

Biological Actions, Electrical Conductance and Silicon-Containing Microparticles of *Arsenicum Album* Prepared in Plastic and Glass Vials

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Abstract

Introduction According to the “silica hypothesis” formulated to explain homeopathy, the information of starting materials would be transferred to cells by silica nanoparticles detached from the glassware walls by serial dilution and agitation through epitaxy. We compared the biological activity, electrical current and silicon microparticle content (by means of scanning electron microscopy/energy-dispersive X-ray spectroscopy) of high dilutions (HDs) of arsenic prepared in plastic and glass vials to investigate the role of silica in their biological effects in vitro.

Materials and Methods Co-cultures of macrophages and yeast (*Saccharomyces cerevisiae*) were treated with different HDs of arsenic prepared in plastic and glass vials. Macrophage morphology, phagocytosis index, nitric oxide (NO), and cytokine production were evaluated.

Results Measurable amounts of silicon microparticles were detected only in the HDs prepared in glass vials, but ultra-centrifugation eliminated them. Specific and non-specific results were observed. Non-specific pro-inflammatory effects were seen in all dilutions prepared in plastic vials, including elevation of pro-inflammatory cytokines, NO and macrophage phagocytic index. Only the 200th centesimal dilution of arsenic produced specific decrease in interleukin-6 production in macrophages, and it was independent of the vial type or the presence of microparticles of silica in the medicine samples. The nature of the vials had an impact on the electric flow in the respective fluids.

Keywords

- ▶ arsenic
- ▶ macrophages
- ▶ microparticles
- ▶ conductance
- ▶ silicon
- ▶ homeopathy

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Conclusion The non-specific, pro-inflammatory effects might be attributed to organic residuals detached from the vials' plastic walls during manipulation. Instead, specific silica-independent effects of the homeopathic medicine can be attributed to the decrease of interleukin-6 after treatment with the 200th centesimal dilution of arsenic.

Introduction

Nanoparticles (NPs) of starting materials and contaminants were recently detected in homeopathic high dilutions (HDs) above Avogadro's number.^{1,2} Since homeopathic medicines are traditionally prepared using glass vials, it was postulated that contaminant silica NPs detaching from vials during the step of agitation might represent a critical component involved in the biological effects of these medicines observed experimentally.¹⁻⁶ The understanding of their real effects (specific or non-specific) is crucial to characterize the mechanisms involved.

It is known that nanosilica can form stable three-dimensional structures using DNA, proteins or living cells as template (epitaxial growth).⁷⁻¹³ This phenomenon led to a microstructural hypothesis to account for the action of homeopathic medicines. Briefly, nanosilica would behave as non-specific amplifier or vehicle for the specific (structural or electromagnetic) information contained in starting materials, subsequently identified and processed by biological systems.¹⁴⁻¹⁶ Hypothetically, initial interaction and adsorption of starting material with silica NPs in low homeopathic dilutions would create "nanoseeds" that would be preserved in the preparation of the subsequent dilutions.^{6,17-19} Nanoparticles might also be associated with changes in electrical conductivity, as was shown for the homeopathic medicine *Ferrum metallicum*,²⁰ although other factors can also alter the conductivity of homeopathic aqueous solutions.²¹

Few studies have sought to assess the influence of glass (silica) on the effects of homeopathic HDs. In the pioneer study carried out by Boiron et al.,²² HDs prepared in paraffin-covered glass vials had smaller effect compared with the ones prepared in standard glass vials. One single study investigated trace elements in plastic and glass vials used to prepare and store homeopathic HDs.²³ The results showed absence of silica in the propylene vials and increase in the concentration of trace elements, mainly silica, lithium and sodium, when bi-distilled water was transferred from propylene to glass vials. The largest difference in trace element concentration occurred in the step from mother tincture to the first centesimal dilution (1cH); the later steps to 30cH were not associated with increase in trace element concentration. To the best of our knowledge, no study investigated differences in biological effects between HDs prepared in plastic and glass vials.

Therefore, the aim of the present study was to compare biological effects of homeopathic HDs—*Arsenicum album* (Ars) 6cH and 200cH—prepared in glass and plastic vials to investigate the role of silica microparticles in such effects. The presence of metal microparticles in the drug samples and other relevant physical properties of the solution (electrical conductance) were analyzed. The experimental design and choice of

the medicine were based on the literature²⁴ and the model chosen, macrophages in co-culture with microorganisms, was previously used in homeopathic fundamental research.²⁵ Macrophages are myeloid cells specialized for phagocytosis of micro-organisms, such as yeasts, and stand out for their ability to respond epigenetically to small environmental stimuli, mainly by production of cytokines/chemokines^{26,27} which makes these cells adequate biological sensors in HD research.

Materials and Methods

High Dilutions

Homeopathic matrix samples were produced according to the Good Manufacturing Practices at the Pharmacy Faculty of Federal University of Rio de Janeiro, following the Brazilian Homeopathic Pharmacopoeia guidelines.²⁸ The starting materials, arsenic and lactose monohydrate, were purchased from Sigma Aldrich (As₂O₃, batch number BCBD 1149V) and *Pharma Nostra Comercial Ltda* (Rio de Janeiro, Brazil), respectively.

Arsenic raw material containing 99.984% and 0.99984 mol/mol² of arsenic trioxide, as declared by the supplier, was acquired as a fine powder. The homeopathic manufacturing process followed the statements of Brazilian Homeopathic Pharmacopoeia²⁸ and started with the separation of lactose powder in three equal parts. One-third of the lactose was placed in a porcelain vessel to avoid As₂O₃ loss. Following this, As₂O₃ was added and triturated, meeting the centesimal scale (1 part in 99 parts of inert vehicle), and the complex (*Arsenicum album* plus lactose) was submitted to two independent and vigorous cycles of crushing and scraping for 20 minutes. After this first cycle, the second and third parts of lactose were added to the porcelain vessel, and submitted to the second and third cycles of crushing and scraping, for 20 minutes each, respectively. The trituration process was concluded at the end of 60 minutes. Each sample received the designation of first triturated 1/100, coded as *Arsenicum album* 1c. For the second triturated compounding, one part of the first one (1cH) was mixed in 99 parts of lactose (centesimal scale), and all crushing and scraping procedure repeated to obtain *Arsenicum album* 2cH. The last trituration process produced *Arsenicum album* 3cH, following the same protocol described above, using one part of *Arsenicum album* 2cH for 99 parts of lactose.

The *Arsenicum album* 3cH was used as matrix to the following dynamizations, always according to Brazilian Homeopathic Pharmacopoeia.²⁸ HDs were prepared using standard type II amber glass or PET (polyethylene terephthalate, R18/400, Apace, São Paulo, Brazil) vials. The plastic vials were sterilized by microwave heating, three 5-minute shocks at high intensity. The glass vials were sterilized in an

autoclave at 121 °C for 15 minutes. The PET material was silica free, according to the manufacturer.

