Asymmetric Total Synthesis and Biological Evaluation of (+)-Cycloclavine

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Published as part of the 50 Years SYNTHESIS – Golden Anniversary Issue

Received: 29.10.2018
Accepted: 31.10.2018
Published online: 20.11.2018

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Abstract The first total synthesis of natural (+)-cycloclavine uses a catalytic asymmetric cyclopropanation of allene, a regiospecific Pd-catalyzed enone formation, and two intramolecular Diels–Alder reactions for indole/indoline annulations. The binding properties of natural (+)- and unnatural (−)-cycloclavine on 16 CNS receptors revealed significant stereospecificity and unique binding profiles in comparison to LSD, psilocin, and DMT. Differential 5-HT affinities, as well as novel sigma-1 receptor properties bode well for potential therapeutic developments of clavine alkaloid scaffolds.

Key words clavine ergot alkaloids, enantioselective allene cyclopropanation, psychedelics, stereospecific GPCR binding, LSD, psilocin, DMT, 5-HTA, sigma-1 receptors

While the unique properties of naturally occurring compounds have always fascinated researchers from all branches of Science, the total synthesis of alkaloids currently experiences a remarkable renaissance, motivated by the complex architectures, diverse functionalities, and profound biological and cultural impact of this large family of natural products.1 Indole alkaloids, in particular, are attracting significant attention in Chemistry and Medicine.2 With the goal to explore both innovative synthetic strategies and new biological applications, we have recently established a program in the total synthesis of ergot alkaloids of the clavine and lysergic acid subclasses (Figure 1).3

In 1969, A. Hofmann and co-workers at Sandoz in Basel, Switzerland, reported the isolation of a novel, cyclopropane-containing ergot alkaloid, (+)-cycloclavine, from the seeds of the morning glory Ipomoea hildebrandtii VAT-KE, collected in Nairobi, Kenya.4 After a latency of almost 40 years, cycloclavine has now become a popular target for organic synthesis, and a number of groups have reported innovative synthetic approaches. In pioneering studies from the group of Szántay, 4-bromo-Uhle’s ketone was subjected to an alkylation and intramolecular aldol reaction for indole/indoline annulations. The binding properties of natural (+)- and unnatural (−)-cycloclavine on 16 CNS receptors revealed significant stereospecificity and unique binding profiles in comparison to LSD, psilocin, and DMT. Differential 5-HT affinities, as well as novel sigma-1 receptor properties bode well for potential therapeutic developments of clavine alkaloid scaffolds.
opened two formal syntheses of racemic cycloclavine and a formal synthesis of (+)-cycloclavine starting from substituted indoles and intersecting with the late stage alkene in Szántay's synthesis.9

Another formal synthesis of (±)-cycloclavine that converged with Szántay's approach was accomplished by Netz and Opatz in 2016, utilizing a β-alkylation of a pyrrolinone followed by a Heck coupling.10

The first asymmetric synthesis of (–)-cycloclavine, the enantiomer of the natural alkaloid, was accomplished by our group in 2017.11 Key features of this synthesis were a catalytic asymmetric cyclopropanation of allene, an intramolecular Diels–Alder reaction to methylenecyclopropane (IMDAMC), and an intramolecular Diels–Alder reaction to furan (IMDAF). Subsequently, a formal synthesis of both enantiomers of cycloclavine was realized by Bisai and co-workers based on a D- or L-proline catalyzed α-aminoxylatation and a Heck coupling (Scheme 2).12 Most recently, Dong and co-workers developed a benzyne cycloaddition/alkene carboacylation route to both (–)-5-epi-cycloclavine and (–)-cycloclavine, utilizing a ring-enlargement of a benzocyclobutenone intermediate as a key reaction.13 The impressive publication surge and the diverse strategies of these synthetic approaches illustrate the high level of current interest in architecturally novel alkaloid natural products. We now report the details of the first enantioselective total synthesis of (+)-cycloclavine.

Our retrosynthetic analysis is summarized in Scheme 3. In analogy to our route to (–)-cycloclavine,11 we selected an asymmetric rhodium-catalyzed cyclopropanation of allene with a diazopropanoate active ester, followed by an aminolysis with 4-(methylamino)but-3-en-2-one, for the assembly of the key precursor for the IMDAMC reaction. After installing the enone in the six-membered ring by a Diao–Stahl ketone dehydrogenation,14 the thermally removable Tempoc group for amine protection15 would be used to stabilize an aminomethylithium reagent and favor enone 1,2-addition versus lactam ring opening. The final indole ring fusion was envisioned to be accomplished by the IMDAF cycloaddition.6b,d,f,7,16

Biographical Sketches

Stephanie R. McCabe received her B.Sc. (Honours) in 2012 from the Australian National University, conducting research under the supervision of Professor Martin Banwell. In 2018, she obtained her Ph.D. in the Wipf group at the University of Pittsburgh. Her research interests centered on natural product total synthesis.

Peter Wipf received his Ph.D. in 1987 from the University of Zürich under the direction of Professor Heinz Heimgartner. He then joined the laboratory of Professor Robert E. Ireland at the University of Virginia as a Swiss NSF postdoctoral fellow, and, in 1990, the University of Pittsburgh as an Assistant Professor. Since 2004, he is the Distinguished University Professor of Chemistry at the University of Pittsburgh. Wipf’s research focuses on the total synthesis of natural products, organometallic, heterocyclic and medicinal chemistry.
Among the enantioselective variants, bulky carboxylates derived from L-tert-leucine, such as Rh$_6$(S-PTTL)$_4$ (8), were found to be particularly effective. More recently,
Hashimoto et al. showed that the substrate scope could be further expanded when the dirhodium complex \( \text{Rh}_2(\text{S-TBPTTL})_4 \) (9) was used as the catalyst (Figure 2).\(^{18}\) We also prepared the enantiomer of 9, \( \text{Rh}_2(\text{R-TBPTTL})_4 \) (10), from \( \text{d-}\text{tert-leucine} \) ([R]-18) and anhydride 17 in toluene at reflux, followed by a ligand exchange reaction with \( \text{Rh}_2(\text{OAc})_4 \) in a chlorobenzene/MeCN mixture at 130 °C (Scheme 6). Furthermore, we reasoned that the 4,7-diphenyl substitution pattern on the phthalimide ring of the novel, sterically demanding dirhodium catalyst 11 would impose even greater steric discrimination than the corresponding bromide substituents in 9 and 10, hopefully leading to greater differentiation between the enantiotopic faces of the allene. This catalyst was prepared in a Diels-Alder reaction of diphenylbutadiene (19) and maleic anhydride (20), followed by DDQ oxidation to afford anhydride 21, which was reacted with \( \text{(S)}-18 \) in the presence of triethylamine to give 22 and subjected to a ligand exchange reaction to yield 11 (Scheme 7).

