GlucoSiFA and LactoSiFA: New Types of Carbohydrate-Tagged Silicon-Based Fluoride Acceptors for $^{18}$F-Positron Emission Tomography (PET)

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Abstract GlucoSiFA derivatives bearing an azide or alkynyl side chain were obtained from peracetyl-D-glucose using as key step a tosylate substitution by a SiFA thiolate obtained from 4-(di-tert-butylfluorosilyl)benzenethiol. In analogy, two-fold SiFA-substituted maltose and lactose derivatives were synthesized via bistosylates. Introduction of an acetal-protecting group in $\beta$-D-azidolactose allowed the synthesis of a LactoSiFA derivative bearing only one SiFA moiety.

Key words carbohydrates, silicon-based fluoride acceptors, nucleophilic substitution, tosylation, regioselectivity

The introduction of positron emission tomography (PET) as a non-invasive method for medical diagnostic in vivo imaging has become an indispensable tool in precision medicine development.$^{1}$ PET not only helps to understand the complex interplay between biological targets such as receptors and enzymes and their cognate ligands but furthermore assists devising new therapeutic regimens based on non-invasive biological target validation. Besides PET, a straightforward example for diagnostic imaging is the use of X-rays that has revolutionized medicine. However, this method only yields anatomic/structural information whereas PET and related radioisotope-based imaging methodologies look directly at dynamic biological processes without interference. PET, in addition to magnetic resonance imaging (MRI) and computed tomography (CT), has proved to be an elegant and non-invasive method to elucidate in vivo biochemistry. It allows metabolic tracking of bioactive compounds and quantification of biochemical and/or enzymatic processes in living organisms. Among commonly used radioactive isotopes such as $^{13}$C, $^{15}$O, $^{18}$F, $^{64}$Cu, and $^{68}$Ga,$^{2}$ the use of fluorine-18 has become rather popular due to its favorable physical properties such as a half-life of 109.7 minutes that allows for longer synthesis times and remote shipment to local imaging facilities and a low positron energy leading to PET images of highest resolution.

There are different strategies for incorporating $^{18}$F into radiopharmaceuticals. On the one hand, fluorination at carbon atoms in both aromatic and aliphatic compounds can be achieved by electrophilic as well as nucleophilic reactions and a variety of appropriate reagents has been developed for this purpose.$^{2,3}$ Alternatively, $^{18}$F can also be bound via isotopic exchange to non-carbon elements such as boron, aluminum, silicon, and phosphorus.$^{3a}$ These non-canonical labeling concepts got momentum in the last two decades although some of the labeling principles have already been introduced to the literature as early as 1958$^{4}$ but remained dormant for many years. The progress achieved in these types of chemistry was regularly summarized in review articles.$^{3a,5}$ Aryldialkylsilicon fluorides, ArR$_2$SiF, in which the silicon atom is sufficiently protected with R = isopropyl$^{6}$ or tert-butyl$^{7a}$ substituents are stable under physiological conditions and undergo $^{19}$F to $^{18}$F isotopic exchange with good radiochemical yields. The tert-butyl-
substituted variant is now widely known as SiFA methodology and has become popular in several research groups (Equation 1).7b–e

\[
\begin{align*}
\text{X} & \quad \text{Si}^{19F} \quad \text{Si}^{18F} \\
\xrightarrow{18F^{-}/\text{Kryptofix} 222/K^+} \\
\text{X} & \quad \text{Si}^{18F} \quad \text{Si}^{19F}
\end{align*}
\]

Equation 1 Isotope exchange in silicon-based fluoride acceptors (SiFA)

Biographical Sketches

Anja Wiegand studied chemistry at the Faculty of Chemistry and Chemical Biology of TU Dortmund. She obtained her B.Sc. in 2012 and her M.Sc. in 2014. In 2018, she finished her Ph.D. work on carbohydrate-based NHC-gold complexes and SiFA compounds under the supervision of Prof. Dr. Norbert Krause.

Vera Wiese studied chemical biology at the Faculty of Chemistry and Chemical Biology of TU Dortmund. She obtained her B.Sc. in 2015 and her M.Sc. in 2018. For her Master’s thesis, she worked on the synthesis of discharraride-based SiFA compounds under the supervision of Prof. Dr. Norbert Krause.