High dilutions were prepared as follows: centesimal serial dilutions (6 for 6cH and 200 for 200cH) were made using different flasks and performing 100 strong vertical motions (agitation) for each dilution. All dilutions were prepared from *Ars 3cH* using 70% hydro-alcoholic solution (p/p)—which is equivalent to 77% (v/v)—in the following steps. A minor deviation of the pharmacopeia guidelines was included to prepare the final working homeopathic dilution, in which sterile water was used: this was necessary to treat single cell cultures without inducing biases related to alcohol concentration of the vehicle and to the possibility of culture contamination. Vertical agitation (“potentization”) was performed in each step by means of a mechanical arm used specifically for this purpose (Denise, AUTIC, São Paulo, Brazil). Theoretically, the concentration of *Ars 6cH* and *Ars 200cH* was 1.3×10^{-11} M and 1.3×10^{-366} M, respectively.

Ars 200cH prepared in glass vials was ultra-centrifuged at 5,000 rpm for 30 minutes to separate heavier particles, and the supernatant alone was analyzed. The tested HDs were designated *Ars 6cHP* (prepared in plastic vials), *Ars 6cHG* (prepared in glass vials), *Ars 200cHP* (prepared in plastic vials), *Ars 200cHG* (prepared in glass vials), and *Ars 200cHGC* (prepared in glass vials and centrifuged after). The control was untreated macrophage-yeast co-culture (Control + yeast).

The vials were wrapped with aluminum foil and labeled with a code by a laboratory technician who did not participate in the study (to make the procedures blinded). The investigators were blinded to the vials’ content until statistical analysis was completed.

Macrophage–Yeast Culture and Treatments

RAW 264.7 macrophages were thawed and sown into culture bottles containing RPMI-1640 medium (Cutilab, Brazil) supplemented with 20% fetal bovine serum and 200 U mL⁻¹ penicillin, 100 U mL⁻¹ streptomycin, and 2.6 µg mL⁻¹ amphotericin B (Cutilab, Campinas, Brazil). Plating was performed in triplicate on six-well plates with 25×10^5 cells per well. Glass slides were not used during plating to keep the cell environment silica-free. Yeast cells (*Saccharomyces cerevisiae*) were added onto macrophages adhered to the well bottom in proportion 1:10 (25×10^6 yeast cells per well).

The treatment solutions were prepared with pure water filtered in a 0.22 µm Millipore filter. The test solutions were added in proportion 20% to the well volume. A second application of treatments was performed 24 hours later, in proportion 1%. Forty-eight hours after the onset of treatments, the macrophages were fixed with absolute methanol for 15 minutes and subjected to Giemsa staining. This staining method is useful for clear visualization of the yeasts inside the macrophages.

Histomorphometry

Ten digital photographs were taken per field (NIKON E 200 microscope coupled to Coolpix digital camera, Tokyo, Japan) and 200 cells/well were counted through oil-immersion objective with 100× magnification. The mean area (pixels) of cells, representing cell spreading, and the percentage of phagocy-

tozed yeast cells were counted automatically using the software Metamorph (Molecular Devices; Saint Jose, California, United States). The phagocytic index was calculated as percentage of internalized yeasts to the number of macrophages in each field.

Acridine Orange Test

The co-cultured cells were incubated after 48 hours of treatment with acridine orange (5 µg/mL) under a 5% CO₂ hood for 20 minutes. Next, the wells were rinsed with phosphate buffered saline (PBS); samples were collected by touching the cells with a slide covered with one drop of PBS, and then examined under fluorescence microscope (Olympus-BX60, Tokyo, Japan) to investigate the lysosome activity. Images were taken with a digital camera coupled to the microscope (Dino-Eye AM7023, New Taipei, Taiwan). In this test, DNA-containing structures emit green fluorescence, and acid structures, such as lysosomes and phagolysosomes, emit red/orange fluorescence. Qualitative analysis of cell morphology was performed on serial photomicrographs.

Nitric Oxide Production

The supernatant from cells kept in co-culture for 48 hours was frozen at -80 °C for later measurement of nitrite and nitrate production. Measurement was performed in duplicate by means of the colorimetric method on 96-well plates: to each plate 100 µL of supernatant and 100 µL of Griess reagent (0.1% N-alpha-naphthyl-ethylenediamine (NED); 1% sulfanilamide; 2.5% of phosphoric acid; Sigma-Aldrich, São Paulo, Brazil) were added. A standard curve was plotted starting with the mother solution (140 mg of sodium nitrite [Sigma-Aldrich, São Paulo, Brazil] + 10 mL of distilled water). The plates were left at room temperature for 10 minutes and then read in enzyme-linked immunosorbent assay spectrophotometer (Thermoplate, TP reader basic, Beijing, China) at 540 nm. The results were assessed by comparing the optical density to the nitric oxide (NO) standard curve and expressed as µg of nitrite per 10⁵ cells.

Immunoenzymatic Assay for Cytokine Measurement

Supernatant of the yeast-infected macrophage cultures was collected 48 hours after the onset of treatments, centrifuged at 15,000 rpm for 5 minutes and frozen at -80 °C for later cytokine measurement. Cytokines were measured by means to the MAGPIX-Luminex method (kit MILLIPIX e-Bioscience mouse cytokine Magnet; Darmstadt, Germany) following the manufacturer’s instructions. This kit includes detection systems for interferon-γ, interleukin (IL)-1α, IL-1β, IL-6, IL-10, IL-12 (P240), IL-12 (P70), monocyte chemoattractant protein 1 (MCP-1), macrophage inflammatory protein-1β (MIP-1 beta), granulocyte macrophage colony-stimulating factor (GM-CSF), regulated on activation, normal T cell expressed and secreted chemokine (RANTES), vascular endothelial growth factor (VEGF), and tumor necrosis factor-α (TNF alpha). The samples were analyzed in triplicate and the results expressed as pg/mL. Samples with values below the limit of detection were recorded as 0 (zero).

Scanning Electron Microscopy/Energy-Dispersive X-Ray Spectroscopy

The identification of silicon content in suspended micro-particles probably contained in the tested solutions was performed by means of the micro-evaporation method, followed by scanning electron microscopy (SEM) under 10-kV working conditions and by energy-dispersive X-ray spectroscopy (EDX) (JSM 6510; JEOL Ltd, Tokyo, Japan).

Microtubes containing samples of each tested solution were centrifuged at 5,000 rpm for 30 minutes for the microparticles to sediment. A single drop of 10 μ L was harvested from the bottom of each tube and placed on a stub. The stubs were previously cleansed with 70% acetone and dried in a closed recipient. The stubs were then kept within a closed Petri dish to ensure that only particles obtained through centrifugation of the tested solutions were analyzed. The drops were left to dry naturally on the stubs up to the time of microscopic analysis. Images were taken with a 10- μ m scale (see example in **Fig. 1**). The samples were harvested only once from each tube, because in a pilot study we found that the second and third drops were almost particle free.

