Efficient Synthesis of Pyrazinoic Acid Hybrid Conjugates

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Abstract Benzotriazole-activated pyrazinoic acid was utilized as a versatile building block for the efficient and convenient synthesis of novel hybrid conjugates of pyrazinoic acid with secondary amines via amino acid linkers in high yields.

Key words pyrazinamide, pyrazinoic acid, secondary amines, amino acids, benzotriazole methodology, antituberculosis

Tuberculosis (TB) is a bacterial pathogen caused by Mycobacterium tuberculosis, which is known to cause pulmonary infection and to become extremely pervasive within the lungs. 1–3 TB is considered to be one of the world’s deadliest communicable diseases because of its high virulence and the ability of M. tuberculosis to enter into a dormant state, then subsequently undergo reactivation. 3–5 Pyrazinamide (PZA) is a first-line antituberculosis prodrug that is often used in combinational therapy with drugs such as isoniazide, ethambutol, streptomycin, and rifampicin (Figure 1). 6–8

PZA is perceived to inhibit vital ribosomal proteins after being converted into its active constituent, pyrazinoic acid (POA), by the tuberculosis enzyme, pyrazinamidase (PZAAse) (Scheme 1). 9 It may lower the pH of the area surrounding M. tuberculosis to such an extent that the organism is unable to grow. Due to its low lipophilicity, POA cannot be absorbed by the gastrointestinal tract. Fortunately, the drug can be absorbed in the pyrazinamide configuration.

Figure 1 Current antituberculosis drugs

Figure 2 Synthesis of pyrazinoic acid hybrid conjugates.
One of the drawbacks of using PZA to treat TB is that it inhibits protein synthesis. With prolonged administration of the recommended dose, harmful side effects such as hepatitis, acute hypertension, thrombocytopenia, and gastrointestinal discomfort have been reported. To overcome these issues, several molecular hybridization approaches have been reported for the development of potential antitubercular agents. Most hybridized structures include clinically used drugs such as rifamycin, ethambutol and isoniazid coupled with other hydrophobic structures such as cinamic acid derivatives. Unfortunately the most promising prodrugs of PZA are not stable.

Secondary amines such morpholine, piperidine, N-methylpiperazine, and pyrrolidine are important scaffolds for potential biological molecules. Structural activity relationships (SARs) suggest that these secondary amines play an important role in bioactive molecules. Drug–amino acid conjugates are used to enhance drug delivery and to increase tissue cell penetration. In addition to acting as carriers of these agents, amino acids also amplify their bioavailability while maintaining their bioactive integrity. Recently, we have synthesized and reported several bioconjugates with enhanced biological properties and increased lipophilicity.

In a continuation of our interest in synthesizing amino acid–peptide conjugates with biological significance, we report herein an efficient synthesis of pyrazimide hybrid conjugates, which contain various amino acids and secondary amines, that may decrease the dose of PZA required to fight TB, increase the drug’s lipophilicity and decrease adverse effects.

POA was activated as its benzotriazolide derivative \( \text{POA} \) by following the previously reported procedure and coupled with free amino acids in a mixture of acetonitrile and water (7:3) at 20 °C for 2 h in the presence of 1.5 equivalents of triethylamine to give compounds. Attempts to prepare benzotriazole derivatives of POA–amino acid conjugates failed. We were also unsuccessful in coupling compounds with secondary amines using different coupling reagents and ended up with mixtures of compounds as evidenced by TLC (Scheme 2). We therefore decided to redesign our approach and synthesize bis-conjugates of POA–secondary amines by coupling Boc-protected aminoacylbenzotriazoles with secondary amines. After removing the Boc group with dioxane–HCl, the unprotected conjugates were coupled with benzotriazole–activated POA to produce the desired products in good yields (Scheme 3). Boc-protected aminoacylbenzotriazoles were treated with secondary amines in the presence of triethylamine in acetonitrile at 20 °C for 2 h to obtain the conjugates in good yields without loss of chiral integrity (Scheme 3, Table 1).

Boc-protected amino acid–secondary amine conjugates were deprotected using a dioxane–HCl mixture at 20 °C for 1 h to give unprotected amino acid–secondary amine conjugates. These compounds were then used in the next step without further characterization. The target compounds were prepared by coupling the unprotected amino acid–secondary amine conjugates with POA-benzotriazole in the presence of triethylamine in acetonitrile at 20 °C for 3 h (Scheme 4, Table 2). All compounds were fully characterized by NMR spectroscopy, HRMS and specific rotation. X-ray diffraction analysis of compound further confirmed the formation of the hybrid conjugate (Figure 2).

In conclusion, benzotriazole–activated pyrazinamidic acid has been used as a precursor for the efficient synthesis of pyrazinamidic acid hybrid conjugates. The hybrid conjugates may be candidates for the development of new antituberculosis agents.