The results of the cyclopropanation of allene (23) with pentafluorophenyl diazopropanoate (4) to give methylenecyclopropane 24 in the presence of the chiral catalysts 8–16 are summarized in Table 1. Rh(II)-Catalysts with sterically hindered amino acid ligands but lacking phthalimide substituents, such as 8 and 12, provided a low e.r. of approximately 7:3 (Table 1, entries 1 and 5). Hashimoto’s tetrabromophthaloyl tert-leucine dirhodium Catalyst 9 resulted in a notable improvement, giving the cyclopropane (R)-24 in a high yield with an e.r. of 87:13 (entry 2). As expected, the (R)-tert-leucine derived 10 gave the enantio-meric product (S)-24 in identical yield and e.r. (entry 3). Disappointingly, however, virtually no enantioinduction was observed when allene was reacted with 4 in the presence of the sterically more demanding dirhodium catalyst 11 (product e.r. = 55:45, entry 4). With this catalyst, no reaction occurred at –78 °C and the mixture had to be warmed to –40 °C before conversion was observed. The chiral cyclopropane catalyst 13 delivered the desired product 24 in moderate yields and with poor enantioinduction (entry 6). Davies’ proline-based catalyst 14 also provided only a moderate yield of 61%, and negligible asymmetric induction (entry 7). The reaction of Ru(II)-catalyst 15 did not deliver any of the desired product. Instead, upon warming the reaction mixture from –78 °C to room temperature, full conversion into the undesired \( \beta \)-H migration product 25 was observed (entry 8). Similarly, the (salen)cobalt(II) catalyst 16 showed no catalytic activity even at room temperature (entry 9).

In our synthetic route, the cyclopropanation of allene (23) with diazopropanoate 4 in the presence of 1 mol% of the dirhodium catalyst \( \text{Rh}_2(\text{R-TBPTTL})_4 \) (10) provided the enantiomerically enriched methylenecyclopropane (S)-24 on multi-gram scale. Ester aminolysis with the lithium salt of 26 gave the vinylogous imide 27 in 80% yield and 87:13 e.r. (Scheme 8). Deprotonation of 27 with NaHMDS in THF at –78 to –50 °C formed the corresponding sodium enolate, which was trapped with TBSCI to give the silyl enol ether intermediate 28. When heated at 95 °C in THF in the micro-

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**Figure 2** Cyclopropanation catalysts

**Scheme 6** Preparation of chiral dirhodium catalyst 10

**Scheme 7** Preparation of chiral dirhodium catalyst 11

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wave reactor, this diene underwent the intramolecular strain-promoted Diels–Alder (IMDAMC) reaction, and TBAF cleavage of the product silyl enol ether formed the desired trans-adduct 30 in 66% yield along with the cis-adduct 29 in 15% yield. These two diastereomers were readily separated chromatographically. Dehydrogenation of 30 under modified Diao–Stahl conditions \(^{14}\) with Pd(TFA)\(_2\) and DMSO in AcOH under an atmosphere of oxygen at 55 °C gave the corresponding enone 31 in 66% yield as a single regioisomer. Interestingly, dehydrogenation of the epimer 29 under these conditions provided the opposite enone regioisomer 32 stereospecifically in 56% yield. This complete switch in regioselectivity for the cis- and trans-diastereomers 29 and 30 parallels results obtained for the enolization of cis- and trans-2-decalones.\(^{22}\) The configuration at the decalin ring junction governs the regioselectivity of enolization process due to torsional strain effects. The torsional strain that was proposed to govern the regiochemistry of enolization in cis- and trans-decalones has also been investigated in greater detail in relevant cis- and trans-octalins.\(^{23a,b}\)

Enone 31 was obtained as a crystalline solid, and its enantiomeric ratio could be further enriched by recrystallization from 87:13 to yield product with >99% e.r. Chiral SFC analysis was used to evaluate each batch for enantiomeric purity.

For the completion of the total synthesis, stannane 34 was treated with n-BuLi at low temperature and converted into the corresponding lithium carbamion (Scheme 9). Addition of enone 31 to the reaction mixture delivered two diastereomeric allylic alcohols, which could be separated by chromatography on silica gel to give α-alcohol α-35 and β-alcohol β-35 in 41% and 32% yield, respectively. The intramolecular Diels–Alder (IMDAMF) reaction of α-35 (bearing a pseudo-equatorial hydroxy group) at 135 °C in a sealed tube was followed by spontaneous aromatization and cleavage of the Tempoc protecting group under the reaction conditions to form indole 36. The stereoisomeric alcohol β-35 was inert under these conditions, quite likely due to steric strain in the transition state that requires a pseudo-axial position of the substituent bearing the furan ring. Finally, lactam reduction led to (+)-cycloclavine in 34% yield over two steps from α-35. Overall, the synthesis of the natural enantiomer was accomplished in 8 steps and 4% yield. The specific rotation of the synthetic material was determined

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**Table 1** Catalytic Asymmetric Cyclopropanation of Allene: Catalyst Screen

<table>
<thead>
<tr>
<th>Entry</th>
<th>Catalyst</th>
<th>Temperature</th>
<th>Solvent</th>
<th>Major product</th>
<th>Yield (%)</th>
<th>e.r.*</th>
<th>Major enantiomer</th>
</tr>
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<tbody>
<tr>
<td>1</td>
<td>8</td>
<td>–78 °C</td>
<td>hexanes</td>
<td>24</td>
<td>83</td>
<td>72:28</td>
<td>R</td>
</tr>
<tr>
<td>2</td>
<td>9</td>
<td>–78 °C</td>
<td>CH(_2)Cl(_2)</td>
<td>24</td>
<td>86</td>
<td>87:13</td>
<td>R</td>
</tr>
<tr>
<td>3</td>
<td>10</td>
<td>–78 °C</td>
<td>CH(_2)Cl(_2)</td>
<td>24</td>
<td>86</td>
<td>87:13</td>
<td>S</td>
</tr>
<tr>
<td>4</td>
<td>11</td>
<td>–78 °C to –40 °C</td>
<td>CH(_2)Cl(_2)</td>
<td>24</td>
<td>78</td>
<td>55:45</td>
<td>R</td>
</tr>
<tr>
<td>5</td>
<td>12</td>
<td>–78 °C</td>
<td>hexanes</td>
<td>24</td>
<td>84</td>
<td>73:27</td>
<td>R</td>
</tr>
<tr>
<td>6</td>
<td>13</td>
<td>–78 °C</td>
<td>CH(_2)Cl(_2)</td>
<td>24</td>
<td>59</td>
<td>59:41</td>
<td>R</td>
</tr>
<tr>
<td>7</td>
<td>14</td>
<td>–78 °C</td>
<td>hexanes</td>
<td>24</td>
<td>61</td>
<td>53:47</td>
<td>R</td>
</tr>
<tr>
<td>8</td>
<td>15</td>
<td>–78 °C to r.t.</td>
<td>CH(_2)Cl(_2)</td>
<td>25</td>
<td>ND*</td>
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<td>–</td>
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<tr>
<td>9</td>
<td>16(^{c})</td>
<td>–78 °C to r.t.</td>
<td>THF</td>
<td>–</td>
<td>–</td>
<td>–</td>
<td>–</td>
</tr>
</tbody>
</table>

* Enantiomeric ratios were determined by chiral SFC analysis of the corresponding imide 27.

\(^{a}\) ND: Not determined.