The results of EDX are expressed as the frequency of silicon among all analyzed microparticles. Thirty-four microparticles deposited on the stubs were randomly chosen for EDX analysis; for samples that did not reach this number, all the deposited particles were analyzed. As the stubs were made of copper, this element was blocked in the reader control to avoid interference. Considering the characteristics of the microscope, to warrant a minimum sensitivity threshold, for each identified element, the percentage of atoms per particle below 1.5% was considered as zero (negative result).

Thus, traces of other elements (C, O, Na, Mg, Zn, Ca, Be, Nb, Pb, Cl, Tc, Rb, Al, S, Fe, P) were randomly detected in all the analyzed dilutions, but, since they were present under the limit of sensitivity of the device, they were not considered.

Electrical Current Flow (Conductance)

A micro-ammeter (Ryodoraku; São Paulo, Brazil), calibrated to 200 M/A, was used to measure the capacity to allow electric current in each dilution. The device wires were connected to two parallel electrodes (30 \times 40 mm). The solutions were put into flat cryotubes with parallel walls to avoid contact between the electrodes and the tube wall.²⁹ For measurement, the electrodes were inserted 1 cm into the liquid. The test was performed in three series, in triplicate, at different times of the day, under a closed hood; electronic devices were kept at least 2 m away from the test site to avoid any influence of magnetic fields. A thermo-hygrometer was placed inside the hood to monitor temperature (24.8°C, on average) and humidity (57%). Conductivity was measured after a 5-second interval to allow for stabilization of the device. Pure sterilized water was used as standard for comparison, as its conductivity is quite stable and equal to 100 μ A. Sterilization included autoclaving for 20 minutes followed by filtering through a 22- μ Millipore filter. The vehicle (filtered and agitated 70% alcohol) was used as experimental control. A sample of water subjected to 100 vertical motions (potentization) was used as supplementary control.

Statistical Analysis

Statistical analysis was performed using IBM SPSS, version 21.0, and GraphPad Prism, version 6.0, for Windows.

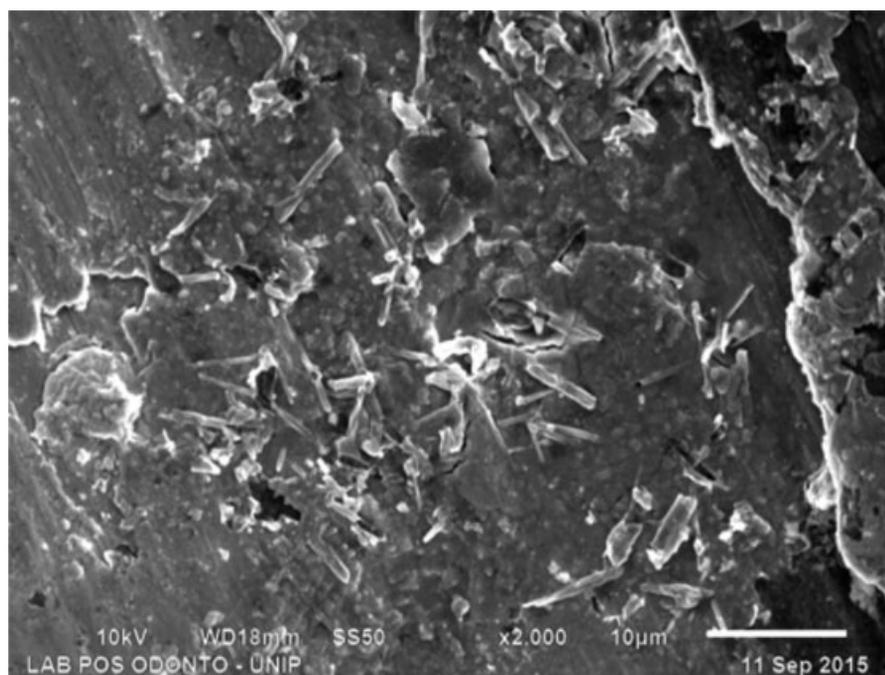


Fig. 1 Scanning electron microscopy (SEM) showing numerous micro-particles deposited on the stub surface. Note that the stub surface is irregular. SEM was used only to identify the presence of silica in the microparticles. Their morphology and size were not evaluated. An illustrative SEM profile showing particles on the stub from a sample of *Ars 6cHP* is represented. Magnification 2,000x.

Normality was evaluated by means of the Shapiro–Wilk test and assessed by inspection of quartile–quartile plots (Q–Q plots). Homogeneity of variances was evaluated by Levene's test and the Welch correction was applied to the ANOVA in cases of non-linearity. Outliers were assessed by inspection of Q–Q plots and were removed when necessary.

Outcomes were evaluated by means of two-way ANOVA and Dunnett's post-test. Partial eta-squared (η^2), a measure of effect size, was also reported. This parameter was particularly useful for variables presenting large deviation values and analyses with p -values that could lead to type I (rejection of a true null hypothesis) or II (retaining a false null hypothesis) errors.

Results are presented as mean \pm standard error and p -values < 0.05 were considered significant.

The profile of the atomic composition of microparticles detected in samples of arsenic dilutions was represented by dot plot graphics with mean and 95% confidence interval.

Results

Assessment of In Vitro Spreading, Phagocytosis and Nitric Oxide Production

On the orange acridine test, *Ars 200cHP* and *Ars 200cHG* induced greater lysosomal activity compared with the controls, manifested as patent fluorescent acid vacuoles (**Fig. 2**).

Macrophages treated with *Ars 200cHP* exhibited increased phagocytic index ($p = 0.001$, $F(2,81) = 5.638$, $\eta^2 = 0.173$) compared with the other groups. Similarly, NO production after 48-hour incubation was higher in *Ars 200cHP* ($p = 0.006$, $F(7,8) = 9.199$, $\eta^2 = 0.775$). However, *Ars 200cHG* and *Ars 200cHGC* presented significant reduction in spreading ($p = 0.001$, $F(3,60) = 11.893$, $\eta^2 = 0.373$) compared with the control and *Ars 200cHP* (**Figs. 3 and 4**).

Neither *Ars 6cHP* nor *Ars 6cHG* induced significant changes in macrophage spreading, phagocytic index or NO production after 48-hour incubation. On the acridine orange test, no significant differences were detected between *Ars 6cH* preparations and control (**Figs. 5 and 6**).

Cytokine Measurement

Interleukin-1 β and GM-CSF were not analyzed because the measured values were below the minimum limit of detection as declared by the manufacturer.

Ars 6cHP induced statistically significant increases in IL-1 α ($F(2,6) = 15.501$; $p = 0.04$; $\eta^2 = 0.838$), IL-6 ($F(2,6) = 10.894$; $p = 0.010$; $\eta^2 = 0.784$), MCP-1 ($F(2,6) = 14.165$; $p = 0.005$; $\eta^2 = 0.825$), and VEGF ($F(2,6) = 44.335$; $p = 0.01$; $\eta^2 = 0.947$) (**Fig. 7**). In turn, treatment with *Ars 200cHP*, *Ars 200cHG*, and *Ars 200cHGC* was associated with statistically significant reduction in IL-6 ($F(3,8) = 4.722$; $p = 0.035$; $\eta^2 = 0.639$) compared with the non-treated control (**Fig. 8**).

