Maternal Age at Delivery and Enzyme Polymorphisms in Children with Type 1 Diabetes Mellitus

Fulvia Gloria-Bottini1 Anna Neri1 Patrizia Saccucci1 Maria Luisa Manca Bitti2 Novella Rapini2
Gabriele Renzetti1 Andrea Magrini1 Egidio Bottini1

1 Department of Biomedicine and Prevention, University of Rome Tor Vergata, Rome, Italy
2 Department of Medical Science, Pediatric Diabetes Unit, University of Rome Tor Vergata, Rome, Italy

J Child Sci 2018;8:e7–e10.

Abstract

Fetal genetic adaptation to environment of aging women could result in positive selection of genes that during extrauterine life increases the risk of type 1 diabetes mellitus (T1DM). We have examined the distribution of three genetic polymorphisms (acid phosphatase locus 1 [ACP1], p53 codon 72, and PTPN22) involved in T1DM risk in relation to maternal age at delivery. ACP1 was determined in 281 T1DM children, ACP1 in 207 children, and PTPN22 in 216 children. Controls (blood donors) were 351 for ACP1, 271 for PTPN22, and 730 for p53 codon 72. Genotypes were determined by DNA analysis. The proportions of the three genotypes associated with T1DM are much greater in T1DM children from older mothers than in those from young mothers and in controls. The data support the hypothesis that advanced maternal age favors a positive selection of genes more adapted to the uterine environment of older women: these genes predispose to T1DM during extrauterine life.

Introduction

The frequency of type 1 diabetes mellitus (T1DM) increases with maternal age at conception.1–5 High pregnancy estrogen concentration in older women may have an important role; however, the mechanism of the association between T1DM and maternal age is unknown.6

Experimental data and clinical observations suggest that delaying childbearing influences the biology of the mother-fetus relationship.7,8 Advanced maternal age could influence intrauterine selection favoring genotype more adapted to the environment of less young women. Indeed, we have found that advanced maternal age is associated with changes in the frequency of haptoglobin phenotypes in the mother and with changes of PGM1/RhC haplotype distribution in both mothers and offspring.9

Thus, we reasoned that fetal genetic adaptation to the environment of aging women could result in a positive selection of genes that during extrauterine life may increase the risk of immunological diseases such as T1DM.

In the present note, we have examined the distribution of three genetic polymorphisms involved in T1DM risk (acid Phosphatase Locus 1 [ACP1], p53 codon 72, and PTPN22)10–12 in children with T1DM in relation to maternal age at delivery.

Acid Phosphatase Locus 1

Cytosolic low molecular weight protein tyrosine phosphatase (cLMWPTP) is encoded by ACP1 gene that shows three codominant alleles: ACP1^A, ACP1^B, and ACP1^C and correspondingly six genotypes with enzymatic activity increasing in the order ACP1^A < ACP1^B < (ACP1^B/A < ACP1^C/A) < (ACP1^B/C < ACP1^C/B) < (ACP1^C/A < ACP1^C/B).13,14

cLMWPTP is able to dephosphorylate a negative phosphorylation site in the ZAP70 tyrosine kinase in T cell,15 and as consequence of the increase of ACP1 activity, there is an increase in ZAP70 activity and signaling from T cell antigen receptor.

p53 Codon 72

p53 codon 72 shows a polymorphism due to a single nucleotide substitution that changes arginine to proline in the

Received
April 28, 2017
Accepted after revision
January 11, 2018
ISSN 2474-5871.

Copyright © 2018 Georg Thieme Verlag KG Stuttgart · New York
License terms

Keywords
► ACP1
► maternal age
► p53 codon 72
► PTPN22
► type 1 diabetes mellitus
protein. There are three genotypes \textsuperscript{‘}Arg/\textsuperscript{‘}Arg, \textsuperscript{‘}Arg/\textsuperscript{‘}Pro, and \textsuperscript{‘}Pro/\textsuperscript{‘}Pro. The arginine variant induces a strong apoptosis activity, while Proline variant induces a strong transcriptional activity.\textsuperscript{16}

p53 is involved in autoimmune diabetes through down-regulation of STAT1.\textsuperscript{17}

\textbf{PTPN22}

Human lymphoid tyrosine phosphatase (Lyp) is encoded by PTPN22 gene and is a regulator of signaling through T cell receptor. The polymorphism is due to a substitution of an arginine with a tryptophan at codon 620. The substitution generates the Lyp-W620 variant that is associated with autoimmune disorders and gain of function of the enzyme.\textsuperscript{12–18}

The polymorphism has two alleles \textsuperscript{‘}C (R-620 variant) and \textsuperscript{‘}T (W-620 variant) and three genotypes \textsuperscript{‘}C/\textsuperscript{‘}C, \textsuperscript{‘}C/\textsuperscript{‘}T, and \textsuperscript{‘}T/\textsuperscript{‘}T.

