Combination Antibiotic Treatment of Serious Methicillin-Resistant Staphylococcus aureus Infections

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Methicillin-resistant Staphylococcus aureus (MRSA) is the most common antibiotic-resistant human pathogen, with an estimated 1,650 cases of bloodstream infection and 500 deaths annually in Australia,1 11,000 deaths annually in the United States,2 and an annual excess cost to Europe’s health care system of €380m.3 MRSA bacteremia has a mortality of 20 to 30%, exceeding that of methicillin-sensitive S. aureus (MSSA) at least in part due to the shortcomings of vancomycin, the standard therapy for MRSA bacteremia. Vancomycin still remains the principal agent of choice in the treatment of MRSA.4 However, multiple shortcomings of glycopeptide monotherapy have been recognized and include poor tissue penetration, slow bactericidal effect, and emergence of resistance during therapy. Combination therapy may theoretically overcome some of these deficiencies.
Consequently, when exploring therapeutic combinations, the addition of a second agent is usually directed to address one or more of the above vancomycin deficiencies: to broaden the spectrum of activity (to include resistant isolates such as heteroresistant vancomycin intermediate S. aureus [hVISA]) and increase the bacteriocidal activity of vancomycin through synergy. Other potential benefits of adding antibiotics include enhancing treatment by providing better tissue or biofilm penetration and reducing toxin production especially in toxin-mediated infections. The alternative to combination therapy is to find a better agent than vancomycin. Several agents with useful activity against MRSA have reached the market in recent years, including daptomycin, linezolid, tigecycline, and quinupristin/dalfopristin. However, none of these agents has been shown to be superior to vancomycin, with clinical trials showing daptomycin is noninferior for S. aureus bacteremia (SAB) and endocarditis and linezolid noninferior for S. aureus catheter-related blood stream infection, and although linezolid achieved improved clinical cure for MRSA pneumonia, it did not result in reduced mortality. Hence, combination therapy is an attractive possibility for improving outcomes from severe MRSA infections.

Most data on combination therapy come from in vitro experiments. These have three possible outcomes: synergy (increased bacteriocidal activity in excess of the additive actions of the two drugs), antagonism, and indifference. The most common in vitro methods used to measure synergy (in increasing order of complexity) are agar diffusion, checkerboard testing, time-kill curves, and simulated pharmacodynamic (PD) models. Agar diffusion methods use combinations of antibiotics either as antibiotic discs or Etest strips (Biomerieux, Paris, France; paper strips of graduated antibiotic concentration), allowing them to diffuse through the agar, and qualitatively examines bacterial growth in different zones of the agar. For example, two Etest strips (one for each antibiotic) can be placed at right angles on the agar plate, and bacterial growth examined near the intersection (where the combination concentration is highest) compared with the outer quadrants. Checkerboard testing uses multiple different dilutions of antibiotic combinations set out on an agar plate as a grid, ranging from below to above the minimum inhibitory concentration (MIC) for each agent. This allows a quantitative estimate of synergy and results are reported as a fractional inhibitory concentration (FIC). An FIC of ≤0.5 indicates synergy, >2.0 antagonism, and an FIC between these values indicates indifference. Time-kill methodology is dynamic rather than examining synergy at a single time point and thus is more likely to correlate with clinical use. Bacteria are cultured in liquid media in the presence of various concentrations of either single or combination antibiotics, and the speed of bacterial killing is quantified over time. In time-kill studies, synergy is defined as a reduction in ≥2 log colony forming units (cfu) of bacteria/mL compared with the cfu/mL of the most potent single drug. Finally, in vitro pharmacokinetic (PK)/PD models attempt to simulate antibiotic dosing within a human or animal host. An example is the fibrin clot model, where an ex vivo human blood clot is seeded with bacteria and exposed to dynamic concentrations of antibiotic, simulating a usual dosing interval.

**Combinations of Two Primarily Active Agents**

**Vancomycin and Linezolid**

Linezolid is an oxazolidinone, a new class of antibiotic with a mechanism of action directed at the early steps of protein synthesis. Its introduction into clinical medicine heralded the first new antibiotic with activity against MRSA since the introduction of vancomycin.