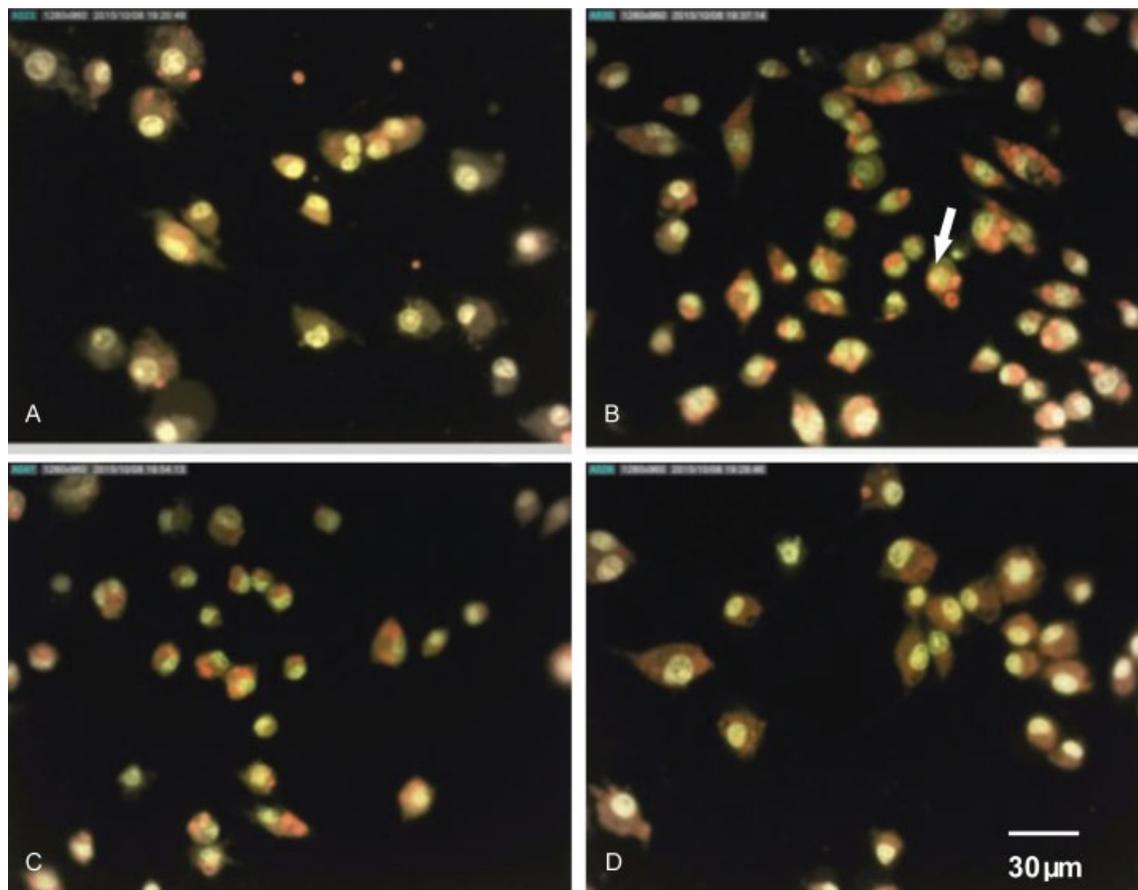


Fig. 2 Photomicrograph of cells stained by means of the acridine orange method under fluorescence microscope. (A) Macrophages co-cultured with yeast. (B) *Arsenicum album* 200cH prepared in plastic vials. (C) *Arsenicum album* 200cH prepared in glass vials. (D) Supernatant of centrifuged *Arsenicum album* 200cH prepared in glass vials. The arrow indicates a phagolysosome with acid content. Magnification 40x.

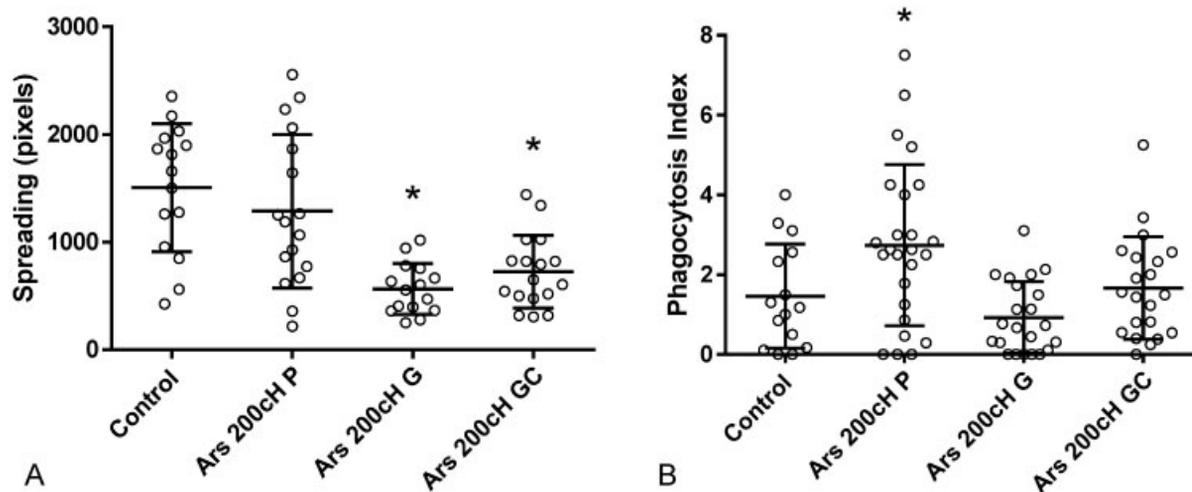


Fig. 3 Spreading (in pixels) (A) and phagocytic index (B) of RAW 264.7 macrophages treated with *Arsenicum album* 200cH. Values represent individual points, mean and standard error. Control + yeast: macrophage-yeast co-culture. *Ars* 200cHG: *Arsenicum album* 200cH prepared in glass vials. *Ars* 200cHGC: *Arsenicum album* 200cH prepared in glass vials and centrifuged. *Ars* 200cHP: *Arsenicum album* 200cH prepared in plastic vials. (A) ANOVA, $p < 0.001$, $F(3,60) = 11.893$, $\eta^2 = 0.373$, compared with *Ars* 200cHP and control; (B) ANOVA, $p < 0.001$, $F(2,81) = 5.638$, $\eta^2 = 0.173$, compared with the other treatments. ANOVA, analysis of variance.

No differences among groups were found regarding the other analyzed cytokines (► Table 1 and 2).