Britta Glowacki studied chemistry at the Faculty of Chemistry and Chemical Biology of TU Dortmund. She obtained her B.Sc. in 2011 and her M.Sc. in 2013. In 2018, she finished her Ph.D. work on amino alcoholate derivatives of group XIV elements under the supervision of Prof. Dr. Klaus Jurkschat.

Ljuba Iovkova was born in Sofia, Bulgaria and moved to Germany to study pharmacy and chemistry. In 2006, she obtained her Diploma from the TU Dortmund. She graduated from the Technische Universität Braunschweig in 1984 and received her Ph.D. in 1986.

Ralf Schirrmacher obtained his Ph.D. in nuclear chemistry from the University of Mainz in 1999. After a brief postdoctoral stay at the University of Pennsylvania, he continued research at the University of Mainz as a civil servant until 2007. In 2008 he was appointed Head of Radiochemistry and Director of Cyclotron at the McConnell Brain Imaging Center at the Montreal Neurological Institute (MNI) of McGill University. During his time at McGill University he held a Canada Research Chair in Molecular Imaging and Radiochemistry. He is currently a Full Professor in Oncologic Imaging at the Faculty of Medicine at the MICF and Cross Cancer Center at the University of Alberta at Edmonton. His research group develops new imaging agents for Positron Emission Tomography (PET) imaging in the field of oncology and neurology using a variety of different radionuclides such as carbon-11, fluorine-18, gallium-68, different radioisotopes of iodine and copper-64.

Klaus Jurkschat received his Ph.D. from Martin Luther University Halle-Wittenberg, Germany, in 1980 and habilitated in 1987 at the same university. After postdoctoral work with Jean-Bernard Robert at CENG Grenoble, France, and Marcel Gielen/Rudolph Willem at VUB Brussels, Belgium, and staying as visiting researcher at SUNY Albany (with Henry G. Kuivila), N.Y., USA, and Deakin University Geelong (with Dainis Dakternieks), Vic. Australia, he went to Dortmund, Germany, where from 1994 to 2018 he was employed as a Full Professor for Inorganic Chemistry at the Technische Universität. He was a Guest Professor at Universite Bordeaux 1 (2000) and Universite Rennes 1 (2012). He is Editor-in-Chief of the journal ‘Main Group Metal Chemistry’ and author of approximately 320 publications. The chemistry of hypercoordinate main group element compounds in all its facets is in the center of his research interests.

Norbert Krause graduated from TU Braunschweig in 1984 and received his Ph.D. in 1986. After postdoctoral stays at the ETH Zürich and Yale University, he joined TU Darmstadt and obtained his Habilitation in 1993. In 1994, he moved to the University of Bonn as Associate Professor, before being appointed to his present position as Full Professor at TU Dortmund in 1998. He was a Fellow of the Japan Society for the Promotion of Science (JSPS) in 2003, 2009, and 2015, and Guest Professor at the Université Catholique de Louvain (2007), at the University of California, Santa Barbara, U.S.A. (2009), and at the École Supérieure de Physique et de Chimie Industrielles de la Ville de Paris (ESPCI), France (2009). He was a member of the Editorial Board of the European Journal of Organic Chemistry (2006–2013). His research focuses on sustainable coinage metal (copper, silver, and gold) catalysis, in particular with water as bulk solvent.
derivatives that also contain a variety of functional groups that hold potential for subsequent protein conjugation by click-type chemistry.

Our synthesis of the β-D-azido-substituted GlucoSiFA derivative 5 (Scheme 1) started with peracetyl-D-glucose (1; mixture of anomers), which was first brominated at the anomeric center with HBr/AcOH. The resulting α-D-glycosyl bromide underwent clean substitution with sodium azide to afford β-D-glycosyl azide 2. Both steps proceeded in excellent yield, as did the subsequent deprotection with sodium methoxide. A selective monotosylation at the primary hydroxy group of the unprotected β-D-azidogalactose gave the difunctionalized monosaccharide 3 in 72% yield. Finally, the desired GlucoSiFA derivative 5 was obtained in 74% yield (51% over 5 steps) by nucleophilic substitution of the tosylate with deprotonated 4-(di-tert-butylfluorosilyl)benzenethiol (4). Due to the pronounced base sensitivity of SiFA derivatives, this step has to be carried out with bulky, less nucleophilic bases. In the event, potassium tert-butoxide in DMSO gave the best results.