Linezolid and vancomycin combination therapy are reported to demonstrate indifference using checkerboard assays, but synergy with time-kill assays. Other studies, using similar methodology, were unable to demonstrate synergy but observed antagonism or indifference. Subsequent animal data have similarly been conflicting with an experimental rabbit endocarditis model, observing some effect with the addition of linezolid, while a rat MSSA osteomyelitis model showed indifference with no additional sterilization or cure rates compared with vancomycin alone. The inconsistency of these results suggests that there are a variety of mechanisms involved in determining antibiotic interactions, which may include the infecting bacterial strain, infection site, and host responses. Thus, this combination should be used with caution as the described antagonism may lead to suboptimal outcomes.

Nevertheless, this combination is still considered in toxin-mediated infections, as subinhibitory concentrations of linezolid inhibit S. aureus toxin production. This modulation of virulence factors through reduced pathogen toxin synthesis may theoretically attenuate disease and influence outcomes. This hypothesis has not been shown in vivo to be effective, with no animal model data available. Although a single case report has been published showing the effectiveness of linezolid in the treatment of MSSA toxic shock syndrome, no comparative data of the combination of linezolid with vancomycin in toxin-mediated clinical syndromes has been published.

**Vancomycin and Daptomycin**

Daptomycin is a lipopeptide, which acts on the cell membrane through a complex process resulting in cell membrane depolarization and permeabilization, ion leakage, and ultimately cell death. There is a paucity of in vitro and in vivo data for this combination due to concerns about the relationship between reduced vancomycin susceptibility and daptomycin nonsusceptibility. This cross-resistance was first documented in vitro when a collection of VISA isolates underwent susceptibility testing and 80% were found to be daptomycin nonsusceptible, despite these isolates not having been exposed to daptomycin. This association was later confirmed by serial passage studies, with all S. aureus isolates developing daptomycin nonsusceptibility in the presence of vancomycin. Subsequently, several clinical cases have described the same phenomenon. Although the precise mechanism remains unclear, experts speculate that increased cell wall thickness, as occurs in VISA, may also prevent daptomycin penetration to its site of action.
Nevertheless, Tsuji and Rybak performed Etest synergy testing and time-kill experiments on one vancomycin heteroresistant and one vancomycin intermediate clinical S. aureus isolate. There was moderate agreement between the two methodologies with vancomycin/daptomycin combination showing either indifference or an additive effect (but not synergy). This combination, albeit with the addition of rifampin, has been used in one published report of two cases. Both patients had orthopedic infections that relapsed after initial vancomycin/rifampin therapy and were cured with several weeks of triple therapy. As both patients also underwent surgical debridement and prosthesis removal, the effectiveness of daptomycin/vancomycin remains unknown, but based on the above concerns, this combination is unlikely to be successful in most cases.

**Vancomycin and Tetracyclines including Tigecycline**

hVISA was first isolated in Japan in the 1990s. Given the lack of treatment options against hVISA, combination therapy was studied. Time-kill experiments using the Mu3 strain, the first recognized hVISA isolate, revealed antagonism when using minocycline/vancomycin in combination. The authors went on to examine possible bacterial changes that may result in this antagonism and found that cell wall thickening did not play a major role.

Tigecycline is a semisynthetic derivative of minocycline and is the first glycycline antibiotic licensed for clinical use. It offered great promise as a new agent with a broad spectrum of activity against gram-negative and -positive bacteria, including multiresistant organisms. However, postmarketing experience has resulted in more selective indications, as it is associated with increased mortality when used in certain clinical settings. Nevertheless, given its spectrum of activity, using it in combination with vancomycin remains an attractive option. Mercier et al performed time-kill studies and found vancomycin and tigecycline to be indifferent when used with four MRSA (three of which were VISA) isolates. Petersen et al obtained similar results using checkerboard and time-kill experiments on 10 S. aureus isolates. There have been few experimental animal studies evaluating tigecycline combinations. A rabbit osteomyelitis model found no difference between combination therapy compared with vancomycin or tigecycline monotherapy. Similarly, no difference was observed in bactericidal activity in a biofilm model for this combination.