Scanning electron microscopy + energy-dispersive X-ray Particle Analysis

The presence of measurable silicon among identified particles was only found in *Ars* 6cHG and *Ars* 200cHG. No measurable silicon was detected in *Ars* 200cHP, *Ars* 200cHGC, and *Ars* 6cHP (► Table 3).

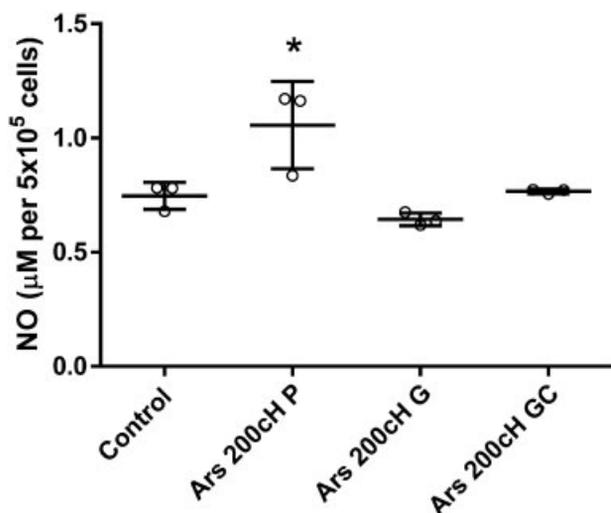


Fig. 4 Nitric oxide ($\mu\text{g}/10^5$ cells) in RAW 264.7 macrophage-yeast co-culture supernatant after 48-hour incubation. Values represent individual points, mean and standard error. Control + yeast: macrophage-yeast co-culture. *Ars* 200cHG: *Arsenicum album* 200cH prepared in glass vials. *Ars* 200cHGC: *Arsenicum album* 200cH prepared in glass vials and centrifuged. *Ars* 200cHP: *Arsenicum album* 200cH prepared in plastic vials. ANOVA, $p < 0.006$, $F(7,8) = 9.199$, $\eta^2 = 0.775$, compared with the other treatments. ANOVA, analysis of variance.

Electric Current Flow

The capacity for an electric current to flow in the fluid (conductance) of the pure filtered water was used as a standard for comparison. Significant reduction in electric flow in relation to water was found in 30% alcohol and *Ars* 200cHP, probably due to the presence of organic non-polar molecules. *Ars* 6cHP also presented low level of conductance, though without statistical significance due to the dispersion of the datapoints. The greatest conductance was exhibited by *Ars* 200cHGC and *Ars* 200cHG, in relation to the controls and *Ars* 200cHP ($p = 0.001$, $F(6,95) = 134.342$, $\eta^2 = 0.895$) (► Fig. 9).

Discussion

The results of the present study show plastic-related non-specific effects and very specific effects on IL production, which were independent of microparticles containing silicon but probably dependent on the smallest nanostructures, which could include water and the silica itself. In this sense, dilutions of arsenic prepared in plastic vials exhibited pro-inflammatory effects attributed to putative organic residuals detached from the vials' walls during disinfection and agitation, since these effects were not seen in glass-prepared medicines (► Supplementary Fig. 1, available in the online version only).

Macrophage spreading, phagocytic activity and increase in IL-1 α , IL-6, MCP-1, and VEGF in the culture medium were found only in samples treated with medicines prepared in plastic vials—*Ars* 6cHP and *Ars* 200cHP, but not in *Ars* 6cHG or *Ars* 200cHG. These findings suggest that plastic-derived soluble contaminants present in higher potencies might be somehow related to the increase in macrophage activity.

Available evidence indicates that PET bottles subjected to high temperature (40–60°C) do not exhibit cytotoxic

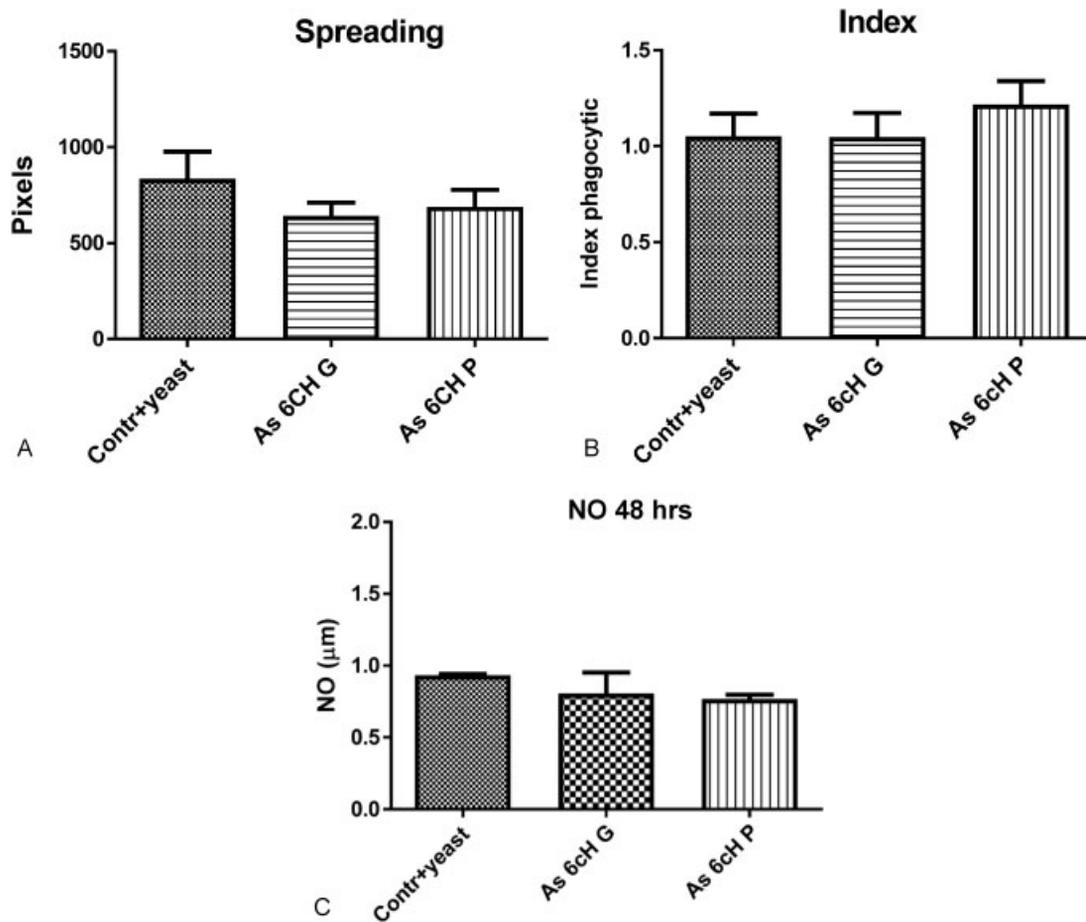


Fig. 5 Spreading (in pixels) (A), phagocytic index (B) and NO production (μg in 10^5 cells) (C) of RAW 264.7 macrophages treated with *Arsenicum album* 6CH. Values represent mean and standard error. Control + yeast: macrophage-yeast co-culture. *Ars 6CHG*: *Arsenicum album* 200cH prepared in glass vials. *Ars 6CHP*: *Arsenicum album* 6CH prepared in plastic vials. ANOVA, analysis of variance.