\textbf{Materials and Methods}

We have studied 281 children with T1DM from the white population of Rome: p53 codon 72 was determined in all subjects, ACP\textsubscript{1}, in 207, and PTPN22 in 216 subjects. Controls (blood donors) from the same population were 351 for ACP\textsubscript{1}, 271 for PTPN22, and 730 for p53 codon 72. Maternal age at delivery had been registered in clinical records in 90 mothers. These subjects have also been considered in previous studies.\textsuperscript{9,11}

Genotype of ACP\textsubscript{1}, PTPN22, and p53 codon 72 was determined by DNA analysis as previously described.\textsuperscript{19}

\textbf{ACP\textsubscript{1}}

Total genomic DNA was extracted from frozen whole blood samples collected in ethylenediaminetetraacetic acid. The amplification conditions consisted of an initial denaturation at 94°C for 2 minutes, followed by 35 cycles of 94°C for 45 seconds, 54°C for 45 seconds, 72°C for 45 seconds, and a final extension of 72°C for 5 minutes.

Ten microliters of the 341-bp exon 3 amplicon was fully cleaved by Cfol at 37°C for 1 hour according to the manufacturer’s instructions and then electrophoresed on 1.8% agarose gels. The digestion created two fragments of 255- and 86-bp for ACP\textsubscript{1}; A and ACP\textsubscript{1}; B alleles, while the ACP\textsubscript{1}; C allele was not out. Similarly, the 299-bp polymerase chain reaction (PCR) product was digested by Taq1 at 65°C for 1 hour according to the manufacturer’s instructions, generating two fragments of 100 and 199 bp for the ACP\textsubscript{1}; A allele but not for the B and C alleles.

\textbf{PTPN22}

A DNA fragment was amplified by PCR in a 25 µL total-volume reaction, containing 100 ng of genomic DNA, 2.5 nM of MgCl\textsubscript{2}, 1× buffer Gold (Applied Biosystems, Foster City, California, United States), 10 pmol of each primer, 0.2 mM of deoxyribonucleotide triphosphate, and 0.5U of AmpliTag Gold (Applied Biosystems). Thirty cycles (30 s at 95°C, 30 s at 54°C, and 30 s at 72°C) were performed with the DNA thermal cycler (Perkin Elmer).

The C/T transition at codon 620 (NCBI refSNP ID: rs2476601) creates a XcmI restriction site in the T allele. The polymorphism was identified by XcmI restriction endonuclease (NEB, Beverly, Massachusetts, United States) digestion of the polymerase chain reaction amplified fragment. After electrophoresis, the gel was stained with ethidium bromide, and the fragments were visualized under ultraviolet light.

\textbf{p53 Codon 72}

Polymerase chain reactions were performed in a total volume of 25 µL containing 200 ng of genomic DNA, 0.4 pmol of each primer, 2 nmol/L of MgCl\textsubscript{2}, 200 mmol/L of deoxynucleotide triphosphates, 1× buffer, and 2U of Taq polymerase. The amplification was performed for 35 cycles with an annealing temperature of 62°C. The amplified DNA was digested for 3 hours with 3U of AccI restriction enzyme. The DNA fragments were resolved by electrophoresis on a 3% agarose gel.

Informed consent was obtained by the mothers to participate in the research project that was approved by the Council of Department.

Chi-square test of independence and principal component analysis were performed by commercial software (SPSS). The median value of mothers’ age was 32 years.

Controls were compared with the whole sample of T1D children and with T1D children from mothers aging 32 years or more at delivery.

\textbf{Results}

\textbf{Table 1} shows the distribution of ACP\textsubscript{1}, p53 codon 72, and PTPN22 genotypes in relation to maternal age at delivery of children with T1DM. As previously described, the proportions of ACP\textsubscript{1}; A/B, \textsuperscript{‘}Arg/\textsuperscript{‘}Arg, p53 codon 72, and carriers of \textsuperscript{‘}T allele of PTPN22 are greater in subjects with T1DM than in controls. The proportion of these genotypes, however, is much greater in children with T1DM from mothers aging more than 32 years (median value of mothers’ age) at delivery. These results suggest that in the environment of aging mothers, zygotes carrying these genotypes could be advantaged with respect to zygotes carrying others genotypes.

\textbf{Figs. 1 to 3} show the correlation between maternal age and the proportion of \textsuperscript{‘}Arg/\textsuperscript{‘}Arg genotype, \textsuperscript{‘}A/B genotype, and carriers of W620 variant of PTPN22 in children with T1DM. A principal component analysis has shown a clear correlation of maternal age with the first two variables and a moderate correlation of maternal age with PTPN22 genotype of children (data not shown).