\(^{b}\) 10 mol% N-methylimidazole was added.
to be +61.4 (c 0.2, CHCl₃), which was consistent with the literature value +63 (c 1, CHCl₃). Mass spectra, IR, ¹H, and ¹³C NMR data were also consistent with the previously reported data for the natural product as well as its enantiomer.⁴,¹¹

Scheme 9 Completion of total synthesis of (+)-cycloclavine

In contrast to the vast literature on lysergic acid derivatives, relatively little is known about the pharmaceutical potential of clavine ergot alkaloids.²b Lysergic acid derivatives, most notably lysergic acid diethylamide (LSD) have significant hallucinogenic properties that can interfere with their therapeutic potential. Most of these effects are thought to be mediated through agonist action at the 5-hydroxytryptamine receptor 2A, 5-HT₂A. In the tailwind of the rapidly expanding medical uses of cannabinoids, the mushroom metabolite psilocybin, and even LSD, are now moving to the forefront of clinical research on the management of mental health, anxiety, neurodegeneration, and substance-use disorders.²⁴ It would appear that more fundamental research on efficacy, tolerability, and safety of serotonergic psychedelics is highly warranted.

It is well known that binding of lysergic acid derivatives to brain membrane receptors is stereospecific, since L-LSD, the psychotropically inactive enantiomer of LSD, is ca. 1000 times weaker as a brain membrane receptor radioligand displacing agent,²⁵ and l-LSD as well as the other diastereomers, d-iso-lysergic acid diethylamide (iso-LSD) and l-iso-lysergic acid diethylamide (l-iso-LSD), show no psychic effects in humans up to a dose of 0.5 mg, which corresponds to a 20-fold increase over a still distinctly active D-LSD dose.²⁶ Having gained ready synthetic access to both natural (+)-cycloclavine and its unnatural enantiomer (−)-cycloclavine,¹¹ we were therefore interested to determine the receptor profiles of both compounds, and compare them to other serotonergic agents.

For an initial survey, we selected 13 pertinent CNS receptors and profiled both enantiomers at 10 µM concentration (Table 2). Compound binding was calculated as a percent inhibition of a radioactively labeled ligand specific for each target. As a group, the cycloclavines were more selective in this receptor panel than D-LSD,²⁷,²⁸ the bioactive LSD stereoisomer. D-LSD was active at the adrenergic ³¹ and histamine H₁ receptors (Table 2, entries 1 and 6), whereas both

<table>
<thead>
<tr>
<th>Entry</th>
<th>Receptor</th>
<th>(+)-Cycloclavine [10 µM]</th>
<th>(+)-Cycloclavine [1 µM]</th>
<th>(-)-Cycloclavine [10 µM]</th>
<th>(-)-Cycloclavine [1 µM]</th>
<th>D-LSD [10 µM]</th>
</tr>
</thead>
<tbody>
<tr>
<td>1</td>
<td>Adrenergic α₁</td>
<td>57</td>
<td>59</td>
<td>100⁶</td>
<td></td>
<td></td>
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<tr>
<td>2</td>
<td>Dopamine D₁</td>
<td>91</td>
<td>30</td>
<td>93</td>
<td></td>
<td></td>
</tr>
<tr>
<td>3</td>
<td>Dopamine D₃</td>
<td>81</td>
<td>14</td>
<td>85</td>
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<td></td>
</tr>
<tr>
<td>4</td>
<td>Dopamine D₃</td>
<td>93</td>
<td>75</td>
<td>64</td>
<td>32</td>
<td>78</td>
</tr>
<tr>
<td>5</td>
<td>GABAₐ</td>
<td>9</td>
<td>0</td>
<td>0</td>
<td></td>
<td></td>
</tr>
<tr>
<td>6</td>
<td>Histamine H₁</td>
<td>17</td>
<td>27</td>
<td>85⁵</td>
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<td>7</td>
<td>Muscarinic M₂</td>
<td>0</td>
<td>0</td>
<td>2</td>
<td></td>
<td></td>
</tr>
<tr>
<td>8</td>
<td>Muscarinic M₃</td>
<td>11</td>
<td>15</td>
<td>15</td>
<td></td>
<td></td>
</tr>
<tr>
<td>9</td>
<td>Nicotinic acetylcholine α4β2</td>
<td>9</td>
<td>11</td>
<td>0</td>
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<td>10</td>
<td>Opiate κ</td>
<td>73</td>
<td>75</td>
<td>29</td>
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</tr>
<tr>
<td>11</td>
<td>Orexin OX₁</td>
<td>23</td>
<td>6</td>
<td>ND⁴</td>
<td></td>
<td></td>
</tr>
<tr>
<td>12</td>
<td>Serotonin 5-HT₁A</td>
<td>97</td>
<td>97</td>
<td>91</td>
<td>73</td>
<td>73</td>
</tr>
<tr>
<td>13</td>
<td>Serotonin 5-HT₂A</td>
<td>100</td>
<td>100</td>
<td>89</td>
<td>51</td>
<td>93</td>
</tr>
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</table>

* Biochemical assays were performed in duplicate at human receptors at Eurofins Cerep Panlabs and are presented as the percent inhibition of specific binding or activity of a radioligand, unless otherwise indicated.

³⁴ Data from ref. 27.

⁵ Rat receptor data.

⁶ Data from ref. 28.

⁴ ND: Not determined.
cycloclavines were moderately active at the opiate \( \kappa \) receptor (entry 10). Neither ergot chemotype showed significant activity at \( \text{GABA}_A \), muscarinic \( M_2 \) and \( M_3 \), and nicotinic acetylcholine \( \alpha_4 \beta_2 \) receptors (entries 5, 7, 8, and 9). We were unable to find LSD data on orexin \( \text{OX}_1 \), but cycloclavine did not perturb radioligand binding at this site at a 10 \( \mu \text{M} \) concentration (entry 11).

Significant differences between (+)- and (−)-cycloclavine revealed themselves in the dopamine \( \text{D}_1 \), \( \text{D}_2 \), and \( \text{D}_3 \) monoamine receptor family (Table 2, entries 2–4). In close analogy to D-LSD, natural (+)-cycloclavine maintained strong affinity to these receptors, which stimulate cognitive and motor functions. (−)-Cycloclavine showed comparatively moderate activity at the dopamine \( \text{D}_1 \) receptor at 10 \( \mu \text{M} \) concentration, but fell below the threshold of 50% inhibition at 1 \( \mu \text{M} \), whereas (+)-cycloclavine still maintained significant binding at this concentration (entry 4). A less prominent but still distinctive stereospecificity was observed at the serotonin 5-HT\(_{1A}\) and 5-HT\(_{2A}\) receptors (entries 12 and 13). Natural (+)-cycloclavine had very potent binding properties at both 10 and 1 \( \mu \text{M} \), whereas (−)-cycloclavine tailed off at 1 \( \mu \text{M} \). Serotonin receptors regulate a plethora of behavioral responses, from aggression, anxiety, appetite, to learning, memory, sleep, and even aging.25 D-LSD is one of the most potent agonists at 5-HT\(_{1A}\) and the affinity at the 5-HT\(_{2A}\) and possibly the 5-HT\(_{2C}\) receptors versus the 5-HT\(_{1A}\) receptor correlates with the mental effects of psychedelics in humans.30 In view of this interesting stereospecificity, and the significance of 5-HT receptors to human behavior, we decided to pursue additional studies on 5-HT subtypes (Table 3).