### Table 1 Boc-Protected Amino Acid–Secondary Amine Conjugates \( 11a–t \)

<table>
<thead>
<tr>
<th>Entry</th>
<th>Product</th>
<th>Yield (%)</th>
<th>Mp (°C)</th>
<th>([\alpha]_D^{20}) (c 1.0 in MeOH)</th>
</tr>
</thead>
<tbody>
<tr>
<td>1</td>
<td>Boc-Gly-Mor</td>
<td>11a</td>
<td>62</td>
<td>114–116</td>
</tr>
<tr>
<td>2</td>
<td>Boc-l-Ala-Mor</td>
<td>11b</td>
<td>67</td>
<td>oil</td>
</tr>
<tr>
<td>3</td>
<td>Boc-3-3-Ala-Mor</td>
<td>11b</td>
<td>74</td>
<td>oil</td>
</tr>
<tr>
<td>4</td>
<td>Boc-l-Val-Mor</td>
<td>11c</td>
<td>75</td>
<td>135–137</td>
</tr>
<tr>
<td>5</td>
<td>Boc-l-Ile-Mor</td>
<td>11d</td>
<td>81</td>
<td>oil</td>
</tr>
<tr>
<td>6</td>
<td>Boc-l-Phe-Mor</td>
<td>11e</td>
<td>84</td>
<td>129–131</td>
</tr>
<tr>
<td>7</td>
<td>Boc-Gly-Pip</td>
<td>11f</td>
<td>90</td>
<td>oil</td>
</tr>
<tr>
<td>8</td>
<td>Boc-l-Ala-Pip</td>
<td>11g</td>
<td>97</td>
<td>oil</td>
</tr>
<tr>
<td>9</td>
<td>Boc-l-Val-Pip</td>
<td>11h</td>
<td>78</td>
<td>oil</td>
</tr>
<tr>
<td>10</td>
<td>Boc-l-Ile-Pip</td>
<td>11i</td>
<td>79</td>
<td>oil</td>
</tr>
<tr>
<td>11</td>
<td>Boc-l-Phe-Pip</td>
<td>11j</td>
<td>81</td>
<td>122–124</td>
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<tr>
<td>12</td>
<td>Boc-Gly-NMP</td>
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<td>11l</td>
<td>81</td>
<td>oil</td>
</tr>
<tr>
<td>14</td>
<td>Boc-l-Val-NMP</td>
<td>11m</td>
<td>84</td>
<td>oil</td>
</tr>
<tr>
<td>15</td>
<td>Boc-l-Ile-NMP</td>
<td>11n</td>
<td>69</td>
<td>oil</td>
</tr>
<tr>
<td>16</td>
<td>Boc-l-Phe-NMP</td>
<td>11o</td>
<td>83</td>
<td>111–113</td>
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<tr>
<td>17</td>
<td>Boc-Gly-Pyr</td>
<td>11p</td>
<td>89</td>
<td>oil</td>
</tr>
<tr>
<td>18</td>
<td>Boc-l-Ala-Pyr</td>
<td>11q</td>
<td>83</td>
<td>oil</td>
</tr>
<tr>
<td>19</td>
<td>Boc-l-Val-Pyr</td>
<td>11r</td>
<td>67</td>
<td>oil</td>
</tr>
<tr>
<td>20</td>
<td>Boc-l-Ile-Pyr</td>
<td>11s</td>
<td>72</td>
<td>oil</td>
</tr>
<tr>
<td>21</td>
<td>Boc-l-Phe-Pyr</td>
<td>11t</td>
<td>78</td>
<td>126–128</td>
</tr>
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</table>
Melting points were determined with a capillary melting-point apparatus equipped with a digital thermometer. Reactions were monitored by using thin-layer chromatography (TLC) on 0.2 mm silica gel F254 plates (Merck). The chemical structures of final products and intermediates were characterized by 1H and 13C NMR spectroscopy with a Bruker NMR spectrometer (500 MHz, 125 MHz). 13C NMR spectra were fully decoupled. Chemical shifts are reported in parts per million (ppm) using the deuterated solvent peak or tetramethylsilane as an internal standard. Mass spectrometric analysis was carried out with a high-resolution Biosystems QStar Elite time-of-flight electrospray mass spectrometer or an Agilent 6210 instrument using time-
Synthesis of (1- of-flight mass spectrometry (TOF-MS) with electrospray ionization (ESI) and dried over anhydrous magnesium sulfate. After filtration, the solution, 20% aqueous sodium bicarbonate was added and the organic layer mixture was stirred for 2–3 h at r.t. Upon completion of the reaction, the acetonitrile was evaporated under reduced pressure and the residue was extracted with EtOAc. The organic layer was washed with aqueous sodium carbonate and dried over sodium sulfate. After filtration, the EtOAc was evaporated under reduced pressure to obtain the desired amino acid–secondary amine conjugate in good yield.

tert-Butyl (2-Morpholinol-2-oxoethoxy) carbamate (Boc-Gly-Mor, 11a)

Yield: 62%; colorless microcrystals; mp 114–116 °C.