The main conclusion from the available data is that there is no benefit with tetracycline and vancomycin combinations. Unless tigecycline is required for a coexistent multiresistant gram-negative infection, combination with vancomycin is not recommended for the treatment of MRSA infections.

**Daptomycin and Tigecycline**

Time-kill and Etest experiments demonstrated an indifferent effect when using this combination. These results are in contrast to a subsequent study, which showed this combination to be synergistic based on time-kill studies and checkerboard titration assays using 10 S. aureus isolates. To corroborate their results, the authors went on to perform an animal surgical site infection model. Tigecycline/daptomycin still showed synergy. No clinical data are available. The role of tigecycline is likely to be limited as discussed previously and thus further studies are unlikely to occur with this combination.

**Daptomycin and Linezolid**

The impact of biofilm-associated infection remains significant especially in light of the aging population and increased joint replacements undertaken. Linezolid/daptomycin combination was studied by Parra-Ruiz et al in their validated in vitro PK/PD biofilm formation model. This study observed that linezolid/daptomycin combination theory showed better efficacy than either agent alone and confirmed the results of one previous study using a simulated endocardial vegetation model. Although a single published case report showed clinical benefit, this occurred in the setting of triple therapy with daptomycin, linezolid, and rifampin. Thus, the benefit of this combination is unclear especially as checkerboard and time-kill experiments showed antagonism.

**Vancomycin and Quinupristin–Dalfopristin**

Quinupristin–dalfopristin is an injectable streptogramin antibiotic with in vitro activity against MRSA. The combination of quinupristin–dalfopristin with vancomycin has resulted in mixed results with studies demonstrating both antagonism and synergy. There remain very limited published clinical data to guide the use of this combination. Given that quinupristin–dalfopristin is not recommended for MRSA bacteremia due to reports of treatment failures and emergence resistance, and there are no published original studies since 2002 on the combination of quinupristin–dalfopristin with vancomycin, it is unlikely to be further advanced as a clinical treatment option.

**β-Lactam Combination Therapy**

Empirical therapy for SAB often includes both vancomycin and antistaphylococcal penicillin such as nafcillin. This is not only because incorrect initial empiric therapy for MRSA bacteremia is associated with a twofold increase in mortality but also because vancomycin monotherapy for MSSA infections is associated with higher rates of hospitalization, treatment failure, and mortality compared with β-lactam therapy (e.g., with nafcillin or cefazolin). Given the increasing prevalence of community-acquired MRSA infection, β-lactam combination therapy is often used in patients with positive blood cultures where the Gram stain shows clustered gram-positive cocci for the first 24 to 48 hours of therapy, but before identification and susceptibility profile of the organism has been determined. Hence a potentially synergistic combination is unwittingly being increasingly used in the subset that turn out to have MRSA infection. Considering the very definition of MRSA is that it is resistant to antistaphylococcal penicillins, it is counter-intuitive to hypothesize that β-lactams might have any benefit when added to standard therapy for MRSA infections. However, unexpected synergy between β-lactams and both vancomycin and daptomycin has been found to occur in vitro, initially only in VISA and hVISA strains, and then more broadly in MRSA.
Vancomycin/β-Lactam Combinations

In Vitro Studies
At least 16 in vitro studies have explored synergy between vancomycin and β-lactams against MRSA isolates, but one of which found evidence of synergy in some or all of the tested strains (Table 1). These studies varied in their methodology (checkerboard synergy testing or time-kill curves), types of strains tested (MRSA vs. hVISA vs. VISA) and the β-lactams used, but a consistent finding across nearly all the studies was synergistic bacterial killing in most but not all strains tested. There are no consistent characteristics in these reports in the strains where synergy was not demonstrated. However, there was a general tendency across these studies (and within some studies) to an increasing degree of synergy with increasing vancomycin MICs. The one study that did not demonstrate synergy did not actually include any MRSA isolates. In this study, Joukhadar et al tested 10 clinical isolates of methicillin sensitive S. aureus and found evidence of neither synergy nor antagonism in any strain, both using fixed drug concentrations, and in a dynamic model simulating clinical dosing.