Starting point of the synthesis of the β-D-alkynyl-substituted GlucoSiFA derivative 8 (Scheme 2) was the Lewis acid catalyzed substitution of the anomeric acetyl group of peracetyl-D-glucose (1) with propargyl alcohol, which provided β-D-glycoside 6 in 76% yield. The following steps via monotosylate 7 proceeded as before and gave target molecule 8 in 33% yield over four steps.

In order to further increase the hydrophilicity of carbohydrate-tagged SiFA derivatives, we next used disaccharides as starting material. Here, serious reactivity and selectivity issues were encountered. For example, even though β-D-azidogalactose can be monotosylated selectively at the primary hydroxy group, all attempts of a monotosylation of β-D-azidolactose (which also contains only one primary hydroxy group) failed and provided either mixtures of bistosylated products at low yield, or no product at all. We then shifted our attention to maltose and lactose as starting disaccharides and first prepared two-fold SiFA-substituted derivatives (Scheme 3).

Whereas the tosylation of β-azido-D-maltose (9a) with pTsCl in pyridine proceeded rather sluggishly, the reactivity was strongly increased in the presence of zinc bromide. Under these conditions, complete conversion was observed after 1 hour at −20 °C, and the bistosylate 10a was isolated in 50% yield. Both leaving groups could be replaced with SiFA moieties under standard conditions using 4 and tBuOK to afford the target molecule 11a in 67% yield. The corresponding SiFA-tagged β-alkynyl-D-maltose 11b was obtained in the same manner from 9b via bistosylate 10b in 31% and 55% yield, respectively.

Similar to the maltose derivatives, the corresponding β-azido- and alkynyl-substituted D-lactoses 12a/b could not be selectively monotosylated at one of the two primary hydroxy groups. Rather, the bistosylates 13a/b were obtained in 44% and 36% yield, which were converted into the two-fold SiFA-modified lactoses 14a/b in moderate yields (33/36%).

It is evident from these results that a protection of one of the two primary hydroxy groups is required for the synthesis of a disaccharide bearing only one SiFA unit. Gratifyingly, the presence of an axial OH group in the galactose ring of β-D-azidolactose 12a allows a selective acetalization to afford product 15 in 74% yield (Scheme 4). For this substrate, the selectivity of the tosylation was examined in detail (Table 1).

With 4 equivalents of zinc bromide and 5 equivalents of tosyl chloride in pyridine at −20 °C, a mixture of the desired monotosylate 16 (41% yield) and the bistosylate 17 (21% yield) was obtained after 10 minutes (Table 1, entry 1). Thus, not only the primary, but also the secondary hydroxy group at C-4′ are reactive under these conditions. As expected, increasing the reaction time up to 40 minutes fa-
vored the formation of the bistosylate 17 (entries 2–4); the best yield of the monotosylate 16 (49%) was obtained after 30 minutes and decreased to 30% after 40 minutes. Smaller amounts of pTsCl afforded a higher selectivity in favor of the monotosylate 16 (entries 5 and 6), which was isolated as the sole product in the presence of 1.3 equivalents of pTsCl (entry 6), albeit in a low yield of 30%. Interestingly, the tosylation of the corresponding lactose derivative bearing a propargyl glycoside instead of the azido group gave only a bistosylate bearing the tosyl groups at the 2-CH2 group and C-4. At this point, it is not clear which factors govern the regioselectivity of these transformations. Unfortunately, attempts to introduce other leaving groups (mesylate, 4-bromophenylsulfonate, bromide) failed.

The structural assignment of tosylation products 16 and 17 is based on extensive NMR studies. Moreover, both products were isolated in crystalline form and characterized by single crystal X-ray diffraction analysis. The quality of the crystals of monotosylate 16 was rather low and allowed only the structural assignment of the constitution, but not of the absolute stereochemistry of the acetal stereocenter. In contrast to this, the acetal stereocenter of the major isomer (87%) of bistosylate 17 (Figure 1) shows S-configuration as estimated by the twin law of the measured crystal.11 The X-ray diffraction analysis also proved the presence of the two tosyl groups at the 2-CH2 and 4′ positions.