The purpose of our second generation functional assays on human 5-HT receptors was to determine effective concentrations \( \text{EC}_{50} \) or inhibitory constants (\( K_i \)) for (+)-cycloclavine and (−)-cycloclavine. Cellular agonist effects were calculated as a percentage of a validated control activity at both 10 and 1 \( \mu \text{M} \), whereas (−)-cycloclavine revealed themselves in the dopamine \( \text{D}_1 \), \( \text{D}_2 \), and \( \text{D}_3 \) monoamine receptor family (Table 2, entries 2–4). In close analogy to D-LSD, natural (+)-cycloclavine maintained strong affinity to these receptors, which stimulate cognitive and motor functions. (−)-Cycloclavine showed comparatively moderate activity at the dopamine \( \text{D}_1 \) receptor at 10 \( \mu \text{M} \) concentration, but fell below the threshold of 50% inhibition at 1 \( \mu \text{M} \), whereas (+)-cycloclavine still maintained significant binding at this concentration (entry 4). A less prominent but still distinctive stereospecificity was observed at the serotonin 5-HT\(_{1A}\) and 5-HT\(_{2A}\) receptors (entries 12 and 13). Natural (+)-cycloclavine had very potent binding properties at both 10 and 1 \( \mu \text{M} \), whereas (−)-cycloclavine tailed off at 1 \( \mu \text{M} \). Serotonin receptors regulate a plethora of behavioral responses, from aggression, anxiety, appetite, to learning, memory, sleep, and even aging.25 D-LSD is one of the most potent agonists at 5-HT\(_{1A}\) and the affinity at the 5-HT\(_{2A}\) and possibly the 5-HT\(_{2C}\) receptors versus the 5-HT\(_{1A}\) receptor correlates with the mental effects of psychedelics in humans.30 In view of this interesting stereospecificity, and the significance of 5-HT receptors to human behavior, we decided to pursue additional studies on 5-HT subtypes (Table 3).

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As suggested by the preliminary assays, (+)-cycloclavine provided considerably more potent at the 5-HT\(_{1A}\) receptor than (−)-cycloclavine with an activation potency \( \text{EC}_{50} = 0.14 \) \( \mu \text{M} \) versus ~5 \( \mu \text{M} \) for (−)-cycloclavine (Table 3, entry 1). Both stereoisomers are poor activators at 5-HT\(_{2A}\), suggesting that hallucinogenic or strongly euphoric effects in humans might be limited in comparison to D-LSD, even though (+)-cycloclavine displays its most potent activation potential \( \text{EC}_{50} = 16 \) nM at 5-HT\(_{2C}\), a receptor that is thought to contribute to the observed mental effects of psychedelic drugs (entries 2 and 4). With the exception of DMT, which has only moderate potency, none of the tested agents activated 5-HT\(_{2B}\), a 5-HT receptor subtype that has been associated with cardiotoxicity. Overall, the 5-HT profile of (+)-cycloclavine closely mirrors that of psilocin, and to a lesser extent, that of DMT. It is substantially different from D-LSD, a property that we believe bodes well for future therapeutic investigations of this compound class.

The unusual activity on the opioid \( \kappa \) receptor, and the relative similarity to psilocin and DMT in the 5-HT panel inspired us to also evaluate the activity of cycloclavines in the sigma-1 assay, a receptor that was originally mischaracterized as an opioid receptor and has now been implicated in neuroinflammation and neuroprotection.36 DMT was identified as an endogenous sigma-1 receptor regulator.33,37 Surprisingly, while (+)-cycloclavine was inactive, the unnatural (−)-cycloclavine was determined to have a \( K_i \) = 8.3 \( \mu \text{M} \) for

### Table 3 Effects of Cycloclavines, DMT, Psilocin, and D-LSD on 5-HT Subtypes and Sigma-1 Receptors\(^{a,b}\)

<table>
<thead>
<tr>
<th>Entry</th>
<th>Receptor</th>
<th>(+)-Cycloclavine [( \mu \text{M} )]</th>
<th>(−)-Cycloclavine [( \mu \text{M} )]</th>
<th>DMT(^{a} ) [( \mu \text{M} )]</th>
<th>Psilocin(^{c} ) [( \mu \text{M} )]</th>
<th>D-LSD(^{c} ) [( \mu \text{M} )]</th>
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</thead>
<tbody>
<tr>
<td>1</td>
<td>Serotonin 5-HT(_{1A})</td>
<td>0.14</td>
<td>–5</td>
<td>0.075(^{d} )</td>
<td>0.123(^{d} )</td>
<td>0.003(^{d} )</td>
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<td>2</td>
<td>Serotonin 5-HT(_{2A})</td>
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<td>&gt;50</td>
<td>0.076</td>
<td>0.721</td>
<td>0.261</td>
</tr>
<tr>
<td>3</td>
<td>Serotonin 5-HT(_{2B})</td>
<td>&gt;20</td>
<td>&gt;20</td>
<td>3.4</td>
<td>&gt;20</td>
<td>12</td>
</tr>
<tr>
<td>4</td>
<td>Serotonin 5-HT(_{2C})</td>
<td>0.016</td>
<td>3.2</td>
<td>0.424(^{d} )</td>
<td>0.094(^{d} )</td>
<td>0.015(^{d} )</td>
</tr>
<tr>
<td>5(^{d})</td>
<td>Sigma-1</td>
<td>–50</td>
<td>8.3</td>
<td>5.2(^{c} )</td>
<td>&gt;10(^{c} )</td>
<td>ND(^{c} )</td>
</tr>
</tbody>
</table>

\(^{a}\) Biochemical assays were performed in duplicate at human receptors at Eurofins Cerep Panlabs and results are based on 5-point concentration response curves, unless otherwise indicated.

\(^{b}\) Activation potency \( \text{EC}_{50} \) values are shown, unless otherwise specified.

\(^{c}\) Data from ref. 31, unless otherwise specified.

\(^{d}\) Inhibition constants \( K_i \).

\(^{e}\) Data from ref. 32.

\(^{f}\) ND: Not determined.
the inhibition of the binding of the radiolabeled agonist haloperidol to sigma-1, and therefore found to be very similar to DMT (Kᵢ = 5.2 µM) (Table 3, entry 5). To the best of our knowledge, this is the first time that stereospecific binding of ergot alkaloids to a sigma receptor has been observed, and, accordingly, it is feasible to consider (–)-cycloclavine as a potential lead structure for sigma receptor modulator design.

In conclusion, we have successfully completed a total synthesis of natural (+)-cycloclavine, featuring an optimization of the catalyst for the asymmetric cyclopropanation of allene with an active ester diazopropanoate, a regiospecific Pd-catalyzed ketone dehydrogenation to the enone, and two intramolecular Diels–Alder reactions for indole/indoline annulations. Furthermore, we have characterized the binding effects of (+)- and (–)-cycloclavine against 16 CNS receptors, and discovered significant stereospecificity properties. (+)-Cycloclavine has at least 10-fold higher potency at the serotonin 5-HT₁c receptor than at any of the other tested receptors, making it one of the most selective tryptamines discovered to date. Furthermore, the receptor subtype profile of (+)-cycloclavine resembles that of the clinically validated mushroom metabolite psilocin more closely than the related psychedelics LSD and DMT. Finally, we determined that the unnatural (–)-cycloclavine has considerably lower affinities at all 5-HT receptors than (+)-cycloclavine, but is quite active at the sigma-1 receptor, a property that it shares with the endogenous sigma-1 ligand DMT. We suggest that these results, in combination with the excellent synthetic tractability of the cycloclavine scaffold, encourage future research on the medicinal chemistry of clavine alkaloids.