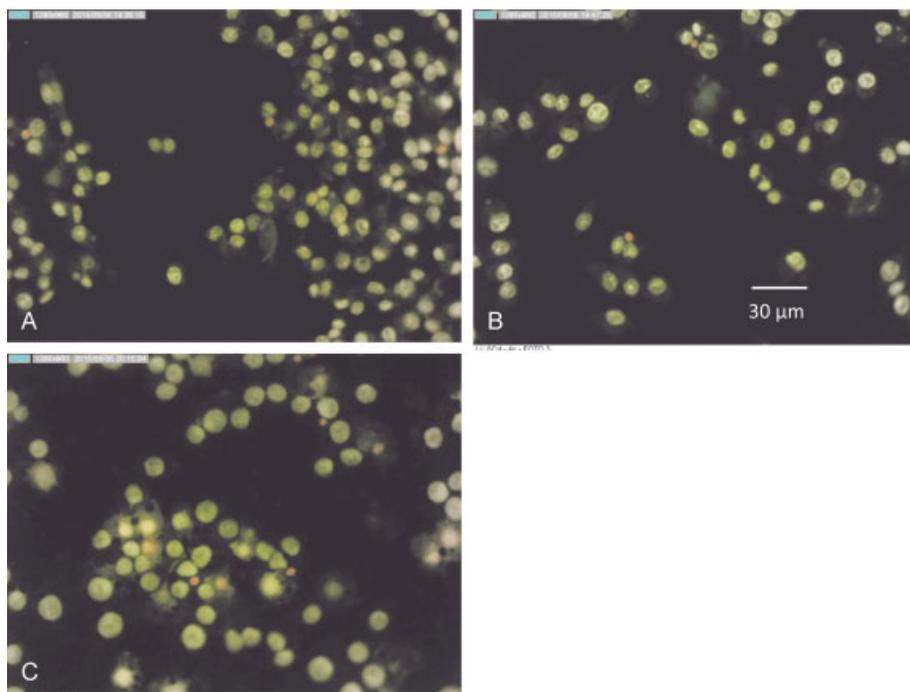


Fig. 6 Photomicrograph of cells stained by means of the acridine orange method under fluorescence microscope. (A) macrophages co-cultured with yeast. (B) *Arsenicum album* 6CH prepared in glass vials. (C) *Arsenicum album* 6CH prepared in plastic vials. 40x magnification.

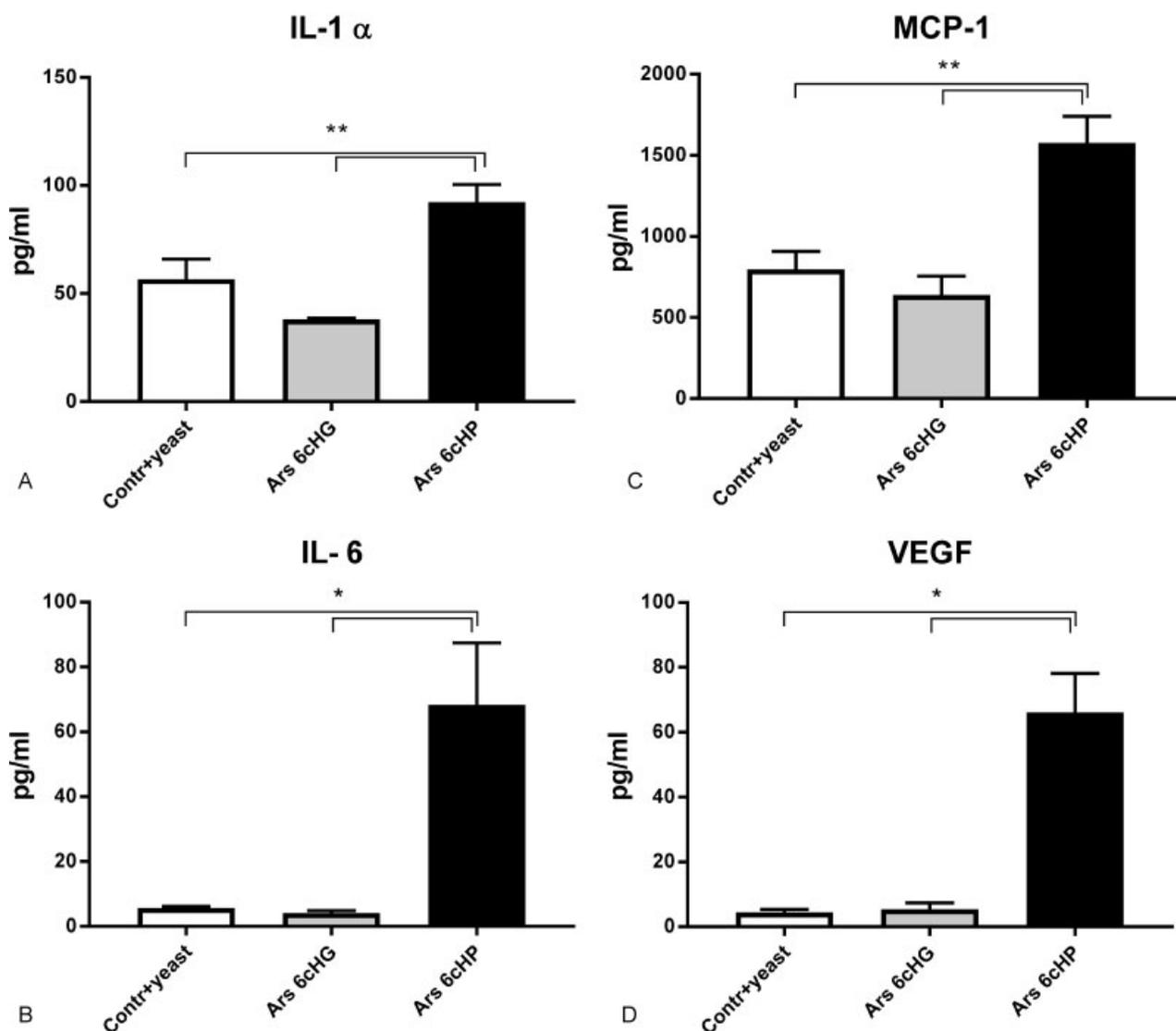


Fig. 7 Concentration (pg/ml) of cytokines and chemokines in the supernatant collected from RAW 264.7-yeast co-culture. IL-1 α (A), IL-6 (B), MCP-1 (C), VEGF (D) produced by cells treated with *Arsenicum album* 6cH prepared in glass and plastic vials. Values represent mean and standard error. * $p < 0.05$ compared with the untreated control, two-way ANOVA. ** $p < 0.005$ compared with the untreated control, two-way ANOVA. See text for η^2 values. ANOVA, analysis of variance; IL-1 α , interleukin-1 α ; MCP-1, monocyte chemoattractant protein-1; VEGF, vascular endothelial growth factor.