The protected monotosylated β-azido-D-lactose derivative 16 obtained in 49% yield under the conditions of Table

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**Scheme 3** Synthesis of two-fold SiFA-substituted maltose and lactose derivatives 11 and 14

**Scheme 4** Synthesis of β-α-azido-substituted LactoSIFA derivative 19
Introduction of an acetal protecting group in β−D-azido-lactose 12a allowed the synthesis of the LactoSiFA derivative 19 in 14% overall yield. Further work is devoted to improved procedures for the monofunctionalization of suitable carbohydrates, as well as, the synthesis of carbohydrate-tagged SiFA derivatives bearing different handles for protein conjugation.

Reactions were carried out under argon atmosphere using oven- or flame-dried glassware. Air- and moisture-sensitive reagents were transferred via syringe. All reagents were obtained commercially and used without further purification. THF, Et₂O, CH₂Cl₂, and MeCN were dried using a MB-8PS-800 system (M. Braun). Reactions were monitored by TLC using silica gel 60 plates provided by Merck and Macherey-Nagel. Visualization was accomplished with UV light (254 nm), ceric ammonium molybdate, KMnO₄, or anisaldehyde.

1H, 13C, 19F, and 26Si NMR spectra were recorded with Bruker Avance III HD (400–600 MHz) and calibrated against residual solvent peaks. IR spectra were obtained with a PerkinElmer Spectrum Two UATR spectrophotometer. Mass spectra were recorded with a Thermo Fisher Scientific TSQ (LCMS-ESI) and a Thermo Electron LTQ Orbitrap spectrometer (HPLC-ESI).

<table>
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<th>Feature</th>
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<td>Figure 1 X-ray crystal structure of bistosylate 17</td>
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Table 1: Tosylation of Disaccharide 15

<table>
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<tr>
<th>Entry</th>
<th>ZnBr₂·2H₂O (equiv)</th>
<th>pTsCl (equiv)</th>
<th>Time (min)</th>
<th>16 (Yield %)</th>
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<tr>
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<td>4</td>
<td>1.3</td>
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*In pyridine at −20 °C.*
A solution of (2R,3S,5S,6R)-2-azido-6-[(4-di-tert-butylfluorosilyl)phenyl]methyl)tetracydro-2H-pyran-3,4,5-triyl)methyl 4-Methylbenzosulfonate (7)  

A solution of (2R,3S,5S,6R)-2-azido-6-[(4-di-tert-butylfluorosilyl)phenyl]methyl)tetracydro-2H-pyran-3,4,5-triol (8) was treated with tBuOK (50 mg, 0.18 mmol, 1.2 equiv) in anhyd DMSO (1 mL). The mixture was stirred at 50 °C for 24 h. Cooling to rt was followed by addition of excess aq 1 M HCl. The mixture was dissolved in Et3O and washed with aq 1 M HCl (3 × 20 mL) and H2O (3 × 20 mL). After drying (MgSO4), the solvent was removed under reduced pressure, and the crude product was purified by column chromatography (silica gel, EtOAc); yield: 36.8 mg (52%); colorless powder.  


A solution of 4 (50 mg, 0.18 mmol, 1.2 equiv) in anhyd DMSO (1 mL) was treated with tBuOK (20 mg, 0.18 mmol, 1.2 equiv). The mixture was stirred at 50 °C for 30 min. After the addition of 3 (54 mg, 0.15 mmol, 1 equiv) in anhyd DMSO (1 mL), the mixture was stirred at 50 °C for 24 h. Cooling to rt was followed by addition of excess aq 1 M HCl. The mixture was dissolved in Et3O and washed with aq 1 M HCl (3 × 20 mL) and H2O (3 × 20 mL). After drying (MgSO4), the solvent was removed under reduced pressure, and the crude product was purified by column chromatography (silica gel) using a gradient starting with pentane/Et3O (15:1) and ending with Et3O/MeOH (20:1); yield: 36.8 mg (52%); colorless powder.  