All glassware were dried in an oven at 140 °C for at least 2 h prior to use. All air and moisture-sensitive reactions were performed under a dry N₂ atmosphere. Reactions carried out at 0 °C employed an ice bath. All air and moisture-sensitive reactions were performed under a dry N₂ atmosphere. Reactions carried out at –78 °C employed a dry ice/acetone bath. Monowave 300 reactor in glass microwave vials (cap sealed) with continuous magnetic stirring and internal ruby thermometer and/or external infrared surface temperature sensor. IR spectra were obtained at 300, 400, 500, 600, or 700 MHz NMR in CDCl₃, unless otherwise specified. ¹³C NMR spectra were obtained at 100, 125, or 150 MHz with proton-decoupling. Chemical shifts (δ) are reported in parts per million with the residual solvent peak used as an internal standard: ¹H/¹³C (Solvent) δ = 7.26/77.16 (CDCl₃); 7.16/128.06 (CD₂Cl₂), 5.32/54.00 (CDCl₃), and 2.08/20.43 (toluene-d₈) and are tabulated as follows: chemical shift, multiplicity (standard abbreviations), and coupling constant(s), number of protons. Reactions were monitored by TLC analysis (pre-coated silica gel 60 F₂₅₄ plates, 250 µm layer thickness) and visualization was accomplished with a KMnO₄ solution (1.50 g of KMnO₄, 10 g of K₂CO₃, and 1.25 mL of 10% NaOH in 200 mL of H₂O₂), when needed. Flash chromatography was performed using SiO₂ (40–63 µm). Specific rotations were measured on a polarimeter equipped with a sodium lamp. Pentfluoro-phenyl 2-diazopropanoate (4)¹¹ and stannane 34¹¹ were prepared as reported previously.

**N-Tetrabromophthaloyl-(R)-tert-leucine**

An oven-dried flask topped with a Dean–Stark apparatus and condenser was charged sequentially with (R)-tert-leucine ([R]-18; 0.500 g, 3.77 mmol), tetramethylphthalic anhydride (17; 1.79 g, 3.77 mmol), anhyd toluene (10 mL), and NEt₃ (0.0536 mL, 0.377 mmol). The resulting heterogeneous mixture was heated at reflux while the solvent was removed at a rate of ca. 1 mL/h. The solution was cooled to r.t. and treated with aq 5% HCl (6 mL) and EtOAc (15 mL). The aqueous layer was extracted with EtOAc (6 mL) and the combined organic layers were dried (Na₂SO₄) and concentrated to give N-tetramethylphthaloyl-(R)-tert-leucine as a white solid; yield: 2.11 g (97%); [α]_D²⁰ +22.3 (c = 0.40, EtOH).

¹¹H NMR (300 MHz, acetone-d₆): δ = 4.68 (s, 1 H), 1.19 (s, 9 H).

The experimental data were consistent with the literature-reported data for the enantiomer.¹⁸

**Dirhodium(II) Tetrakis[N-tetramethylphthaloyl-(R)-tert-leucinate] [Rh₂(R-TBPTTL)₄, 10]**

An oven-dried flask topped with a Dean–Stark apparatus and condenser was charged with Rh₂(OAc)₄ (0.130 g, 0.294 mmol), N-tetramethylphthaloyl-(R)-tert-leucine (0.933 g, 1.62 mmol), and a mixture of chlorobenzene and MeCN (9:1, 13 mL). The resulting dark purple denser was charged with Rh₂(OAc)₄ (0.130 g, 0.294 mmol), tetrabromophthalic anhydride (1.1 g, 3.77 mmol), tetrabromophthaloyl-[(tert-leucine)₂]-c₁₂h₁₀ (0.555 g, 75%) as a green solid, which was used directly without further purification.

¹¹H NMR (300 MHz, benzene-d₆): δ = 5.37 (s, 4 H), 1.45 (s, 36 H).

The experimental data were consistent with the literature-reported data for the enantiomer.²⁸

**4,7-Diphenyl-3a,4,7a-tetrahydroisobenzofuran-1,3-dione**

A clear, yellow solution of butadiene (20 g, 0.96 mmol) and maleic anhydride (201 (1.1 g, 11 mmol) in xylene (25 mL) was heated at 140 °C for 16 h. After this time, the solution was cooled to 0 °C and treated sequentially with toluene (40 mL) and sat. aq NaHCO₃ (40 mL). The organic layer was washed with NaHCO₃ (40 mL), brine (40 mL), filtered, and dried (Na₂SO₄) to provide 10 (0.555 g, 75%) as a white solid; yield: 2.29 g (78%).

¹¹H NMR (500 MHz, CDCl₃): δ = 7.43 (app t, J = 7.3 Hz, 4 H), 7.39–7.36 (m, 6 H), 6.56 (s, 2 H), 3.84 (d, J = 4.0 Hz, 2 H), 3.74 (dd, J = 4.5, 2.0 Hz, 2 H).

The experimental data were consistent with the literature-reported data.³⁹

**4,7-Diphenylisobenzofuran-1,3-dione (21)**

A red solution of 4,7-diphenyl-3a,4,7a-tetrahydroisobenzofuran-1,3-dione (1.0 g, 3.3 mmol) and DDQ (1.5 g, 6.6 mmol) in toluene (10 mL) was heated at 110 °C for 17 h. The reaction mixture was concentr-
trated and filtered, and the solid was washed with EtOH to provide crude 21 (0.832 g, 84%) as a light pink solid that was used without further purification.

\[^{1}H\text{NMR (300 MHz, CDCl}_3\): } \delta = 7.85 \text{ (s, 2 H), 7.61–7.57 (m, 4 H), 7.54–7.51 (m, 6 H).}

The experimental data were consistent with the literature-reported data.\(^{39}\)

(5)-2-(1,3-Dioxo-4,7-diphenylisoindolin-2-yl)-3,3-dimethylbutanoic acid (22)

An oven-dried microwave vial was charged sequentially with 4,7-diphenylbenzofuran-1,3-dione (21: 0.300 g, 1.00 mmol), (5)-tart-leucine (0.199 g, 1.50 mmol), and NEt\(_3\) (0.0283 mL, 0.200 mmol), and the resulting homogeneous brown solution was heated at reflux for 19 h. The solution was cooled to r.t., and extracted with aq 5% HCl and EtOAc. The aqueous layer was back-extracted with EtOAc (3 ×), and the resulting heterogeneous brown solution was heated at reflux for temperature, oil bath. The reaction mixture was cooled and concentrated. The resulting residue was purified by chromatography on SiO\(_2\) (0–2% acetone/CH\(_2\)Cl\(_2\)) to provide the carboxylic acid 22 as a white foam; yield: 0.128 g (31%); mp 142.7–145 °C; [\(\alpha\)]\(_{D}\)\(^{20} = \text{10.6 (c 0.17, CHCl}_3\).\]

IR (ATR): 2963, 1770, 1710, 1603, 1475, 1382, 1122, 903, 752, 698 cm\(^{-1}\).

\[^{13}C\text{NMR (100 MHz, CDCl}_3\): } \delta = 129.6, 128.9, 128.3, 128.1, 60.0, 35.8, 28.2.

\[^{1}H\text{NMR (300 MHz, CDCl}_3\): } \delta = 7.65 (s, 2 H), 7.56 (app dd, \(J = 7.6, 1.9\) Hz, 4 H), 7.52–7.45 (m, 6 H), 4.68 (s, 1 H), 1.14 (s, 9 H).