activity,³⁰ but biomaterials containing PET might modify the macrophage activity in vitro.^{31,32} In any case, plastic components have relevant biological actions, this fact being of interest for homeopathic pharmaceuticals and, thus, deserving of more thorough study. For instance, one study compared homeopathic medicine *Gelsemium sempervirens* 6c, 30c, and 200c to agitated vehicle controls (6c, 30c, and 200c) and non-agitated solvent prepared in vials with natural cork stoppers and with silicone stoppers. All the vials with cork stoppers exhibited more than 4×10^8 NPs/mL, significantly more than with the non-agitated silicone stoppers. On ultraviolet-visible spectroscopy, the agitated control 30c exhibited significantly higher absorbance, while the non-agitated silicone stopper controls had significantly lower mean absorbance than all the other samples.³³

These findings suggest that the agitation inherent to homeopathic remedy manufacture released trace materials

from natural cork stoppers that could interfere with various properties of HDs, and eventually also with their biological activity.³³ Also, potentized water exhibited a residual effect on macrophage spreading, which might be attributed to a non-specific action of micro- and/or nano-bubbles.³⁴ A direct action of medicines on yeast is unlikely in the present case because, according to the literature, treatment with *Ars* in decimal dilutions 8x and 16x did not induce any change in *S. cerevisiae* growth.³⁵

The decrease in IL-6 production might be considered as the main specific biological effect of *Arsenicum album* 200cH, since it was observed not only in this potency but in all situations: *Ars* 200cHG, *Ars* 200cHGC, and *Ars* 200cHP. Therefore, these specific effects were not related to the identification of silicon-containing microparticles and agree with the effects of *Ars* 200cH on leukocytes previously described.²⁴ The decrease in macrophage spreading after

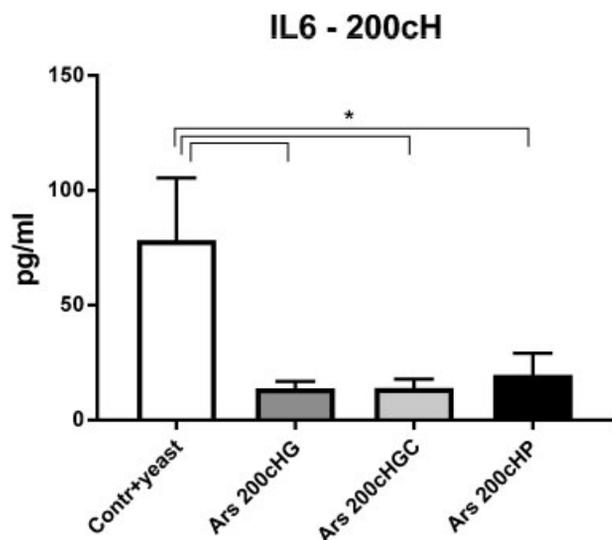


Fig. 8 Concentration (pg/mL) of IL-6 in the supernatant collected from RAW 264.7-yeast co-culture treated with *Arsenicum album* 200cH prepared in glass (centrifuged or not) and plastic vials. Values represent mean and standard error. **p* = 0.035 compared with the untreated control, two-way ANOVA. See text for η^2 values. ANOVA, analysis of variance; IL-6, interleukin-6.

treatment with *Ars 200cHG* and *Ars 200cHGC* corroborates this result; besides it is very specific, focused on the reduction in IL-6 production itself. This specific feature is also seen in other studies involving gene expression and homeopathy.^{27,36,37}

The acridine orange test showed an interesting result. While *Ars 200cHP* and *Ars 200cHG* induced a similar mor-

phological pattern, characterized by patent acid vacuoles (lysosomes), *Ars 200cHGC* (superficial phase after centrifugation) induced a less active pattern. Maybe the ultra-centrifugation of *Ars 200cHG* and the resulting changes in its microparticle composition can be related to this result, since the biggest particles present before centrifugation could also be phagocytosed, stimulating the cells.³⁸⁻⁴¹

The correlation between electric current flow through the liquids and the type of the vial used corroborates the hypothesis in which metal particles in suspension could improve conductivity^{1,2,21,41} and possible organic contaminants from plastic could impair it. The size of these particles seems not to be important in this case, since *Ars 200cHGC* (only the supernatant of the medicine, after ultra-centrifugation) presented the highest values of electric current flow.

In short, the results of the present study indicate lack of correlation between the presence of silicon-containing microparticles and biological (specific or non-specific) effects of the tested HDs. On these grounds, one might not infer that silicon-containing microparticles could carry the information of homeopathic medicines to biological systems. However, smaller particles, as NPs,^{1,2,4-6,22,32,33} might explain the divergence. Moreover, in this model, NPs of the original substance could not be observed, maybe due to the limitations of the microscope used. Some studies showing these particles and their putative hormetic effects are well described in the literature.⁴¹⁻⁴³

Another hypothesis remains to be checked in the future: that the capacity of water itself acts as information carrier, based on electromagnetic properties derived from its dipole behavior.⁴⁴⁻⁴⁸ Alternatively, the participation of both elements together—water and particles, mixed in complex

Table 1 Cytokines, chemokines and VEGF-A measurements (pg/mL) in the supernatant of macrophages treated with different protocols of *Arsenicum album*. The supernatants were harvested after 48 hours of culture and treatment. Values are expressed in mean and standard deviation. Experiment using *Arsenicum album* 6cH

Treatments	Untreated control	<i>Ars 6cHG</i>	<i>Ars 6cHP</i>
INF- γ	4.55 \pm 1.19	4.69 \pm 0.73	4.11 \pm 0.97
IL-1 α	56.64 \pm 16.03	38.04 \pm 1.15	92.34 \pm 13.95**
IL-6	5.68 \pm 0.95	4.16 \pm 1.25	68.25 \pm 33.20*
IL-10	3.61 \pm 1.25	3.27 \pm 0.39	3.98 \pm 0.66
IL-12 p40	6.01 \pm 2.30	4.63 \pm 0.74	4.98 \pm 0.95
IL-12 p70	10.86 \pm 9.53	4.53 \pm 1.22	5.42 \pm 2.18
MCP-1	795.33 \pm 193.08	638.66 \pm 202.74	1576.33 \pm 286.19**
MIP-1 β	12727.00 \pm 768.09	12647.33 \pm 494.54	13796.33 \pm 643.12
RANTES	261.33 \pm 58.52	262.00 \pm 75.43	352.66 \pm 104.57
VEGF	4.48 \pm 1.53	5.46 \pm 3.28	66.08 \pm 17.02*
TNF- α	890.66 \pm 139.95	760.00 \pm 113.84	714 \pm 65.50

Abbreviations: IL-1 α , interleukin-1 α ; INF- γ , interferon- γ ; MCP-1, monocyte chemoattractant protein-1; MIP-1 β , macrophage inflammatory protein-1 β ; RANTES, regulated on activation, normal T cell expressed and secreted chemokine; TNF- α , tumor necrosis factor- α ; VEGF, vascular endothelial growth factor.