IR (ATR): 3347, 3293, 2859, 1582, 1470, 1365, 1214, 1106, 1007, 824, 811, 740, 716, 645 cm–1.
A solution of 9a (100 mg, 0.27 mmol, 1.0 equiv) in anhyd pyridine (2.4 mL) was cooled to −20 °C and treated with ZnBr₂·H₂O (243 mg, 1.08 mmol, 4.0 equiv) and then with tPTsCl (257 mg, 1.35 mmol, 5.0 equiv) in anhyd pyridine (1 mL). The mixture was stirred at −20 °C for 1 h. The reaction was carried out three times in separate flasks. The reaction mixtures were combined, the solvent was removed under reduced pressure, and the crude product was purified by column chromatography (silica gel, EtOAc); yield: 367 mg (50%); colorless solid.

IR (ATR): 3325, 2934, 2859, 2117, 1715, 1598, 1534, 1516, 1470, 1373, 1250, 1056, 1012, 939, 816, 740, 645, 600, 504 cm⁻¹.

HRMS (ESI): m/z calc for C₃₉H₄₆N₆O₈S₂Na [M + Na⁺]: 698.1286; found: 698.1295.

A solution of 10b (100 mg, 0.26 mmol, 1.0 equiv) in anhyd pyridine (2.4 mL) was cooled to −20 °C and treated with ZnBr₂·H₂O (237 mg, 1.05 mmol, 4.0 equiv) and then with tPTsCl (248 mg, 1.33 mmol, 5.0 equiv) in anhyd pyridine (1 mL). The mixture was stirred at −20 °C for 1 h. The reaction was carried out twice in separate flasks. The reaction mixtures were combined, the solvent was removed under reduced pressure, and the crude product was purified by column chromatography (silica gel, EtOAc); yield: 115 mg (31%); colorless solid.

IR (ATR): 3284, 2924, 1731, 1598, 1354, 928, 812, 691, 660, 551 cm⁻¹.

HRMS (ESI): m/z calc for C₃₉H₄₆N₆O₈S₂Na [M + Na⁺]: 689.15684; found: 689.15570.
IR (ATR): 3298, 2933, 2859, 1732, 1582, 1471, 1365, 1245, 1062, 1012, 824, 812, 741, 646, 601, 504, 430 cm⁻¹.

1H NMR (600 MHz, DMSO-d₆): δ = 7.44–7.39 (m, 4 H, ArH), 7.36 (d, J = 8.1 Hz, 2 H, ArH), 7.26 (d, J = 8.1 Hz, 2 H, ArH), 5.72 (d, J = 2.6 Hz, 1 H, OH), 5.64 (d, J = 6.6 Hz, 1 H, OH), 5.34–5.29 (m, 2 H, 2 × OH), 5.13 (d, J = 3.7 Hz, 1 H, α-), 5.06 (d, J = 5.1 Hz, 1 H, OH), 4.93 (td, J₁ = 6.5 Hz, J₂ = 3.9 Hz, 1 H, 1H-β₁–1H), 4.11 (d, J = 2.6 Hz, 1 H, 1H-β₂–1H), 3.99 (d, J = 13.6 Hz, 1 H, 1H–β₃–1H), 3.80–3.77 (m, 1 H, H₃–3), 3.53–3.44 (m, 2 H), 3.43–3.34 (m, 7 H), 3.23–3.18 (m, 1 H), 3.14–3.08 (m, 1 H), 3.08–2.98 (m, 2 H), 0.97 [d, J = 8.1 Hz, 36 H, 4 × CH(CH₃)₃].

13C NMR (150 MHz, DMSO-d₆): δ = 140.0, 139.8 (2 s, Ar), 133.9 (d, J = 4.1 Hz, Ar), 133.8 (d, J = 4.1 Hz, Ar), 128.5 (s, J = 13.9 Hz, Ar), 126.4 (d, Ar), 125.8 (d, Ar), 101.2 (d, 100.5 (d, 82.9 (d), 79.3 (s), 77.5 (s), 76.0, 74.2 (2 d), 72.8 (d, J = 7.1 Hz), 72.6, 72.3 (2 d), 72.01 (d, J = 3.4 Hz), 68.8 (d), 58.3 (d), 54.7 (t), 31.4, 28.0 (2 t), 27.0 [q, C(CH₃)₃], 19.8, 19.7 [2 j = 4.7 Hz, C(CH₃)₂].

19F NMR (565 MHz, CDClin): δ = –187.5, –187.5 (2 d, J = 297.8 Hz).