Asymmetric Cyclopropanation of Allenes; General Procedure (Table 1)

Pentafluorophenyl (R)-1-Methyl-2-methylene cyclopropane-1-carboxylate ([R]-24)\(^{11}\)

An oven-dried three-necked flask was charged with the respective catalyst (0.0056 mmol) and hexanes or CH\(_2\)Cl\(_2\) (2.8 mL), and the resulting green solution was cooled to \(-78^\circ\text{C}\) and treated dropwise with an excess of condensed gaseous allene (23: ca. 14 mmol). The reaction mixture was then treated with a solution of pentafluorophenyl 2-diazopropanoate (4: 0.56 mmol) in hexanes or CH\(_2\)Cl\(_2\) (1.7 mL) via syringe pump at a rate of 1 mL/h. After the addition was complete, the mixture was allowed to warm to r.t. The solution was concentrated under reduced pressure and the resulting residue was purified by chromatography on SiO\(_2\) (0–2% EtO\(_2\) /hexanes) to provide pentafluorophenyl cyclopropane (R)-24.

\[^{1}H\text{NMR (400 MHz, CDCl}_3\): } \delta = 5.64 (t, \(J = 2.8\) Hz, 1 H), 5.56 (t, \(J = 2.2\) Hz, 1 H), 2.30 (dt, \(J = 9.2, 2.5\) Hz, 1 H), 1.63 (dt, \(J = 9.2, 2.4\) Hz, 1 H), 1.51 (s, 3 H).

(\(R\),\(E\))-N1-Methyl-2-methylene-N\(\_\) (3-oxobut-1-en-1-yl)cyclopropane-1-carboxamide ([\(R\)-\(E\)]-27)\(^{11}\)

An oven-dried flask charged with vinylogous amide 26 (0.0178 g, 0.18 mmol, 1.0 equiv) was evacuated and backfilled with N\(_2\) (3 ×). Freshly distilled THF (0.6 mL) was added and the resulting solution was cooled to \(-78^\circ\text{C}\) and treated dropwise with n-BuLi (2.29 M solution in hexanes, 0.082 mL, 0.19 mmol, 1.05 equiv). The resulting clear, pale yellow solution was stirred for 5 min at \(-78^\circ\text{C}\), then treated with a solution of ester (R)-24 (0.053 g, 0.19 mmol, 1.05 equiv) in THF (1 mL). The resulting bright yellow solution was treated with DMAP (0.023 g, 0.19 mmol, 0.1 equiv) and stirred for 10 min at \(-78^\circ\text{C}\). The cold bath was removed and the reaction mixture was allowed to warm to r.t. The reaction was quenched with sat. aq NaHCO\(_3\). After addition of EtOAc, the aqueous layer was extracted with EtOAc (3 ×) and the combined organic layers were dried (Na\(_2\)SO\(_4\)), filtered, and concentrated. The crude product was purified by chromatography on SiO\(_2\) (10–15% EtO\(_2\)/hexanes) to provide vinylogous imide (R)-27 as a clear, pale yellow oil; yield: 28 mg (80%).

\[^{1}H\text{NMR (300 MHz, CDCl}_3\): } \delta = 8.36 (d, \(J = 13.8\) Hz, 1 H), 5.71 (d, \(J = 13.8\) Hz, 1 H), 5.69 (app d, \(J = 3.0\) Hz, 1 H), 5.53 (app s, 1 H), 3.15 (s, 3 H), 2.29 (s, 3 H), 1.79 (dt, \(J = 9.6, 2.2\) Hz, 1 H), 1.49 (s, 3 H), 1.32 (dt, \(J = 9.6, 2.2\) Hz, 1 H).

SFC conditions for r.r. analysis: Chiralpak-IC semi-prep column (250 × 10 mm), gradient elution: 5-15% i-ProH, 7 mL/min, 254 nm detection, P = 100 bar.

Pentafluorophenyl (S)-1-Methyl-2-methylene cyclopropane-1-carboxylate ([S]-24) (Table 1, entry 3)

An oven-dried three-necked flask fitted with a N\(_2\) inlet, a dry-ice condenser, and a septum was charged with Rh\(_2\)(R-TPFPTTL)\(_4\) (10; 0.566 g, 0.226 mmol) and CH\(_2\)Cl\(_2\) (1.41 mL), and the green solution was cooled to \(-78^\circ\text{C}\) (dry ice/acetone bath) and treated dropwise with an excess of condensed gaseous allene (23: ca. 65 drops, 5.14 g, 5.52 mmol). The resulting solution was then treated with a solution of pentafluorophenyl 2-diazopropanoate (4: 6.00 g, 22.6 mmol) in CH\(_2\)Cl\(_2\) (12 mL) via syringe pump at a rate of ca. 2 mL/h. The reaction mixture was allowed to stir at \(-78^\circ\text{C}\) for 1 h before warming to r.t. The resulting green residue was purified by chromatography on SiO\(_2\) (0–1% EtO\(_2\)/hexanes) to afford methylene cyclopropane (S)-24 as a clear and pale yellow oil; yield: 5.41 g (86%); [\(\alpha\)]\(_{D}\)\(^{22} = 2.8 (c 7.6, CHCl\(_3\)).\]
(S,E)-N,1-Dimethyl-2-methylene-N-(3-oxobut-1-en-1-yl)cyclopropane-1-carboxamide [(S)-27]

An oven-dried round bottomed flask was charged with vinylogous amide 26 (2.77 g, 27.9 mmol) and evacuated and backfilled with N₂ (3 ×). Distilled THF (93 mL) was added and the resulting solution was cooled to −78 °C and treated dropwise with n-Buli (11.7 mL, 2.5 M solution in hexanes, 29.3 mmol). The mixture was stirred for 5 min at −78 °C, then treated dropwise with a solution of pentfluorophenyl (S)-1-methyl-2-methylene-cyclopropane-1-carboxylate [(S)-24] (214.5 g, 9.3 mmol) in THF (15 mL). The reaction mixture was treated with DMAP (0.0341 g, 0.279 mmol) and stirred for 10 min at −78 °C. The cold bath was removed and the solution was allowed to warm to r.t., and quenched with sat. aq NaHCO₃ and EtOAc. The aqueous layer was extracted with EtOAc (3 ×) and the combined organic layers were dried (Na₂SO₄) and concentrated. The crude product was purified by chromatography on SiO₂ (10−15% EtOAc/hexanes) to afford [(S)-27]; yield: 4.30 g (80%); e.r. 87:13 by SFC analysis) as a clear and pale yellow oil; [α]D⁰ 26 −243.2 (c 0.93 CHCl₃).

1H NMR (400 MHz, CDCl₃): δ = 3.41 (dd, J = 12.8, 4.0, 1.0 Hz, 1 H), 2.80 (dd, J = 13.6, 4.0, 1.6 Hz, 1 H), 2.69 (ddt, J = 15.5, 4.8, 1.0 Hz, 1 H), 2.66 (s, 3 H), 2.54 (dd, J = 15.4, 12.2, 7.2 Hz, 1 H), 2.35 (app t, J = 13.4 Hz, 1 H), 2.13 (ddt, J = 13.6, 5.0, 1.4 Hz, 1 H), 1.72 (ddd, J = 13.1, 7.3, 1.5 Hz, 1 H), 1.38 (s, 3 H), 0.95 (d, J = 3.6 Hz, 1 H), 0.66 (d, J = 4.0 Hz, 1 H).