Animal Studies
The few studies that have assessed combinations of vancomycin with β-lactams in animal models have all found evidence of synergy. Climo et al found faster sterilization of infection with vancomycin plus nafcillin in MRSA rabbit endocarditis and renal abscess models. Ribes et al tested various combinations of linezolid, vancomycin, and imipenem in a murine peritonitis VISA model using time-kill curves, and found faster bacterial killing with vancomycin plus imipenem compared with vancomycin alone, in both strains tested. Finally, Fernandez et al investigated the anti-MRSA cefepime ceftobiprole against an MRSA and a VISA strain in a rat endocarditis model. They found good activity of ceftobiprole against both strains in terms of sterilizing vegetations and preventing mortality; the combination of vancomycin plus ceftobiprole led to faster killing on time-kill curves, but similar rates of mortality and of sterilization of vegetations compared with ceftobiprole alone.

Human Studies
There are currently no published prospective controlled trials of vancomycin/β-lactam combination therapy in patients with MRSA bacteremia, but one observational study has recently been published. In this single-center retrospective cohort study, Dilworth et al described the outcomes of 50 patients with MRSA bacteremia who received combination therapy with vancomycin and at least 24 hours of β-lactam (at their clinicians’ discretion), and compared them with 30 patients treated at the same hospital, during the same time period with vancomycin alone. They found a higher rate of microbiological eradication in the combination therapy group (96 vs. 80%, p = 0.02), which persisted on a multivariate model attempting to control for potential confounders (adjusted odds ratio for achieving microbiological eradication in the combination group = 11.24, p = 0.01).

Daptomycin/β-Lactam Combinations

In Vitro Studies
At least 10 in vitro studies have examined the combination of daptomycin with various β-lactams against MRSA and VISA strains (Table 2). The findings of these studies are remarkably similar to the vancomycin/β-lactam synergy articles cited earlier: synergy for most but not all strains tested, and an increasing degree of synergy with increasing MICs to both vancomycin and daptomycin. No studies have found evidence of antagonism with this combination.

Animal Studies
A recently published animal study mirrored the findings of the in vitro studies. Garrigós et al used a rat tissue cage model of MRSA infection to study the combination of daptomycin with cloxacillin, and found superior cure rates with the combination than with daptomycin alone.

Human Studies
As for the vancomycin/β-lactam combination, there are no clinical trials of daptomycin with β-lactams either published or in trials registries. However, limited observational data suggest this combination may be effective, particularly MRSA with poor response to daptomycin. In a case series of seven patients with persistent MRSA bacteremia for more than 1 week despite high-dose daptomycin, all cleared their bacteremia within 48 hours once nafcillin or oxacillin was added to their therapy. In a second case series of 22 patients with persistent MRSA bacteremia despite daptomycin for a median of 10 days, the addition of ceftaroline lead to clearance of bacteremia in all cases, in a median of 2 days.

Summary
Although the studies on β-lactam combination therapy are heterogeneous, there are some consistent findings (with either vancomycin or daptomycin as the companion agent): adding a β-lactam leads to synergistic bacterial killing in the majority of strains tested and in all animal models tested. The most consistent data come from more resistant strains and from antistaphylococcal penicillins or ceftaroline rather than other β-lactams. β-lactam combination therapy (with either vancomycin or daptomycin) is not recommended in Infectious Diseases Society of America (IDSA) or other guidelines at this stage, but the emerging data are intriguing, and at least one phase 2b randomized controlled trial (RCT) of this strategy is underway (Australia and New Zealand Clinical Trials Registry number ACTRN1261000940077). A key question that emerges from these data is: what is the mechanism of the observed synergy? The mechanisms have not been entirely elucidated, but are becoming clearer over time. Increasing vancomycin resistance in S. aureus is paradoxically associated with decreasing MICs to oxacillin, and this so-called “see-saw effect” is at least in part due to alteration of the MecA gene in some strains of VISA and vancomycin resistant Staphylococcus aureus (VRSA). and possibly to other structural changes in penicillin-binding proteins. β-lactams have been shown to enhance binding of daptomycin to the bacterial cell wall. Finally, Sakoulas et al recently
Table 1 In-vitro and animal studies investigating synergy between vancomycin and beta-lactams against MRSA

