S.; Suryavanshi, G. S.; Sudalai, A. *Tetrahedron Lett.* **2008**, *49*, 6791; and therein. (b) Venkataramasubramanian, V.; Kiran, I. N. C.; Sudalai, A. *Synlett* **2015**, *26*, 355.
- Velmourougane, G.; Harbut, M. B.; Dalal, S.; McGowan, S.; Oellig, C. A.; Meinhardt, N.; Whisstock, J. C.; Klemba, M.; Greenbaum, D. C. *J. Med. Chem.* **2011**, *54*, 1655.
- Wasserman, H. H.; Petersen, A. K.; Xia, M. *Tetrahedron* **2003**, *59*, 6771.
- Lee, B. W.; Lee, J. H.; Jang, K. C.; Kang, J. E.; Kim, J. H.; Park, K.-M.; Park, K. H. *Tetrahedron Lett.* **2003**, *44*, 5905.
- Righi, G.; Achille, C. D.; Pescatore, G.; Bonini, C. *Tetrahedron Lett.* **2003**, *44*, 6999.
- Feske, B. D.; Stewart, J. D. *Tetrahedron: Asymmetry* **2005**, *16*, 3124.
- Kudyba, I.; Raczko, J.; Jurczak, J. *Tetrahedron Lett.* **2003**, *44*, 8685.
- Seo, Y.; Kim, H.; Chae, D. W.; Kim, Y. G. *Tetrahedron: Asymmetry* **2014**, *25*, 625.
- Lee, J. H.; Lee, B. W.; Jang, K. C.; Jeong, I.-Y.; Yang, M. S.; Lee, S. G.; Park, K. H. *Synthesis* **2003**, 829.
- Semple, J. E.; Owens, T. D.; Nguyen, K.; Levy, O. *Org. Lett.* **2000**, *2*, 2769.

- (25) Nemoto, H.; Ma, R.; Suzuki, I.; Shibuya, M. *Org. Lett.* **2000**, *2*, 4245.
- (26) Kudyba, I.; Raczek, J.; Jurczak, J. *J. Org. Chem.* **2004**, *69*, 2844.
- (27) (a) Bergmeier, S. C.; Stanchina, D. M. *J. Org. Chem.* **1999**, *64*, 2852. (b) Shang, S.; Willems, A. V.; Chauhan, S. S. *J. Pept. Sci.* **2018**, *24*, 3067.
- (28) Richter, A.; Hedberg, C. *Synthesis* **2010**, 2039.
- (29) Lee, J. H.; Kim, J. H.; Lee, B. W.; Seo, W. D.; Yang, M. S.; Park, K. H. *Bull. Korean Chem. Soc.* **2006**, *27*, 1211.
- (30) (a) see ref 15b (b) Kumar, P.; Dwivedi, N. *Acc. Chem. Res.* **2013**, *46*, 289. (c) Zhong, G. *Angew. Chem. Int. Ed.* **2003**, *42*, 4247. (d) Mukherjee, S.; Yang, J. W.; Hoffmann, S.; List, B. *Chem. Rev.* **2007**, *107*, 5471. (e) Vilaivan, T.; Bhanthumnavin, W. *Molecules* **2010**, *15*, 917. (f) Lalwani, K. G.; Sudalai, A. *Synlett* **2016**, *27*, 1339. (g) Hayashi, Y.; Yamaguchi, J.; Hibino, K.; Shoji, M. *Tetrahedron Lett.* **2003**, *44*, 8293. (h) Mangion, I. K.; MacMillan, D. W. C. *J. Am. Chem. Soc.* **2005**, *127*, 3696. (i) Momiyama, N.; Yamamoto, H. *J. Am. Chem. Soc.* **2003**, *125*, 6038. (j) Lee, L. G.; Whitesides, G. M. *J. Org. Chem.* **1986**, *51*, 25. (k) Brown, S. P.; Brochu, M. P.; Sinz, C. J.; MacMillan, D. W. C. *J. Am. Chem. Soc.* **2003**, *125*, 10808.
- (31) Janey, J. M. *Angew. Chem. Int. Ed.* **2005**, *44*, 4292.