29Si NMR (119 MHz, DMSO-d₆): δ = 15.5, 13.0 (2 d, J = 298.1 Hz).


A solution of 12a (100 mg, 0.27 mmol, 1.0 equiv) in anhyd pyridine (2.4 mL) was cooled to –20 °C with stirring and treated with ZnBr₂·2H₂O (237 mg, 1.0 equiv). The mixture was stirred at –20 °C for 15 min. The reaction was carried out twice in separate flasks. The reaction mixtures were combined, the solvent was removed under reduced pressure, and the crude product was purified by column chromatography (silica gel) using a gradient starting with cyclohexane/EtOAc (100:1) and ending with EtOAc/MeOH (1:1); yield: 160 mg (54%); colorless solid.

IR (ATR): 3369, 2912, 2116, 1728, 1582, 1451, 1352, 1255, 1173, 1060, 971, 811, 769, 689, 662, 550 cm⁻¹.

1H NMR (500 MHz, DMSO-d₆): δ = 7.83–7.76 (m, 4 H, ArH), 7.47–7.45 (m, 4 H, ArH), 5.82 (d, J = 5.6 Hz, 1 H, 1H-β₁–1H), 5.16 (d, J = 3.8 Hz, 1 H, 1H-β₂–1H), 4.96 (d, J = 4.7 Hz, 1 H, 1H-β₃–1H), 4.82 (d, J = 4.7 Hz, 1 H, 1H-β₄–1H), 4.59–4.55 (m, 2 H), 4.46 (d, J = 9.9 Hz, 1 H, 1H–β₅–1H), 4.20–4.10 (m, 3 H), 3.91 (t, J = 9.2 Hz, 1 H), 3.76–3.72 (m, 2 H), 3.30–3.23 (m, 3 H), 3.04–3.01 (m, 1 H), 2.42 (d, J = 1.5 Hz, 6 H, 2 × ArCH₃). OH groups were not detected.

IR (ATR): 3343, 2912, 1728, 1598, 1451, 1352, 1173, 1122, 1049, 1019, 971, 931, 836, 819, 783, 691, 661, 551 cm⁻¹.

1H NMR (500 MHz, DMSO-d₆): δ = 7.84–7.77 (m, 4 H, ArH), 7.50–7.46 (m, 4 H, ArH), 5.46 (d, J = 5.4 Hz, 1 H, 1H-β₁–1H), 5.16 (d, J = 4.1 Hz, 1 H, 1H-β₂–1H), 4.96 (d, J = 5 Hz, 1 H, 1H-β₃–1H), 4.82 (d, J = 4.9 Hz, 1 H, 1H-β₄–1H), 4.54 (d, J = 2.1 Hz, 1 H, 1H-β₅–1H), 4.47 (d, J = 9.3 Hz, 1 H, 1H–β₆–1H), 4.31 (d, J = 7.8 Hz, 1 H, 1H–β₇–1H), 4.23–4.07 (m, 5 H), 4.02 (d, J = 6.8 Hz, 1 H, 1H–β₈–1H), 3.91 (dd, J₁ = 9.9 Hz, J₂ = 8.5 Hz, 1 H, 1H–β₉–1H), 3.78–3.76 (m, 1 H), 3.59–3.51 (m, 3 H), 3.27–3.24 (m, 1 H), 3.04–2.97 (m, 1 H), 2.43 (s, 6 H, 2 × ArCH₃).

IR (ATR): 1258, 1257, 1256, 1255 (7 d, Ar), 1028 (d, CHO), 89.2 (s, CH₂), 78.5, 74.0, 73.3, 72.6, 72.4, 72.2 (6 H, CH₃), 69.9 (t, CH₂OTs), 68.9 (d, CH₂), 62.2 (s, ArCH₂).
A solution of 15 (100 mg, 0.22 mmol, 1.0 equiv) in anhyd pyridine (2 mL) was cooled to −20 °C and treated with ZnBr₂·2 H₂O (198 mg, 0.88 mmol, 4.0 equiv) and then with pTsCl (209 mg, 1.1 mmol, 5.0 equiv) in anhyd pyridine (1 mL). The mixture was stirred at −20 °C for 30 min. The solvent was removed under reduced pressure, and the crude product was purified by column chromatography (silica gel) using a gradient starting with EtOAc/cyclohexane (1:1) and ending with EtOAc/MeOH (30:1, 1% Et₃N) and ending with EtOAc/MeOH (30:1, 1% Et₃N) and ending with EtOAc/MeOH (30:1, 1% Et₃N).