<table>
<thead>
<tr>
<th>Author (y)</th>
<th>Setting</th>
<th>Organism</th>
<th>Base antibiotic(s)</th>
<th>Beta-lactam(s)</th>
<th>Finding</th>
<th>Comments</th>
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<tbody>
<tr>
<td>Seibert et al (1992)</td>
<td>In vitro</td>
<td>MRSA</td>
<td>Vancomycin</td>
<td>Cefpirome Cefoperazone Ceftazidime</td>
<td>Synergy with cefpirome/Vancomycin and cefoperazone/Vancomycin but not ceftazidime/Vancomycin</td>
<td>Synergy with “most” MRSA strains</td>
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<td>Palmer and Rybak (1997)</td>
<td>In vitro</td>
<td>MRSA</td>
<td>Vancomycin</td>
<td>Pip-Tazo Imipenem Nafcillin</td>
<td>Synergy with Vancomycin/Imipenem and Vancomycin/Nafcillin, for hVISA but not VISA</td>
<td>Time-kill studies in infected fibrin clots</td>
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<td>Climo et al (1999)</td>
<td>In vitro</td>
<td>Rabbit endocarditis model Rabbit renal abscess model</td>
<td>MRSA MRCNS</td>
<td>Vancomycin Oxacillin Ceftriaxone Ceftazidime Amoxy-clav</td>
<td>In vitro synergy for all combinations In vivo only nafcillin tested—synergy for sterilizing vegetations and renal abscesses</td>
<td>Synergy was proportional to Vancomycin MIC</td>
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<td>Ribes et al (2010)</td>
<td>In vitro</td>
<td>hVISA VISA</td>
<td>Vancomycin Linlinezolid</td>
<td>Imipenem</td>
<td>Linezolid plus imipenem most effective Combo in vivo</td>
<td>Linezolid plus Vancomycin antagonistic</td>
</tr>
<tr>
<td>Sika et al (2011)</td>
<td>In vitro</td>
<td>MRSA MRCNS</td>
<td>Vancomycin</td>
<td>Imipenem</td>
<td>Synergy in 21/22 strains</td>
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<td>Hagihara et al (2012)</td>
<td>In vitro</td>
<td>MRSA hVISA VISA</td>
<td>Vancomycin</td>
<td>Cefazolin</td>
<td>Synergy for all strains</td>
<td>In vitro PK/PD model</td>
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<td>Fernandez (2012)</td>
<td>Rat endocarditis model</td>
<td>MRSA VISA</td>
<td>Vancomycin</td>
<td>Cefotobiprole</td>
<td>Synergy for Vancomycin + cefotobiprole</td>
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<td>Leonard (2012)</td>
<td>In vitro</td>
<td>hVISA MRSA MSSA</td>
<td>Vancomycin</td>
<td>Nafcillin</td>
<td>23/25 h VISA strains synergy. Also against MRSA and MSSA, but least effect against MSSA</td>
<td>5 strains had PK/PD simulations—also demonstrated synergy</td>
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<tr>
<td>Werth et al (2013)⁵⁸</td>
<td>In vitro</td>
<td>hVISA hVISA</td>
<td>Daptomycin Vancomycin</td>
<td>Ceftroline</td>
<td>Daptomycin + ceftaroline synergistic against both strains; Vancomycin plus ceftaroline only against the daptomycin S strain</td>
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<tr>
<td>Werth et al (2013)⁵⁹</td>
<td>In vitro</td>
<td>VISA VISA</td>
<td>Vancomycin</td>
<td>Ceftaroline Oxacillin</td>
<td>Vancomycin/Oxa synergy in 3/5 VISA. Vancomycin/ceftaroline in 5/5 VISA</td>
<td>Inverse correlation between Vancomycin and β-lactam susceptibility</td>
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<td>Dilworth et al (2014)⁶⁷</td>
<td>In vitro</td>
<td>MRSA VISA</td>
<td>Vancomycin</td>
<td>Pip-Tazo Oxacillin</td>
<td>Synergy in all strains for both oxa and Pip-Tazo</td>
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Abbreviations: (h)VISA, (heteroresistant) vancomycin intermediate Staphylococcus aureus; MRSA, methicillin-resistant Staphylococcus aureus; MSSA, methicillin susceptible Staphylococcus aureus; MRCNS, methicillin-resistant coagulase negative Staphylococcus; Pip-Tazo, piperacillin–tazobactam; PK-PD, pharmacokinetic/pharmacodynamic.