1H NMR (500 MHz, CDCl3): δ = 5.50 (br s, 1 H), 3.95 (d, J=3.5 Hz, 2 H), 3.51–3.62 (m, 6 H), 3.40 (t, J=4.4 Hz, 2 H), 1.45 (s, 9 H).

13C NMR (125 MHz, CDCl3): δ = 167.5, 156.0, 80.0, 66.9, 66.5, 45.0, 42.4, 42.3, 28.5.

HRMS (ESI): m/z[M + H]+ calcd for C11H20N2O4: 244.1424; found: 244.1425.

tert-Butyl (5)-(1-Morpholinol-1-oxopropan-2-yl) carbamate (Boc-t-Ala-Mor, 11b)

Yield: 67%; oil.

1H NMR (500 MHz, CDCl3): δ = 5.51 (br s, 1 H), 4.59 (br s, 1 H), 3.67–3.47 (m, 8 H), 1.43 (s, 9 H), 1.29 (d, J=5.8 Hz, 3 H).

13C NMR (125 MHz, CDCl3): δ = 171.5, 155.3, 79.8, 67.0, 66.8, 46.1, 42.6, 28.6, 19.5.

HRMS (ESI): m/z[M + H]+ calcd for C12H21N2O4: 258.1584; found: 258.1580.

tert-Butyl (1-Morpholinol-1-oxopropan-2-yl) carbamate (Boc-t-Ala-Mor, 11b’)

Yield: 74%; oil.

1H NMR (500 MHz, CDCl3): δ = 5.55 (d, J=5.8 Hz, 1 H), 4.54 (d, J=5.8 Hz, 1 H), 3.62–3.40 (m, 8 H), 1.37 (s, 9 H), 1.24 (d, J=7.9 Hz, 3 H).

13C NMR (125 MHz, CDCl3): δ = 171.4, 155.1, 79.6, 66.8, 66.6, 46.0, 42.4, 28.4, 19.2.

HRMS (ESI): m/z[M + H]+ calcd for C12H21N2O4: 258.1584; found: 258.1582.

tert-Butyl (5)-(3-Methyl-1-morpholinol-1-oxobutan-2-yl) carbamate (Boc-t-Val-Mor, 11c)

Yield: 75%; colorless microcrystals; mp 135–137 °C.

1H NMR (500 MHz, CDCl3): δ = 3.57 (d, J=7.9 Hz, 1 H), 4.34 (t, J=7.9 Hz, 1 H), 3.64–3.48 (m, 8 H), 1.89–1.83 (m, 1 H), 1.36 (s, 9 H), 0.88 (d, J=6.6 Hz, 3 H), 0.83 (d, J=6.6 Hz, 3 H).

13C NMR (125 MHz, CDCl3): δ = 170.8, 155.9, 79.4, 66.9, 66.7, 54.6, 46.3, 42.4, 31.4, 28.3, 19.6, 17.3.


tert-Butyl ((2S,3S)-3-Methyl-1-morpholinol-1-oxopentan-2-yl) carbamate (Boc-t-Ile-Mor, 11d)

Yield: 81%; oil.

1H NMR (500 MHz, CDCl3): δ = 3.46 (d, J=7.9 Hz, 1 H), 4.39 (t, J=7.9 Hz, 1 H), 3.68–3.48 (m, 8 H), 2.30–2.23 (m, 1 H), 1.65–1.64 (m, 1 H), 1.50–1.45 (m, 1 H), 1.38 (s, 9 H), 0.88–0.81 (m, 6 H).

13C NMR (125 MHz, CDCl3): δ = 171.1, 155.9, 79.6, 66.9, 66.8, 54.0, 46.5, 42.5, 38.1, 28.4, 24.1, 15.9, 11.4.

HRMS (ESI): m/z[M + H]+ calcd for C12H21N2O4: 300.2049; found: 300.2054.