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Yield: 66 mg (49%); colorless solid.

IR (ATR): 3334, 2911, 2113, 1598, 1359, 1248, 1174, 1095, 1033, 994, 964, 903, 839, 810, 774, 741, 698, 670, 597, 554, 520, 481 cm⁻¹.

1H NMR (600 MHz, DMSO-d₆): δ = 7.76 (dd, J = 8.1 Hz, J₂ = 7.2 Hz, 1 H), 7.44 (dd, J₁ = 6.1 Hz, J₂ = 7.2 Hz, 2 H, ArH), 7.41–7.35 (m, 5 H, ArH), 5.74 (d, J = 5.6 Hz, 1 H, OH), 5.56 (s, 1 H), 5.19 (dd, J₁ = 4.1 Hz, J₂ = 1.0 Hz, 2 H, ArH), 5.06–4.99 (m, 2 H, OH), 4.63–4.56 (m, 2 H), 4.33 (d, J = 7.4 Hz, 1 H), 4.13 (dd, J₁ = 11.0 Hz, J₂ = 7.2 Hz, 1 H), 4.09–4.05 (m, 2 J), 3.96 (d, J = 12.2 Hz, 1 H), 3.79 (t, J = 8.3 Hz, 1 H), 3.58 (s, 1 H), 3.48–3.39 (m, 3 H), 3.36 (d, J = 9.5 Hz, 1 H), 3.03 (td, J₁ = 8.6 Hz, J₂ = 5.8 Hz, 1 H), 2.38 (s, 3 H, ArCH₃).

13C NMR (150 MHz, DMSO-d₆): δ = 145.3 (s, Ar), 139.0 (s, Ar), 132.7 (s, Ar), 130.5, 129.2 (2 d, Ar), 128.4 (d, Ar), 128.1, 126.7 (2 d, Ar), 102.8 (d), 100.2 (d), 89.7 (d), 77.5, 76.1, 74.6, 73.9, 73.3, 72.0, 70.0 (7 d), 69.8, 68.9 (2 t), 66.8 (d, 21.6 ArCH₃).

HRMS (ESI): m/z calcd for C₂₉H₂₆N₄O₉SNa [M + Na⁺]: 610.17012; found: 610.17125.

17

Yield: 52 mg (31%); colorless solid.

IR (ATR): 3486, 3037, 2875, 2116, 1729, 1598, 1357, 1246, 1173, 1093, 1043, 963, 905, 868, 812, 697, 669, 552 cm⁻¹.

1H NMR (600 MHz, DMSO-d₆): δ = 7.82 (d, J = 8.2 Hz, 2 H, ArH), 7.74 (d, J = 8.2 Hz, 2 H, ArH), 7.42 (d, J = 8.2 Hz, 2 H, ArH), 7.39–7.35 (m, 5 H, ArH), 7.29–7.26 (m, 2 H, ArH), 5.76 (d, J = 5.5 Hz, 1 H, OH), 5.61 (d, J = 5.5 Hz, 1 H, OH), 5.35 (s, 1 H), 5.04 (d, J = 3.1 Hz, 1 H, OH), 4.62–4.51 (m, 4 H), 4.48 (d, J = 7.6 Hz, 1 H), 4.25 (d, J = 3.4 Hz, 1 H), 4.03–4.09 (m, 2 H), 3.99–3.95 (m, 1 H), 3.78–3.73 (m, 1 H), 3.70 (s, 1 H), 3.54 (ddd, J₁ = 9.5 Hz, J₂ = 7.9 Hz, J₃ = 5.5 Hz, 1 H), 3.43–3.36 (m, 2 H), 3.01 (td, J₁ = 8.5 Hz, J₂ = 5.8 Hz, 1 H), 2.38 (d, J = 3.7 Hz, 6 H, 2 x ArCH₃).