Rifampin

Rifampin is an attractive as an adjunctive agent as it is bactericidal at a low concentration. However, it has activity against some extracellular bacterial forms, such as biofilms. Despite this, there is evidence suggesting that rifampin therapy increases bacterial resistance to vancomycin. In animal and in vitro studies, rifampin was able to penetrate the biofilm and increase the killing of vancomycin-resistant bacteria. However, these findings have not been replicated in other in vitro and animal experiments. Specifically, addressing the in vivo and animal experiments, addressing the use of rifampin in the presence of biofilms is important. There is some evidence in vitro and in animal models of osteomyelitis that the combination of rifampin and vancomycin is synergistic. In contrast, rifampin plus vancomycin is not synergistic in the presence of biofilms. rifampin’s ability to increase the killing of vancomycin-resistant bacteria in vitro and in animal models may be of therapeutic benefit in the treatment of MRSA infections. However, these findings have not been replicated in other studies.

Other Combination Therapy

Rifampin is often used as an adjunctive agent in the treatment of MRSA infections. rifampin is bactericidal at low concentrations and has activity against some extracellular bacterial forms, such as biofilms. However, there is evidence suggesting that rifampin therapy may increase bacterial resistance to vancomycin. In animal and in vitro studies, rifampin was able to penetrate the biofilm and increase the killing of vancomycin-resistant bacteria. However, these findings have not been replicated in other in vitro and animal experiments. Specifically, addressing the in vivo and animal experiments, addressing the use of rifampin in the presence of biofilms is important. There is some evidence in vitro and in animal models of osteomyelitis that the combination of rifampin and vancomycin is synergistic. In contrast, rifampin plus vancomycin is not synergistic in the presence of biofilms. rifampin’s ability to increase the killing of vancomycin-resistant bacteria in vitro and in animal models may be of therapeutic benefit in the treatment of MRSA infections. However, these findings have not been replicated in other studies.
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<td>Silva et al (1988)</td>
<td>In vitro</td>
<td>MRSA, Enterococci, Peptococci</td>
<td>Daptomycin</td>
<td>Aztreonam, Ceftriaxone</td>
<td>Synergy against all MRSA strains tested</td>
<td>Time-kill curves</td>
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<td>Rand and Houck (2004)</td>
<td>In vitro</td>
<td>MRSA (18 strains)</td>
<td>Daptomycin</td>
<td>Oxacillin, Amp-Sul, Tic-Clav, Pip-Tazo</td>
<td>All combinations synergistic</td>
<td>Sub-MICs of daptomycin</td>
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<td>Snydman et al (2005)</td>
<td>In vitro</td>
<td>MRSA, MSSA, VRE</td>
<td>Daptomycin</td>
<td>Oxacillin, Ceftriaxone, Ceftazidime, Imipenem</td>
<td>Indifference of all combinations for MSSA; Synergy in 7/10 MRSA isolates</td>
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<td>Cilli et al (2006)</td>
<td>In vitro</td>
<td>VRE, MRSA, MRSE</td>
<td>Daptomycin</td>
<td>Amp-Sul, Tic-Clav, Pip-Tazo</td>
<td>Synergy in &gt; 70% of MRSA strains</td>
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<td>Tsuji and Rybak (2006)</td>
<td>In vitro</td>
<td>hVISA (2 strains)</td>
<td>Daptomycin, Vancomycin</td>
<td>Ampicillin-Sulbactam</td>
<td>Synergy with Vancomycin/AS but indifference with daptomycin/AS</td>
<td>Etest and time kill</td>
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<td>Mehta et al (2012)</td>
<td>In vitro</td>
<td>MRSA (Dap S and Dap R)</td>
<td>Daptomycin</td>
<td>Nafillin, Cefotaxime, Azmyo-Clav, Imipenem</td>
<td>Synergy with all blc combinations, esp oxacillin</td>
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<td>Garrigós et al (2012)</td>
<td>Animal model</td>
<td>MRSA</td>
<td>Daptomycin</td>
<td>Cloxacillin</td>
<td>Combo better cure rates, but only mildly. Daptomycin-Rif was better</td>
<td>Rat tissue cage model (foreign body infection)</td>
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<tr>
<td>Werth et al (2013)</td>
<td>In vitro</td>
<td>hVISA, DNS hVISA</td>
<td>Daptomycin, Vancomycin</td>
<td>Ceftaroline</td>
<td>Combination synergistic for both strains with daptomycin, for one strain with Vancomycin</td>
<td>Time-kill curves</td>
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<td>Leonard and Rolek (2013)</td>
<td>In vitro</td>
<td>VISA</td>
<td>Daptomycin</td>
<td>Nafillin</td>
<td>Synergy in 11/20</td>
<td>Increasing synergy with increasing daptomycin MIC</td>
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<td>Werth et al (2014)</td>
<td>In vitro</td>
<td>hVISA, Daptomycin R VISA</td>
<td>Daptomycin, Vancomycin</td>
<td>Ceftaroline</td>
<td>Daptomycin + ceftaroline syner- gistic against both strains; Vancomycin plus ceftaroline only against the daptomycin S strain</td>
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<td>Barber et al (2014)</td>
<td>In vitro</td>
<td>MRSA (20 isolates)</td>
<td>Daptomycin</td>
<td>Ceftobiprole</td>
<td>Dapt plus ceftobiprole syner- gistic in all isolates</td>
<td>Time-kill curves</td>
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Abbreviations: (h)VISA, (heteroresistant) vancomycin intermediate *Staphylococcus aureus*; MRSA, methicillin-resistant *Staphylococcus aureus*; MSSA, methicillin susceptible *Staphylococcus aureus*; MR CNS, methicillin-resistant coagulase negative *Staphylococcus*; Pip-Tazo, piperacillin–tazobactam; PK-PD, pharmacokinetic/pharmacodynamic; DNS, Daptomycin non susceptible.
aureus bacteremia [ARREST study]. In a review of four previously published RCTs, Thwaites et al determined that adjunctive rifampin for serious staphylococcal infections was associated with a reduction in infection-related deaths by 55% \((p = 0.02)\). Notably, two of these studies principally focused on MSSA infections treated with oxacillin, and the combined findings in this systematic review probably do not apply to MRSA and vancomycin therapy. In the ARREST trial, MRSA bacteremia is not a prespecified subgroup. Thus, despite this being the largest planned RCT for SAB to date, the study may be underpowered to make specific conclusions regarding the MRSA subgroup, particularly as reductions in numbers of MRSA bacteremia in the United Kingdom may result in MRSA bacteremia being a minority of infections.