### Table 2 POA Hybrid Conjugates 5a–t

<table>
<thead>
<tr>
<th>Entry</th>
<th>Product 5</th>
<th>Yield (%)</th>
<th>Mp (°C)</th>
<th>[M+H]+ (c 1.0 in MeOH)</th>
</tr>
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<tbody>
<tr>
<td>1</td>
<td>POA-Gly-Mor</td>
<td>5a</td>
<td>75</td>
<td>180–181 –</td>
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<tr>
<td>2</td>
<td>POA-t-Ala-Mor</td>
<td>5b</td>
<td>69</td>
<td>sticky –25.4</td>
</tr>
<tr>
<td>3</td>
<td>POA-ε-Ala-Mor</td>
<td>5b’</td>
<td>72</td>
<td>oil racemic</td>
</tr>
<tr>
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<td>POA-ε-Val-Mor</td>
<td>5c</td>
<td>76</td>
<td>–19.2</td>
</tr>
<tr>
<td>5</td>
<td>POA-ε-Ile-Mor</td>
<td>5d</td>
<td>70</td>
<td>–20.3</td>
</tr>
<tr>
<td>6</td>
<td>POA-ε-Phe-Mor</td>
<td>5e</td>
<td>78</td>
<td>–25.6</td>
</tr>
<tr>
<td>7</td>
<td>POA-Gly-Pip</td>
<td>5f</td>
<td>74</td>
<td>123–125 –</td>
</tr>
<tr>
<td>8</td>
<td>POA-ε-Ala-Pip</td>
<td>5g</td>
<td>75</td>
<td>–19.9</td>
</tr>
<tr>
<td>9</td>
<td>POA-ε-Phe-Pip</td>
<td>5h</td>
<td>81</td>
<td>–23.6</td>
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<tr>
<td>10</td>
<td>POA-ε-Phe-Pip</td>
<td>5i</td>
<td>64</td>
<td>–22.0</td>
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<tr>
<td>11</td>
<td>POA-ε-Phe-Pip</td>
<td>5j</td>
<td>79</td>
<td>–20.1</td>
</tr>
<tr>
<td>12</td>
<td>POA-ε-Phe-Pip</td>
<td>5k</td>
<td>69</td>
<td>sticky –</td>
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<tr>
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<td>POA-ε-Ala-NMP</td>
<td>5l</td>
<td>73</td>
<td>–26.5</td>
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<tr>
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<td>POA-ε-Val-NMP</td>
<td>5m</td>
<td>62</td>
<td>–20.9</td>
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<tr>
<td>15</td>
<td>POA-ε-Ile-NMP</td>
<td>5n</td>
<td>64</td>
<td>–23.0</td>
</tr>
<tr>
<td>16</td>
<td>POA-ε-Phe-NMP</td>
<td>5o</td>
<td>71</td>
<td>–28.1</td>
</tr>
<tr>
<td>17</td>
<td>POA-Gly-Pyr</td>
<td>5p</td>
<td>81</td>
<td>148–150 –</td>
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<td>18</td>
<td>POA-ε-Val-Pyr</td>
<td>5q</td>
<td>79</td>
<td>–23.0</td>
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<tr>
<td>19</td>
<td>POA-ε-Phe-Pyr</td>
<td>5r</td>
<td>82</td>
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<tr>
<td>20</td>
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<td>74</td>
<td>–26.6</td>
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<tr>
<td>21</td>
<td>POA-ε-Phe-Pyr</td>
<td>5t</td>
<td>73</td>
<td>–25.1</td>
</tr>
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**tert-Butyl (5)-(1-Morpholino-1-oxo-3-phenylpropan-2-yl)carbamate (Boc-i-Ph-Mor, 11e)**

Yield: 84%; colorless microcrystals; mp 129–131 °C.

1H NMR (500 MHz, CDCl3): δ = 7.29–7.19 (m, 5 H), 5.44 (d, J=7.5 Hz, 1 H), 4.82–4.78 (m, 1 H), 3.62–3.28 (m, 6 H), 3.05–2.89 (m, 4 H), 1.43 (s, 9 H).

13C NMR (125 MHz, CDCl3): δ = 170.5, 155.2, 136.5, 129.7, 128.8, 127.3, 80.0, 66.7, 66.3, 51.0, 46.2, 42.4, 40.7, 28.5.


**tert-Butyl (2-Oxo-2-(piperidin-1-yl)ethyl)carbamate (Boc-Gly-Pip, 11f)**

Yield: 90%; oil.

1H NMR (500 MHz, CDCl3): δ = 5.56 (br s, 1 H), 3.94 (d, J=3.5 Hz, 2 H), 3.56 (t, J=5 Hz, 2 H), 3.31 (t, J=5 Hz, 2 H), 1.65–1.63 (m, 6 H), 1.45 (s, 9 H).

13C NMR (125 MHz, CDCl3): δ = 167.9, 164.3, 79.6, 45.9, 44.2, 41.9, 28.4, 25.9, 24.8, 24.6.


**tart-Butyl (5)-(1-Oxoo-1-(piperidin-1-yl)propan-2-yl)carbamate (Boc-i-Ala-Pip, 11g)**

Yield: 97%; oil.

1H NMR (500 MHz, CDCl3): δ = 5.61 (d, J=8.4 Hz, 1 H), 4.64–4.58 (m, 1 H), 3.62–3.38 (m, 4 H), 1.68–1.53 (m, 6 H), 1.44 (s, 9 H), 1.29 (d, J=6.7 Hz, 1 H).

13C NMR (125 MHz, CDCl3): δ = 171.0, 155.3, 79.5, 46.6, 46.4, 43.4, 28.6, 28.6, 28.5, 26.6, 25.7, 24.7, 19.7.

HRMS (ESI): m/z[M + H]+ calcd for C26H29N3O3: 356.2131; found: 356.2134.

**tert-Butyl (5)-(3-Methyl-1-oxo-1-(piperidin-1-yl)butan-2-yl)carbamate (Boc-i-Val-Pip, 11h)**

Yield: 78%; oil.