HRMS (ESI+): m/z calcd for C₁₁H₁₆NO₂ (M + H)+: 194.1176; found: 194.1179.

All other experimental data were consistent with the reported data for the enantiomer.11

(1aS,3aR,7aR)-1a,3-Dimethylhexahydro-2H-cyclopenta[jindole-2,5(3H)-dione (30) and (1aS,3aS,7aR)-1a,3-Dimethylhexahydro-2H-cyclopenta[jindole-2,5(3H)-dione (29)

An oven-dried flask was charged with NaHMDS (2.50 g, 12.9 mmol) and anhyd THF (150 mL) under an atmosphere of N₂. The resulting clear and colorless solution was stirred for 15 min at r.t., then cooled to −78 °C, and stirred for a further 15 min at this temperature prior to treatment with a solution of 27 (2.75 g, 14.2 mmol) in THF (20 mL) slowly at a rate of 4 mL/h, maintaining the temperature of the aceto/dry ice bath below −50 °C. During the slow addition of the amide, the solution changed color from clear and pale yellow to clear and orange. The resulting clear, orange solution was allowed to warm to −50 °C over 1 h, cooled to −78 °C and treated dropwise with a solution of TBSCI (2.36 g, 15.5 mmol) in THF (52 mL). The reaction mixture was allowed to stir for a further 5 min at −78 °C before the cold bath was removed, and the solution allowed to reach r.t.

1H NMR analysis of an aliquot (CDCl₃) indicated conversion into enol ether 28.

1H NMR (300 MHz, CDCl₃): δ = 7.65 (d, J = 13.5 Hz, 1 H), 5.67 (app s, 1 H), 5.52 (d, J = 13.8 Hz, 1 H), 5.45 (app s, 1 H), 4.23 (s, 2 H), 3.13 (br s, 3 H), 1.73 (dt, J = 9.6, 2.1 Hz, 1 H), 1.47 (s, 3 H), 1.25 (dt, J = 9.4, 2.2 Hz, 1 H), 1.00 (s, 3 H), 0.25 (s, 3 H), 0.24 (s, 3 H).

The mixture was concentrated and directly subjected without purification to the next reaction. A solution of crude amide 28 (assumed 3.98 g, 12.9 mmol) in THF (28 mL, 3.5 mL × 8 batches) was heated under microwave irradiation at 95 °C for 30 min, then concentrated to deliver the crude Diels–Alder adducts (assumed 3.98 g, 12.9 mmol) as an orange oil that was used directly in the next conversion. The crude enol ether adducts (assumed 3.98 g, 12.9 mmol) were immediately dissolved in THF (130 mL), treated dropwise with TBAF (12.9 mL, 1 M solution in THF, 12.9 mmol) and stirred at r.t. for 30 min. The reaction mixture was filtered through a pad of Florisil (washed with EtOAc) and concentrated. The resulting residue was purified by chromatography on SiO₂ (20% EtOAc/hexanes to elute recovered starting material, then 50−70% EtOAc/hexanes to elute the truns-diastereomer 30, then 90−100% EtOAc/hexanes to elute the cis-diastereomer 29) to provide trans-ketone 30 as a pale yellow solid (1.64 g, 66%) and cis-ketone 29 as a yellow oil (0.371 g, 15%).

HRMS (ESI+): m/z calcd for C₁₁H₁₆NO₂ (M + H)+: 194.1176; found: 194.1179.

A solution of Pd(TFA)₂ (0.500 g, 2.59 mmol) was added and the reaction mixture was allowed to stir at 55 °C for 4 days. Additional Pd(TFA)₂ (2 × 0.11 g) was added after 24 and 48 h to drive the reaction to completion. The mixture was removed via pipette transfer and the white needle-shaped crystals were cooled to r.t., and kept overnight at −20 °C. The mother liquor was re-measured, and the third recrystallization led to 98:2 e.r.

Enantiomeric Enrichment by Recrystallization of Enone 31; Typical Procedure

Enone 31 (0.110 g, 0.573 mmol) was dissolved in boiling MTBE (9.5 mL) and 1,2-dichloroethane (0.4 mL) and the solution was allowed to cool to r.t., and kept overnight at −20 °C. The mother liquor was removed via pipette transfer and the white needle-shaped crystals were washed with MTBE (2 ×) and placed under vacuum to remove trace solvents. The first recrystallization provided compound of >99.5:0.5 e.r. by chiral SFC analysis; a 95:4:6 e.r. was achieved in the second recrystallization, and the third recrystallization led to 98:2 e.r. to deliver a combined yield of enone 31 (0.0654 g, 60%, 69% of theoretical maximum) as a white crystalline solid. Only combined samples with e.r. >97:2:5:2 were carried on in the subsequent reaction; [α]D⁰ 27 + 111.1 (c 0.484, CHCl₃).

Enantiomeric Enrichment by Recrystallization of Enone 31; Typical Procedure

Enone 31 (0.110 g, 0.573 mmol) was dissolved in boiling MTBE (9.5 mL) and 1,2-dichloroethane (0.4 mL) and the solution was allowed to cool to r.t., and kept overnight at −20 °C. The mother liquor was removed via pipette transfer and the white needle-shaped crystals were washed with MTBE (2 ×) and placed under vacuum to remove trace solvents. The first recrystallization provided compound of >99.5:0.5 e.r. by chiral SFC analysis; a 95:4:6 e.r. was achieved in the second recrystallization, and the third recrystallization led to 98:2 e.r. to deliver a combined yield of enone 31 (0.0654 g, 60%, 69% of theoretical maximum) as a white crystalline solid. Only combined samples with e.r. >97:2:5:2 were carried on in the subsequent reaction; [α]D⁰ 27 + 111.1 (c 0.484, CHCl₃).
An oven-dried 3-necked 25 mL flask fitted with two stoppers and N₂ was maintained between –67.4 and –74.5 °C, then cooled to –93.1 °C. EtOAc and quenched with aq 2 M NH₄Cl, maintaining the internal temperature at –35 °C. The crude residue was used directly in the next step.¹¹

**SFC analysis:** Chiralpak-IC semiprep column (250 × 10 mm), gradient elution: 1–30% i-PrOH, 5.5 mL/min, 254 nm detection, P = 100 bar.

A microwave flask was charged with Pd(TFA)₂ (0.0092 g, 0.027 mmol). A solution of Na-metal (0.250 g, 41%) and axial alcohol (0.250 g, 1.31 mmol) in anhyd THF/Et₂O 1:1 (3.5 mL) slowly over 10 min. The reaction mixture was concentrated and purified by chromatography on SiO₂ (50% EtOAc/hexanes) to deliver vinylogous amide as a white solid; yield: 0.029 g (56%).

IR (ATR): 2956, 1732, 1607, 1458, 1241, 1074 cm⁻¹.

²¹H NMR (300 MHz, CDCl₃): δ = 5.49 (s, 1 H), 2.93 (s, 3 H), 2.68–2.65 (m, 1 H), 2.63 (dd, J = 5.0 Hz, 1 H), 2.35 (td, J = 12.4, 6.8 Hz, 1 H), 1.73 (ddd, J = 13.0, 4.7, 2.8 Hz, 1 H), 1.41 (s, 3 H), 1.33 (dd, J = 4.2 Hz, 1 H), 1.17 (d, J = 4.2 Hz, 1 H).