There is stronger but still inconclusive evidence for the use of adjunctive rifampin for prosthetic joint infections (PJJIs), where biofilm assumes a critical importance. For example, Peel et al and Aboltins et al have reported on successful outcomes with debridement and retention of carefully selected patients with PJIs prescribed prolonged courses of rifampin and fusidic acid. However, these studies suffer from their retrospective and observational nature, a limited number of patients with MRSA \(n = 39\), and notably MRSA infection (compared with coagulase negative staphylococci) remained an independent risk factor for treatment failure. The combination of a fluoroquinolone with rifampin has also been demonstrated to be effective in treating selected PJIs with a debridement and retention approach in both a small RCT and several retrospective cohort studies. However, the RCT reported by Zimmerli et al only included 15 PJI patients, included a relatively ineffective control arm (ciprofloxacin monotherapy), and when reanalyzed by intention to treat found no significant difference between the rifampin and nonrifampicin containing arms. Somewhat controversially, based largely on this single RCT, the 2013 IDSA guidelines recommend that rifampin be added to initial parenteral therapy for MRSA PJIs, followed by prolonged combination oral therapy with rifampin with a companion agent such as a fluoroquinolone.

**Daptomycin/Rifampin**

In vitro and animal studies demonstrate an overall pattern of either antagonism or indifference with the addition of rifampin to daptomycin. For example, in an in vitro IE model, the combination of daptomycin with either rifampin or gentamicin antagonized or delayed the bactericidal activity of daptomycin alone. Similarly, in time-kill experiments and a rabbit endocarditis model, the combination of daptomycin and either rifampin or gentamicin demonstrated no enhancement of the effectiveness of daptomycin against MRSA compared with daptomycin alone. There are currently only case reports or small case series of clinical studies involving the combination of daptomycin and rifampin.