1H NMR (500 MHz, CDCl3): δ = 5.37 (d, J=7.5 Hz, 1 H), 4.42 (t, J=7.5 Hz, 1 H), 3.54–3.41 (m, 4 H), 1.89–1.84 (m, 1 H), 1.60–1.45 (m, 6 H), 1.37 (s, 9 H), 0.90 (d, J=6.7 Hz, 3 H), 0.81 (d, J=6.7 Hz, 3 H).

13C NMR (125 MHz, CDCl3): δ = 170.8, 155.9, 79.4, 66.9, 66.7, 54.6, 46.3, 42.4, 31.4, 28.3, 19.6, 17.3.


**tert-Butyl ((2S,3S)-(3-Methyl-1-oxo-1-(piperidin-1-yl)pentan-2-yl)carbamate (Boc-i-Ile-Pip, 11i)**

Yield: 79%; oil.

1H NMR (500 MHz, CDCl3): δ = 5.33 (d, J=7.9 Hz, 1 H), 4.50 (t, J=7.9 Hz, 1 H), 3.59–3.49 (m, 4 H), 3.20–3.17 (m, 1 H), 1.70–1.53 (m, 8 H), 1.43 (s, 9 H), 0.93 (d, J=5.8 Hz, 3 H), 0.88 (t, J=7.5 Hz, 3 H).

13C NMR (125 MHz, CDCl3): δ = 170.7, 156.1, 79.5, 54.4, 47.1, 43.3, 38.5, 28.6, 28.6, 25.8, 24.7, 24.0, 16.2, 11.7.


**tert-Butyl (2S,3S)-(3-Methyl-1-(4-methylpiperazin-1-yl)-1-oxobutan-2-yl)carbamate (Boc-i-Val-NMP, 11j)**

Yield: 84%; oil.

1H NMR (500 MHz, CDCl3): δ = 5.34 (d, J=7.2 Hz, 1 H), 4.30 (d, J=7.2 Hz, 1 H), 3.53–3.39 (m, 4 H), 2.95–2.82 (m, 4 H), 2.15 (s, 3 H), 1.82–1.76 (m, 1 H), 1.29 (s, 9 H), 0.81 (d, J=6.7 Hz, 3 H), 0.74 (d, J=6.7 Hz, 3 H).

13C NMR (125 MHz, CDCl3): δ = 170.4, 155.8, 79.3, 55.2, 54.7, 45.9, 45.6, 41.9, 31.5, 28.3, 19.7, 17.1.


**tert-Butyl ((2S,3S)-3-Methyl-1-(4-methylpiperazin-1-yl)-1-oxobutanyl-2-yl)carbamate (Boc-i-Ile-NMP, 11l)**

Yield: 69%; oil.

1H NMR (500 MHz, CDCl3): δ = 5.26 (d, J=8.2 Hz, 1 H), 4.41 (t, J=8.2 Hz, 1 H), 3.66–3.46 (m, 4 H), 2.35–2.32 (m, 4 H), 2.24 (s, 3 H), 2.15–2.10 (m, 1 H), 1.65–1.60 (m, 1 H), 1.49–1.45 (m, 1 H), 1.29 (s, 9 H), 0.91–0.80 (m, 6 H).

13C NMR (125 MHz, CDCl3): δ = 170.9, 156.0, 79.6, 55.4, 54.9, 54.3, 46.1, 45.9, 42.1, 38.4, 28.5, 24.1, 16.1, 11.6.

tert-Butyl (5S)-1-(4-Methylpiperazin-1-yl)-1-oxo-3-phenylpropan-2-yl)carbamate (Boc-t-Phe-NMP, 11o)

Yield: 89%; oil.

1H NMR (500 MHz, CDCl3): δ = 5.38 (br s, 1 H), 8.70 (br s, 1 H), 8.53 (br s, 1 H), 4.23 (d, J=4.3 Hz, 2 H), 3.68–3.63 (m, 6 H), 3.46–3.41 (m, 2 H), 1.73–1.78 (m, 4 H), 1.30–1.35 (m, 4 H), 1.26–1.31 (m, 4 H).

13C NMR (125 MHz, CDCl3): δ = 168.3, 163.1, 79.6, 56.6, 48.1, 46.9, 45.7, 43.1, 28.6, 26.2, 24.3, 19.5, 17.5.


tert-Butyl (2-Oxo-2-(pyrrolidin-1-yl)ethyl)carbamate (Boc-Gly-Pyr, 11p)

Yield: 83%; oil.

1H NMR (500 MHz, CDCl3): δ = 5.48 (br s, 1 H), 4.47–4.42 (m, 1 H), 3.80–3.85 (m, 4 H), 1.99–1.86 (m, 4 H), 1.43 (s, 9 H), 1.31 (d, J=6.7 Hz, 3 H).