\textbf{Figs. 4} shows the correlation between maternal age and the mean percentage increase in the frequencies of \textsuperscript{‘}A/B genotype, \textsuperscript{‘}Arg/\textsuperscript{‘}Arg genotype, and carriers of \textsuperscript{‘}T allele in children with T1DM as compared with the frequencies in the general population. With the increase in maternal age, there is a progressive increase in these genotypes compared with the frequencies observed in the general population.

\textbf{Table 2} shows the distribution of ACP\textsubscript{1}, \textsuperscript{‘}A/B genotype in healthy newborns in relation to maternal age. In the
offspring of mothers aging at delivery more than 32 years, there is a statistically significant increase in this genotype, suggesting that the increase in *C3* A/*C3* B genotype observed in T1DM offspring of older mothers is a general phenomenon connected with maternal aging. Unfortunately, we have no similar samples of newborns to study this phenomenon for *C3* Arg/*C3* Arg of p53 codon 72 and for carriers for *C3* allele of PTPN22.

### Table 1 Distribution of genetic polymorphisms associated with T1DM in relation to maternal age at conception

<table>
<thead>
<tr>
<th>Genetic polymorphisms</th>
<th>Genotype</th>
<th>Controls (A)</th>
<th>T1DM (all subjects) (B)</th>
<th>T1DM (mothers &gt;32 y at conception) (C)</th>
<th>Comparisons</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td></td>
<td>%</td>
<td>N</td>
<td>%</td>
<td>N</td>
</tr>
<tr>
<td>ACP1</td>
<td><em>A</em>/B</td>
<td>34.5%</td>
<td>351</td>
<td>43.5%</td>
<td>207</td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td>A vs. B: p = 0.035</td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td>A vs. C: p = 0.020</td>
</tr>
<tr>
<td>p53 codon 72</td>
<td><em>Arg</em>/Arg</td>
<td>48.8%</td>
<td>730</td>
<td>58.0%</td>
<td>281</td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td>A vs. B: p = 0.009</td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td>A vs. C: p = 0.006</td>
</tr>
<tr>
<td>PTPN22</td>
<td><em>T</em> allele carriers</td>
<td>5.9%</td>
<td>271</td>
<td>11.6%</td>
<td>216</td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td>A vs. B: p = 0.030</td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td>A vs. C: p = 0.027</td>
</tr>
</tbody>
</table>

Abbreviations: ACP1, acid phosphatase locus 1; T1DM, type 1 diabetes mellitus.

Fig. 1 The correlation between maternal age at birth and the proportion of offspring with T1D carrying the *Arg*/Arg genotype. T1D, type 1 diabetes mellitus.

Fig. 2 The correlation between maternal age at delivery and the proportion of offspring with T1D carrying the A/B genotype. T1D, type 1 diabetes mellitus.

Fig. 3 The correlation between maternal age at delivery and the proportion of offspring with T1D carrying the W620 variant of PTPN22. T1D, type 1 diabetes mellitus.

Fig. 4 Correlation between maternal age and percentage increase in the frequencies of *A*/B genotype, *Arg*/Arg genotype, and carrier of W620 variant in T1D children with respect to the frequencies in the general population. T1D, type 1 diabetes mellitus.
Table 2 Distribution of ACP1* A/B genotype in healthy newborn in relation to maternal age

<table>
<thead>
<tr>
<th>Maternal age</th>
<th>Maternal age</th>
<th>Significance of difference</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>≤ 32 y</td>
<td>&gt; 32 y</td>
</tr>
<tr>
<td>% proportion of <em>A</em>/B genotype</td>
<td>%</td>
<td>N</td>
</tr>
<tr>
<td>31.3%</td>
<td>36.0%</td>
<td>262</td>
</tr>
</tbody>
</table>

Abbreviation: ACP1, acid phosphatase locus 1.

**Discussion**

We have considered three genetic systems involved in immunological functions and associated with T1DM. The analysis of genotype distributions of these systems has shown that in children from mothers aging more than 32 years at delivery, there is a strong increase in genotypes associated positively with T1DM. For ACP1, this increase has been observed also in healthy consecutive newborns delivered by mothers aging more than 32 years.

These data are in favor of the hypothesis that advanced maternal age influences intrauterine selection in favor of genes more adapted to the environment of older women. These genes could favor survival during intrauterine life but predispose to T1DM during extrauterine life.

Several mechanisms have been proposed to explain the association between maternal age and risk to T1DM in the offspring. As far as we know, however, our observation suggests a new mechanism connecting maternal age and susceptibility to T1DM: modification of intrauterine environment due to advancing maternal age favors the survival of fetus carrying genes that predispose to T1DM during extrauterine life.

Further studies on this problem would be rewarding.

**Conflict of Interest**

None.

**References**