- (32) Chacko, S.; Ramapanicker, R. *J. Org. Chem.* **2015**, *80*, 4776.
- (33) Chacko, S.; Kalita, M.; Ramapanicker, R. *Tetrahedron: Asymmetry* **2015**, *26*, 623.
- (34) Petakamsetty, R.; Jain, V. K.; Majhi, P. K.; Ramapanicker, R. *Org. Biomol. Chem.* **2015**, *13*, 8512.
- (35) Chacko, S.; Ramapanicker, R. *Tetrahedron Lett.* **2015**, *56*, 2023.
- (36) Petakamsetty, R.; Das, R. P.; Ramapanicker, R. *Tetrahedron* **2014**, *70*, 9554.
- (37) Chacko, S.; Ramapanicker, R. *Eur. J. Org. Chem.* **2012**, 7120.
- (38) Philip, A. T.; Chacko, S.; Ramapanicker, R. *Synthesis* **2013**, *45*, 1997.
- (39) Philip, A. T.; Chacko, S.; Ramapanicker, R. *J. Pept. Sci.* **2015**, *21*, 887.
- (40) Chacko, S.; Ramapanicker, R. *ChemistrySelect* **2016**, *1*, 4458.
- (41) Petakamsetty, R.; Ansari, A.; Ramapanicker, R. *Carbohydr. Res.* **2016**, *435*, 37.
- (42) Jain, V. K.; Ramapanicker, R. *Tetrahedron* **2017**, *73*, 1568.
- (43) For synthesis of aldehyde **4** see: (a) Guduru, S. K. R.; Chamakuri, S.; Raji, I. O.; MacKenzie, K. R.; Santini, C.; Young, D. W. *J. Org. Chem.* **2018**, *83*, 11777. (b) Cytlak, T.; Skibinska, M.; Kaczmarek, P.; Kazmierczak, M.; Rapp, M.; Kubicki, M.; Koroniak, H. *RSC Adv.* **2018**, *8*, 11957. (c) Shankar, P. S.; Bigotti, S.; Lazzari, P.; Manca, I.; Spiga, M.; Sani, M.; Zanda, M. *Tetrahedron Lett.* **2013**, *54*, 6137. (d) Yao, L.; Wen, J.; Liu, S.; Tan, R.; Wood, N. M.; Chen, W.; Zhang, S.; Zhang, X. *Chem. Commun.* **2016**, *52*, 2273; See also the Experimental Section 'Procedure for Synthesis of Aldehyde **4**'.
- (44) Umezawa, H.; Aoyagi, T.; Morishima, H.; Matsuzaki, M.; Hamada, M.; Takeuchi, T. *J. Antibiot.* **1970**, *23*, 259.
- (45) Moore, M. L.; Bryan, W. M.; Fakhoury, S. A.; Magaard, V. W.; Huffman, W. F.; Dayton, B. D.; Meek, T. D.; Hyland, L.; Dreyer, G. B.; Metcalf, B. W.; Strickler, J. E.; Gorniak, J. G.; Debouck, C. *Biochem. Biophys. Res. Commun.* **1989**, *159*, 420.
- (46) Liu, J.; Chen, W.; Xu, Y.; Ren, S.; Zhang, W.; Li, Y. *Bioorg. Med. Chem.* **2015**, *23*, 1963.
- (47) Si, C.-M.; Shao, L.-P.; Mao, Z.-Y.; Zhou, W.; Wei, B.-G. *Org. Biomol. Chem.* **2017**, *15*, 649.
- (48) Li, X.; Li, Y.-l.; Chen, Y.; Zou, Y.; Zhuo, X.-b.; Wu, Q.-y.; Zhao, Q.-j.; Hu, H.-g. *RSC Adv.* **2015**, *5*, 94654.
- (49) Kondekar, N. B.; Kandula, S. R. V.; Kumar, P. *Tetrahedron Lett.* **2004**, *45*, 5477.
- (50) Andres, J. M.; Pedrosa, R.; Perez, A.; Encabo, A. P. *Tetrahedron* **2001**, *57*, 8521.