13C NMR (125 MHz, CDCl3): δ = 171.4, 155.3, 79.6, 48.0, 46.5, 46.1, 28.6. 26.2, 24.3, 18.9.


tert-Butyl (5S)-(1-Oxo-1-(pyrrolidin-1-yl)propan-2-yl)carbamate (Boc-t-Ala-Pyr, 11q)

Yield: 83%; oil.

1H NMR (500 MHz, CDCl3): δ = 5.26 (d, J=7.8 Hz, 1 H), 4.15 (t, J=7.8 Hz, 1 H), 3.60–3.30 (m, 4 H), 1.87–1.75 (m, 5 H), 1.37 (s, 9 H), 0.86 (d, J=6.7 Hz, 3 H).

13C NMR (125 MHz, CDCl3): δ = 170.7, 155.8, 79.2, 57.0, 46.6, 45.7, 31.3, 28.3, 26.0, 24.1, 19.5, 17.5.


tert-Butyl (5S)-(3-Methyl-1-oxo-1-(pyrrolidin-1-yl)butan-2-yl)carbamate (Boc-t-Val-Pyr, 11r)

Yield: 67%; oil.

1H NMR (500 MHz, CDCl3): δ = 3.50 (d, J=8.5 Hz, 1 H), 1.73–1.55 (m, 2 H), 1.43 (s, 9 H), 0.93–0.86 (m, 6 H).

13C NMR (125 MHz, CDCl3): δ = 171.1, 156.0, 79.6, 56.6, 48.1, 46.9, 46.0, 38.2, 28.6, 26.2, 24.4, 15.8, 11.5.


Synthesis of Bisconjugates 5a–t; General Procedure

The secondary amine–amino acid conjugate was stirred in 4 M HCl–dioxane solution for 1 h. The dioxane was evaporated under reduced pressure and the residue was treated with diethyl ether. The resulting solid was treated without further purification with the benzotriazolide derivative of pyrazinoic acid in the presence of triethylamine (1.5 equiv) in acetonitrile (10 mL). The reaction mixture was stirred at 20 °C for 4–6 h, monitoring by TLC. Upon completion of reaction, the acetonitrile was evaporated and the residue was extracted with EtOAc. The organic layer was washed with aqueous sodium carbonate and dried over anhydrous sodium sulfate. After filtration, the EtOAc was evaporated under reduced pressure to obtain the desired conjugates in good yields.

N-(2-Morpholino-2-oxoethyl)pyrazine-2-carboxamide (POA-Gly-Mor, 5a)

Yield: 75%; colorless microcrystals; mp 180–181 °C.

1H NMR (500 MHz, CDCl3): δ = 9.32 (s, 1 H), 8.70 (br s, 1 H), 8.65 (br s, 1 H), 8.53 (br s, 1 H), 4.23 (d, J=4.3 Hz, 2 H), 3.68–3.63 (m, 6 H), 3.46–3.44 (m, 2 H).

13C NMR (125 MHz, CDCl3): δ = 166.4, 163.3, 147.5, 144.3, 143.1, 66.8, 66.5, 45.1, 42.5, 41.2.


(S)-N-(1-Morpholino-1-oxopropan-2-yl)pyrazine-2-carboxamide (POA-t-Ala-Mor, 5b)

Yield: 68%; oil.

1H NMR (500 MHz, CDCl3): δ = 9.34 (s, 1 H), 8.74–8.70 (m, 2 H), 8.50 (br s, 1 H), 5.13–5.08 (m, 1 H), 3.69–3.53 (m, 8 H), 1.45 (d, J=6.9 Hz, 3 H).

13C NMR (125 MHz, CDCl3): δ = 170.8, 162.6, 147.5, 144.3, 143.0, 66.9, 66.7, 46.2, 45.4, 42.8, 19.0.


(S)-N-(1-Morpholino-1-oxopentan-2-yl)pyrazine-2-carboxamide (POA-t-Ile-Mor, 5b)

Yield: 72%; gum.

1H NMR (500 MHz, CDCl3): δ = 9.33 (s, 1 H), 8.74–8.68 (m, 2 H), 8.50 (br s, 1 H), 5.16–5.07 (m, 1 H), 3.69–3.55 (m, 8 H), 1.44 (d, J=6.9 Hz, 3 H).

13C NMR (125 MHz, CDCl3): δ = 170.8, 162.4, 147.5, 145.0, 143.1, 66.9, 66.7, 46.2, 45.4, 42.7, 19.0.

(5)-N-(3-Methyl-1-morpholinol-1-oxobutan-2-yl)pyrazine-2-carboxamide (POA-1-Va-Mol, 5c)

Yield: 76%; oil.

1H NMR (500 MHz, CDCl3): δ = 9.37 (s, 1 H), 8.76 (br s, 1 H), 8.58 (br s, 1 H), 8.60 (d, J=7.2 Hz, 1 H), 8.50 (d, J=5.6 Hz, 2 H), 8.42 (d, J=8.7 Hz, 1 H), 8.00–7.97 (m, 1 H), 3.77–3.60 (m, 8 H), 2.19–2.13 (m, 1 H), 0.05 (d, J=6.6 Hz, 3 H), 0.10 (d, J=6.6 Hz, 3 H).