**p* < 0.05 compared with the untreated control,
 ***p* = 0.005 compared with the untreated control, two-way ANOVA.
 See text for η^2 values.

Table 2 Cytokines, chemokines and VEGF-A measurements (pg/mL) in the supernatant of macrophages treated with different protocols of *Arsenicum album*. The supernatants were harvested after 48 hours of culture and treatment. Values are expressed in mean and standard deviation. Experiment using *Arsenicum album* 200cH

Treatments	Untreated control	Ars 200cHG	Ars 200cHGC	Ars 200cHP
INF- γ	4.75 \pm 1.36	3.96 \pm 1.09	5.08 \pm 1.47	4.75 \pm 1.50
IL-1 α	77.70 \pm 71.27	82.06 \pm 70.67	48.02 \pm 42.20	71.71 \pm 39.02
IL-6	78.48 \pm 46.63	13.78 \pm 5.22*	13.96 \pm 35.77 *	19.56 \pm 31.49*
IL-10	4.00 \pm 0.94	4.36 \pm 1.55	3.61 \pm 1.27	3.81 \pm 1.22
IL-12 p40	4.85 \pm 1.42	4.89 \pm 1.80	4.28 \pm 1.46	4.22 \pm 1.42
IL-12 p70	7.86 \pm 3.63	6.06 \pm 4.21	6.39 \pm 2.55	6.84 \pm 2.37
MCP-1	3237.00 \pm 1574.47	1453.00 \pm 940.16	1985.33 \pm 1111.32	1742.66 \pm 1053.59
MIP-1 β	14675.33 \pm 1302.98	13161.67 \pm 1685.76	14763.33 \pm 4743.15	13758.33 \pm 4577.11
RANTES	230.00 \pm 118.82	121.97 \pm 84.36	114.12 \pm 80.16	157.50 \pm 82.97
VEGF	8.39 \pm 6.61	8.31 \pm 3.90	8.19 \pm 3.30	5.88 \pm 3.13
TNF- α	1765.00 \pm 474.00	1345.66 \pm 509.48	1448.33 \pm 502.74	1536.33 \pm 557.24

Abbreviations: ANOVA, analysis of variance; IL-1 α , interleukin-1 α ; INF- γ , interferon- γ ; MCP-1, monocyte chemoattractant protein-1; MIP-1 β , macrophage inflammatory protein-1 β ; RANTES, regulated on activation, normal T cell expressed and secreted chemokine; TNF- α , tumor necrosis factor- α ; VEGF, vascular endothelial growth factor.

* p = 0.035 compared with the untreated control, two-way ANOVA.

η^2 = 0.639.

nano-structures that are almost impossible to separate in nature—could elicit some biological effect.⁴⁹

Many studies using refined methods are still needed to establish which hypotheses account for the mechanism of action of homeopathy, since several rationales represent reasonable possibilities.^{50,51}

Conclusions

Homeopathic *Arsenicum album* in both potencies, 6cH and 200cH, can induce changes in conductivity and biological effects on macrophages according to the type of vial used (plastic or glass). Non-specific plastic-related pro-inflammatory effects were seen in both potencies of *Arsenicum album* prepared in plastic vials. Specific anti-inflammatory effect was seen for *Arsenicum album* 200cH, by its reduction in IL-6 whatever the kind of vial used and the presence or absence of silicon-containing microparticles. However, the participation of smaller particles (NPs) and/or other physico-chemical properties of water remains to be determined.

Table 3 Frequency of the element silicon among the identified micro-particles found in different *Arsenicum album* samples, using SEM/EDX system (number of particles “positive” to the presence of silicon in relation to the total)

Frequency	Ars 6cHG	Ars 6cHP	Ars 200cHG	Ars 200cHGC	Ars 200cHP
Per particle	3/34	0/24	3/34	0/13	0/24
Percentage	8.8%	0%	12.5%	0%	0%

Abbreviations: EDX, energy-dispersive X-ray spectroscopy; SEM, scanning electron microscopy.

Highlights

- *Arsenicum album* 200 cH decreases specifically IL-6 expression in macrophages in vitro.
- Silicon content in microparticles found in *Arsenicum album* has no relation to specific biological effects.
- Remedies prepared in plastic vials presented non-specific pro-inflammatory action.

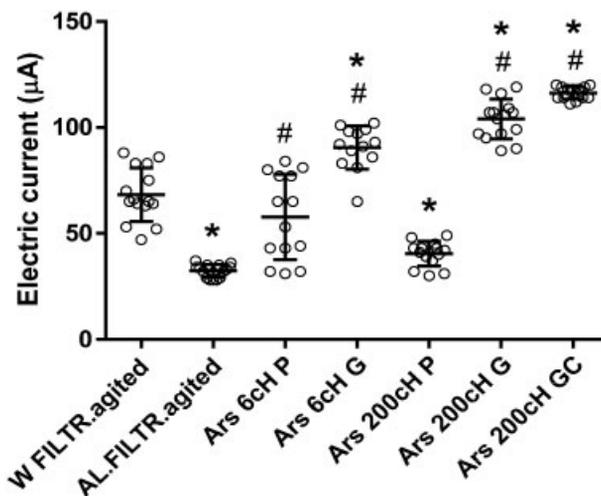


Fig. 9 Electrical current flow (conductance) of arsenic HDs (μ A). Values represent individual points, mean and standard error. W FILTR agitated: pure water filtered in a 22- μ m filter (Millipore) and vertically agitated (100 times). AL FILTR agitated: 70% alcohol filtered in 22- μ m filter (Millipore) and vertically agitated (100 times). *Ars 6cHG*: *Arsenicum album* 6cH prepared in glass vials. *Ars 6cHP*: *Arsenicum album* 6cH prepared in plastic vials. *Ars 200cHG*: *Arsenicum album* 200cH prepared in glass vials. *Ars 200cHGC*: *Arsenicum album* 200cH prepared in glass vials and centrifuged. *Ars 200cHP*: *Arsenicum album* 200cH prepared in plastic vials. ANOVA, $F(6, 95) = 134.342$, $\eta^2 = 0.895$; * $p < 0.001$ in relation to water; ** $p < 0.001$ in relation to the other treatments.

- Decrease in IL-6 was independent of the material of the vials and/or of the silica microparticle content.
- The nature of the vials seems to influence the electric flow in the respective fluids.

Supplementary Files

► **Supplementary Fig. 1** Flow-chart of the study design and the main results.

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Conflict of Interest

The authors declare there is no conflict of interest. Funding sponsors did not interfere with the design of the study, collection, analysis or interpretation of data, manuscript writing, or with the decision to publish the results. The initial project was selected for ABFH grant after ad hoc and blind evaluation of the proposal.

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