²¹³C NMR (100 MHz, CDCl₃): δ = 197.2, 177.1, 164.4, 101.4, 36.9, 30.1, 29.9, 29.2, 26.2, 24.5, 10.4.

HRMS (LCMS ESI+): m/z calcd for C₁₁H₁₈N₂O₂+ (M + H⁺): 212.1019; found: 212.1020.

All data were consistent with the literature-reported data.²

**Allylic Alcohols α-35 and β-35**

An oven-dried 3-necked 25 mL flask fitted with two stoppers and N₂ inlet was charged with stannane (0.893 g, 1.57 mmol) and evacuated under high vacuum, then backfilled with N₂ (3 × 1). A stopper was exchanged for an internal thermocouple thermometer and anhyd Et₂O (5 mL) was added. The clear, pale yellow solution was cooled to –70.5 °C (Et₂O/dry ice) and stirred for 10 min, then treated dropwise with n-BuLi (0.628 ml, 2.5 M solution in hexanes, 1.57 mmol), during which time the temperature rose to –67.4 °C. The resulting clear, yellow solution was stirred for 15 min while the internal temperature was maintained between –67.4 and –74.5 °C, then cooled to –93.1 °C using a liquid N₂:Et₂O bath, and treated with a solution of enone (0.250 g, 1.31 mmol) in anhyd THF/CH₂Cl₂ 1:1 (3.5 mL) slowly over 10 min in 0.4 mL portions. The temperature rose to –86 °C during the addition, and the reaction mixture was subsequently stirred for 30 min below –90 °C, then for 1 h at –76 °C. The solution was diluted with EtOAc and quenched with aq 2 M NH₄Cl, maintaining the internal temperature below –50 °C. The aqueous layer was extracted with EtOAc (5 × 1), and the combined organic layers were dried (Na₂SO₄), and concentrated. The residue was purified by chromatography on SiO₂ (gradient elution 5% acetone/CH₂Cl₂ to elute recovered starting material enone (3 mg); then alcohol α-35; then 10% acetone/CH₂Cl₂ was used to elute alcohol β-35) to deliver equatorial alcohol α-35 (0.250 g, 41%) and axial alcohol β-35 (0.196 g, 32%) as white foams.

α-35

| [α]D25²⁰ | +30.1 (c 0.2, CHCl₃). |

β-35

| [α]D25²⁰ | +2.0 (c 0.3, CHCl₃). |

A solution of α-35 (0.128 g, 0.272 mmol) in anhyd degassed toluene (10 mL) was heated to 135 °C in a sealed tube for 90 h. The reaction mixture was concentrated and filtered through a pad of SiO₂, washing with 2–5% acetone/CH₂Cl₂ to provide crude (26 mg, 39%), which was used directly in the next step.

¹¹H NMR (400 MHz, CDCl₃): δ = 8.14 (br s, 1 H), 7.21 (d, J = 8.0 Hz, 1 H), 7.09 (dd, J = 8.0, 7.2 Hz, 1 H), 7.02 (app s, 1 H), 6.80 (d, J = 6.8 Hz, 1 H), 3.78 (dd, J = 11.8, 4.2 Hz, 1 H), 3.29 (dd, J = 13.6, 4.0 Hz, 1 H), 2.79 (s, 3 H), 2.71–2.64 (m, 1 H), 1.80 (s, 3 H), 1.14 (d, J = 3.6 Hz, 1 H), 0.84 (d, J = 3.2 Hz, 1 H).

A microwave vial was charged with a solution of crude (0.0270 g, 0.107 mmol) in anhyd THF (1.2 mL) under an atmosphere of N₂. The solution was cooled to 0 °C and treated dropwise with LiAlH₄ (0.535 mL, 1 M solution in Et₂O, 0.535 mmol). The reaction mixture was stirred at reflux in a sealed tube for 16 h, diluted with Et₂O, cooled to 0 °C and treated sequentially with H₂O (0.020 mL), aq 15% NaOH (0.020 mL) and H₂O (0.061 mL), warmed to r.t. and stirred for 15 min. Anhyd MgSO₄ was then added and the solution was stirred rigorously for 15 min, and filtered through a pad of Celite. The crude residue was purified by chromatography on SiO₂ (pre-washed column with 0.1% NEt₃/CH₂Cl₂, then gradient elution 0–2% MeOH/CH₂Cl₂ with 0.1% NEt₃ added to each eluent) to deliver a (−)-cycloclavine as a white solid; yield: 22.0 mg (86%, or 34% over 2 steps); mp 161.1–163.8 °C (dec.; [α]D20²⁰ +61.4 (c 0.2, CHCl₃); [lit. ⁴ [α]D20²⁰ +63 (c 1, CHCl₃)].

IR (ATR): 3409, 3166, 3103, 3062, 2942, 2885, 2843, 2788, 1617, 1590, 1442, 1222, 1165, 1095, 922, 749 cm⁻¹.

¹¹H NMR (500 MHz, CDCl₃): δ = 7.89 (br s, 1 H), 7.13 (dd, J = 8.0, 0.5 Hz, 1 H), 7.09 (app t, J = 7.8 Hz, 1 H), 6.90 (app t, J = 1.8 Hz, 1 H), 6.82 (dd, J = 7.0, 0.5 Hz, 1 H), 3.15 (d, J = 9.0 Hz, 1 H), 3.13 (dd, J = 13.8, 3.8 Hz, 1 H), 2.78 (dd, J = 11.5, 4.0 Hz, 1 H), 2.63–2.57 (m, 1 H), 2.40 (d, J = 8.5 Hz, 1 H), 2.36 (s, 3 H), 1.69 (s, 3 H), 1.60 (d, J = 3.5 Hz, 1 H), 0.45 (d, J = 3.5 Hz, 1 H).

¹³C NMR (150 MHz, CDCl₃): δ = 135.6, 133.7, 128.8, 123.1, 118.2, 113.5, 110.5, 108.1, 69.8, 65.7, 40.1, 34.5, 27.9, 25.1, 24.4, 16.6.
DEPT-135 (100 MHz, CDCl₃): δ = 123.1 (CH), 118.2 (CH), 110.5 (CH), 108.1 (CH), 69.8 (CH), 65.8 (CH₂), 40.1 (NCH₃), 25.1 (CH₂), 24.4 (CH₂), 16.6 (CH₃).

HRMS (LCMS ESI+): m/z calcd for C₁₆H₁₉N₂ (M + H)+: 239.1543; found: 239.1544.

All relevant data were consistent with the literature-reported data for the natural product⁴ and the enantiomer.¹¹

Funding Information

The authors are grateful to Boehringer-Ingelheim Pharmaceuticals Inc., Ridgefield CT, for partial financial support of this work. SRM also acknowledges support from the Mary E. Warga and the University of Pittsburgh Arts and Sciences Mellon Fellowships.

Acknowledgment

The authors thank M. K. Wipf for graphical support.

Supporting Information

Supporting information for this article is available online at https://doi.org/10.1055/s-0037-1610395. Spectral data (¹H and ¹³C NMR for selected new compounds and (+)-cycloclavine).

References