13C NMR (125 MHz, CDCl3): δ = 169.8, 163.0, 147.3, 144.3, 142.7, 66.7, 66.6, 53.2, 46.3, 42.4, 31.7, 19.7, 17.5.


N-((2S,3S)-3-Methyl-1-morpholinol-1-oxopentan-2-yl)pyrazine-2-carboxamide (POA-1-Ile-Pip, 5d)

Yield: 70%; oil.

1H NMR (500 MHz, CDCl3): δ = 9.37 (s, 1 H), 8.76 (br s, 1 H), 8.57 (br s, 1 H), 8.46 (d, J=7.9 Hz, 1 H), 5.02 (t, J=7.9 Hz, 1 H), 3.76–3.52 (m, 8 H), 1.94–1.93 (m, 1 H), 1.64–1.59 (m, 1 H), 1.23–1.16 (m, 1 H), 0.92–0.91 (m, 6 H).

13C NMR (125 MHz, CDCl3): δ = 167.4, 163.0, 147.5, 144.5, 142.9, 67.0, 66.9, 52.8, 46.7, 42.7, 38.4, 24.4, 161.1, 11.4.


(5)-N-(1-Morpholinol-1-oxo-3-phenylpropan-2-yl)pyrazine-2-carboxamide (POA-1-Phe-Pip, 5e)

Yield: 78%; gum.

1H NMR (500 MHz, CDCl3): δ = 9.36 (s, 1 H), 8.75 (br s, 1 H), 8.59–856 (m, 2 H), 7.33–7.26 (m, 5 H), 5.35–5.30 (m, 1 H), 3.61–3.32 (m, 6 H), 3.20–3.09 (m, 2 H), 3.00–2.89 (m, 2 H).

13C NMR (125 MHz, CDCl3): δ = 169.7, 162.6, 147.6, 144.5, 143.0, 136.2, 129.8, 128.9, 127.6, 66.7, 66.2, 49.8, 46.3, 42.5, 40.4.


N-(2-Oxo-2-(piperidin-1-yl)ethyl)pyrazine-2-carboxamide (POA-Gly-Pip, 5f)

Yield: 74%; colorless microcrystals; mp 123–125 °C.

1H NMR (500 MHz, CDCl3): δ = 9.38 (s, 1 H), 8.77–8.74 (m, 2 H), 8.58 (br s, 1 H), 4.27 (d, J=4.3 Hz, 2 H), 3.65 (t, J=5.2 Hz, 2 H), 3.41 (t, J=5.2 Hz, 2 H), 1.69–1.59 (m, 2 H).

13C NMR (125 MHz, CDCl3): δ = 165.8, 163.3, 147.5, 144.4, 143.1, 45.8, 43.4, 41.4, 41.6, 26.4, 25.6, 24.6.


(5)-N-(1-Oxopyridin-1-yl)propan-2-yl)pyrazine-2-carboxamide (POA-1-Phe-Pip, 5j)

Yield: 79%; oil.

1H NMR (500 MHz, CDCl3): δ = 9.34 (s, 1 H), 8.71 (br s, 1 H), 8.62–8.56 (m, 2 H), 7.34–7.24 (m, 5 H), 5.40–5.35 (m, 1 H), 3.57–3.47 (m, 4 H), 2.94–2.78 (m, 2 H), 1.54–1.43 (m, 6 H).

13C NMR (125 MHz, CDCl3): δ = 169.8, 163.3, 147.8, 144.5, 143.9, 138.0, 129.5, 128.6, 127.2, 52.5, 49.9, 46.3, 43.2, 26.2, 25.6, 24.4.


N-(2-(4-Methylpiprazin-1-yl)-2-oxoethyl)pyrazine-2-carboxamide (POA-Gly-NMP, 5k)

Yield: 69%; gum.

1H NMR (500 MHz, CDCl3): δ = 9.34 (s, 1 H), 8.71 (br s, 1 H), 8.68 (br s, 1 H), 8.54 (br s, 1 H), 4.26 (d, J=4.3 Hz, 2 H), 3.75–3.00 (m, 4 H), 2.42–2.35 (m, 4 H), 2.31 (s, 3 H).

13C NMR (125 MHz, CDCl3): δ = 169.4, 163.3, 147.8, 144.5, 143.4, 55.3, 54.8, 46.4, 46.1, 45.9, 40.8.


(5)-N-(1-(4-Methylpiprazin-1-yl)-1-oxopropan-2-yl)pyrazine-2-carboxamide (POA-1-Ala-NMP, 5l)

Yield: 73%; oil.

1H NMR (500 MHz, CDCl3): δ = 3.92 (s, 1 H), 8.72–8.70 (m, 2 H), 8.53 (br s, 1 H), 5.07–5.04 (m, 1 H), 3.72–3.51 (m, 4 H), 2.44–2.34 (m, 4 H), 2.27 (s, 3 H), 1.42 (d, J=6.7 Hz, 3 H).