13C NMR (150 MHz, DMSO-d₆): δ = 145.1 (s, Ar), 144.9 (s, Ar), 138.1 (s, Ar), 134.0 (s, Ar), 132.3 (s, Ar), 130.2, 129.1, 128.2 (4 d, Ar), 127.8 (d, J = 13.6 Hz, Ar), 126.2 (d, Ar), 120.8 (d), 100.2 (d), 89.7 (d), 77.5, 76.1, 74.6, 73.9, 73.3, 72.0, 70.0 (7 d), 69.8, 68.9 (t), 66.8 (d), 21.6 (q, ArCH₃).

LRMS (ESI): m/z calcd for C₂₉H₂₆N₄O₉SNa [M + Na⁺]: 786.16; found: 786.32.

X-ray Crystal Data

Intensity data for the colorless crystal of compound 17 were collected on a D8 Venture Bruker Diffractometer, SC-XRD using Cu-Kα radiation at 173(2) K. The molecule structure was solved with direct methods using SHELXS-2014/7 or SHELXT-2014/7 and refinements were carried out using F2 by using SHELXL-2014/7 or OLEX2. The data obtained by the measurement were treated in the refinement procedure as a 2-component twin. Applying the TwinRotMat option in the program PLATON revealed a twin law (BASF 0.13258).

A solution of 4 (110 mg, 0.41 mmol, 1.2 equiv) in anhyd DMSO (3 mL) was treated with bBuOK (45.8 mg, 0.41 mmol, 1.2 equiv). The mixture was stirred at 50 °C for 30 min. After the addition of 16 (207 mg, 0.34 mmol, 1 equiv) in anhyd DMSO (1 mL), the mixture was stirred at 50 °C for 2 d. Cooling to rt was followed by addition of excess aq 1 M HCl. The mixture was dissolved in Et₂O and washed with aq 1 M HCl (3 x 20 mL) and H₂O (3 x 20 mL). After drying (MgSO₄), the solvent was removed under reduced pressure, and the crude product was purified by column chromatography (silica gel) using a gradient starting with cyclohexane/EtOAc (2:1) and ending with EtOAc; yield: 146 mg (61%); colorless solid.

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IR (ATR): 3385, 2934, 2933, 2859, 2115, 1733, 1582, 1471, 1365, 1245, 1163, 1027, 901, 812, 740, 699, 641, 505, 431 cm⁻¹.

1H NMR (600 MHz, DMSO-d₆): δ = 7.47 (d, J = 7.7 Hz, 4 H, ArH), 7.42–7.35 (m, 5 H, ArH), 5.70 (d, J = 5.5 Hz, 1 H, OH), 5.58 (s, 1 H), 5.47 (d, J = 4.4 Hz, 1 H, OH), 5.13–5.07 (m, 1 H, OH), 4.83–4.77 (m, 1 H, OH), 4.64 (d, J = 8.4 Hz, 1 H), 4.50–4.46 (m, 1 H), 4.11 (d, J = 2.6 Hz, 2 H), 4.00 (d, J = 12.1 Hz, 1 H), 3.85–3.81 (m, 1 H), 3.75 (br s, 1 H), 3.65 (s, 1 H), 3.54–3.47 (m, 2 H), 3.45–3.43 (m, 2 H), 3.18–3.08 (m, 2 H), 1.01 [s, 18 H, 2 × C(CH₃)₃].

13C NMR (150 MHz, DMSO-d₆): δ = 139.8, 138.9, 135.3 (3 s, Ar), 134.4 (d, J = 4.4 Hz, Ar), 129.1, 128.3, 126.6, 126.2 (4 d, Ar), 103.7 (d, C-1'), 100.1 (d), 89.9 (d), 82.3, 76.1, 75.8, 74.5, 73.7, 72.2, 70.3 (7 d), 68.8 (t), 66.8 (d), 32.7 (t), 28.4, 27.4 [2 q, C(CH₃)], 20.2 [d, J = 12.1 Hz, C(CH₃)].

19F NMR (565 MHz, CDCl₃): δ = –187.4 (d, J = 296.5 Hz).

29Si NMR (119 MHz, CDCl₃): δ = 15.6 (d, J = 297.1 Hz).


Supporting Information
Supporting information for this article is available online at https://doi.org/10.1055/s-0037-1611656.

References
(11) CCD C1866515 contains the supplementary crystallographic data for this paper. The data can be obtained free of charge from The Cambridge Crystallographic Data Centre via www.ccdc.cam.ac.uk/getstructures.