**Gentamicin**

**Vancomycin/Gentamicin**

Several studies have demonstrated a consistent in vitro synergism between aminoglycosides and vancomycin. However, clinical studies do not support the addition of an aminoglycoside to vancomycin. In a retrospective evaluation of 87 patients with persistent SAB, 48 of whom had MRSA infection, those treated with an aminoglycoside had lower incidence of recurrence within 6 months, but there was no significant association with mortality or other outcomes. In analyzing data from the daptomycin registrational RCT, Cosgrove et al found that 27/122 (22%) of patients who received initial low-dose gentamicin therapy (in combination with either nafcillin or vancomycin) experienced a clinically significant decline in renal function, compared with 8/100 (8%) of those who did not receive gentamicin. Based on these clinical studies, the IDSA recommends that gentamicin should not be added to vancomycin for the treatment of MRSA bacteremia or native valve endocarditis.

**Daptomycin/Gentamicin**

The combination of daptomycin with gentamicin has been tested in vitro with varying results; synergy has been demonstrated in some studies, but not in others. Unfortunately, a RCT comparing daptomycin to daptomycin combined with gentamicin was terminated early after recruiting only 24 patients (Clinical trials NCT00638157). Thus, the combination of daptomycin with gentamicin cannot be recommended at this stage.

**Other Combinations**

**Daptomycin-nonsusceptible S. aureus** (DNS) not infrequently emerges during daptomycin therapy. Among agents tested in combination with daptomycin, trimethoprim-sulfamethoxazole has shown promise in a PK/PD model for the treatment of DNS. Although clinical experience is currently limited for DNS infections that are refractory to standard treatment, the combination with trimethoprim-sulfamethoxazole should be considered.

In vitro studies have determined that subinhibitory concentrations of clindamycin, linezolid, and rifampin can block production of toxins such as Panton–Valentine leukocidin and \(\alpha\)-toxin by \(S. aureus\). Clinical experience of the use of these agents in severe toxin-mediated staphylococcal infections (e.g., toxic shock syndrome or necrotizing pneumonia) is limited. Two retrospective studies suggest that there may be a clinical benefit for suppression of toxins in such cases. Treatment guidelines from the United Kingdom and France recommend that antitoxin therapy be instituted where toxin-mediated staphylococcal disease is suspected or apparent and in the absence of robust evidence for the treatment of these life-threatening infections, these recommendations are clearly sensible.

Studies of combination therapy for MRSA involving novel antibiotics are also beginning to emerge. In a small number of clinical MRSA isolates tested in vitro (five VISA and five hVISA), oritavancin (a novel lipoglycopeptide antibiotic) appears to be synergistic when used in combination with either gentamicin, linezolid or rifampin, as does telavancin (a second novel lipoglycopeptide antibiotic), when used with gentamicin, ceftriaxone, rifampin, or meropenem. Since only these two drugs became Food and Drug
Administration approved in 2014, and are not approved for treatment of MRSA bacteremia, clinical experience is limited and the implications of these in vitro studies are unclear at this stage. Despite its lack of activity as a single agent against MRSA, fosfomycin appears to be synergistic with linezolid against clinical MRSA isolates in an in vitro model mirroring the β-lactam concept, where a seemingly inactive agent makes an important contribution when combined with an active agent.

**Conclusion**

Because of the limitations of vancomycin, the standard therapy for serious MRSA infections, many combinations of antibiotics have been tested, primarily in in vitro models. Unfortunately, studies of the majority of these combinations have reported mixed or negative data. However, several β-lactam antibiotics have consistently been shown to be synergistic for the majority of MRSA strains (including hVISA and daptomycin nonsusceptible strains), when combined with either vancomycin or daptomycin. Although these combinations appear promising, limited clinical data are available, and clinical trials are only just beginning to be performed. Currently, there is insufficient evidence to recommend any combination therapy for serious MRSA infections in actual patient care.

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