13C NMR (125 MHz, CDCl3): δ = 170.3, 162.3, 147.4, 144.3, 142.9, 55.1, 54.6, 46.0, 45.4, 43.2, 19.1.

(S)-N-(3-Methyl-1-(4-methylpiperazin-1-yl)-1-oxobutan-2-yl)pyrazine-2-carboxamide (POA-L-Phe-NMP, 5m)
Yield: 62%; oil.
1H NMR (500 MHz, CDCl3); δ = 9.37 (s, 1 H), 8.75 (br s, 1 H), 8.57 (br s, 1 H), 8.52 (d, J=8.9 Hz, 1 H), 7.50–5.00 (m, 1 H), 3.76–3.61 (m, 4 H), 2.46–2.38 (m, 4 H), 2.31 (s, 3 H), 2.17–2.13 (m, 1 H), 1.04 (d, J=6.7 Hz, 3 H), 0.99 (d, J=6.7 Hz, 3 H).
13C NMR (125 MHz, CDCl3); δ = 169.8, 163.3, 147.5, 144.4, 143.0, 55.4, 54.3, 46.2, 46.0, 42.3, 32.1, 20.1, 17.7.

N-((2S,5S)-3-Methyl-1-(4-methylpiperazin-1-yl)-1-oxopentan-2-yl)pyrazine-2-carboxamide (POA-L-Ile-NMP, 5n)
Yield: 79%; oil.
1H NMR (500 MHz, CDCl3); δ = 9.32 (s, 1 H), 8.70 (br s, 1 H), 8.52 (br s, 1 H), 8.45 (d, J=7.8 Hz, 1 H), 5.01 (t, J=7.8 Hz, 1 H), 3.72–3.60 (m, 4 H), 2.42–2.36 (m, 4 H), 2.27 (s, 3 H), 1.89–1.88 (m, 1 H), 1.57–1.53 (m, 1 H), 1.18–1.13 (m, 1 H), 0.98–0.86 (m, 6 H).
13C NMR (125 MHz, CDCl3); δ = 169.9, 162.9, 147.4, 144.4, 142.9, 55.3, 54.7, 52.9, 46.2, 42.2, 38.4, 24.3, 16.1, 11.4.

(5)-N-(1-(4-Methylpiperazin-1-yl)-1-oxo-3-phenylpropan-2-yl)pyrazine-2-carboxamide (POA-L-Phe-NMP, 5o)
Yield: 71%; oil.
1H NMR (500 MHz, CDCl3); δ = 9.34 (s, 1 H), 8.73 (br s, 1 H), 8.58–8.54 (m, 2 H), 7.30–7.19 (m, 5 H), 5.30–5.24 (m, 1 H), 3.72–3.61 (m, 4 H), 3.20–3.09 (m, 2 H), 2.47–2.38 (m, 4 H), 2.24 (s, 3 H).
13C NMR (125 MHz, CDCl3); δ = 170.1, 162.8, 147.5, 144.7, 143.2, 136.5, 129.3, 128.7, 127.2, 55.3, 54.7, 45.6, 46.2, 42.6, 40.8.

N-(2-Oxo-2-(pyrrolidin-1-yl)ethyl)pyrazine-2-carboxamide (POA-Gly-Pyr, 5p)
Yield: 81%; colorless microcrystals; mp 148–150 °C.
1H NMR (500 MHz, CDCl3); δ = 9.31 (s, 1 H), 8.68–8.64 (m, 2 H), 8.52 (br s, 1 H), 4.15 (d, J=4.3 Hz, 2 H), 3.50–3.40 (m, 4 H), 2.00–1.78 (m, 4 H).
13C NMR (125 MHz, CDCl3); δ = 166.1, 163.3, 147.4, 144.3, 143.0, 68.1, 46.2, 45.7, 42.1, 26.1, 24.3.

(5)-N-(1-Oxo-1-(pyrrolidin-1-yl)propan-2-yl)pyrazine-2-carboxamide (POA-L-Ala-Pyr, 5q)
Yield: 73%; oil.
1H NMR (500 MHz, CDCl3); δ = 9.33 (s, 1 H), 8.73 (br s, 1 H), 8.59–8.55 (m, 2 H), 7.35–7.23 (m, 5 H), 5.14–5.09 (m, 1 H), 3.75–3.34 (m, 4 H), 3.03–2.93 (m, 2 H), 2.18–1.98 (m, 4 H), 1.00 (d, J=6.6 Hz, 3 H), 0.97 (d, J=6.6 Hz, 3 H).
13C NMR (125 MHz, CDCl3); δ = 160.9, 162.3, 147.3, 144.2, 142.8, 136.2, 129.2, 128.3, 126.6, 52.5, 45.9, 45.8, 39.7, 24.0, 21.8.

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