Patient-Activated Three-Dimensional Multifrequency Magnetic Resonance Elastography for High-Resolution Mechanical Imaging of the Liver and Spleen

Patienten-aktivierte dreidimensionale Multifrequenz-MagnetresonanzeLASTAGROFIE zur hochauflösenden mechanischen Bildgebung der Leber und Milz

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Key words

● magnetic resonance elastography
● MR imaging
● abdomen
● spleen
● liver

Zusammenfassung


Ergebnisse: Karten der zwei unabhängigen mechanischen Kenngrößen, Betrag und Phase des komplexen Moduls, wurden in hoher räumlicher Auflösung erzielt. Details der mechanischen Struktur der Leber und der Milz sind gut erkennbar. Die Leber erscheint mit gemittelten Elastizitätswerten von 1,27 ± 0,17 kPa wesentlich weicher als die Milz, für die eine Elastizität von 2,01 ± 0,69 kPa gefunden wurde.

Schlussfolgerung: Patienten-aktivierte 3-D-MMRE kombiniert mit MDEV-Inversion stellt die erste Modalität zur hochauflösenden mechanischen Bildgebung von Leber und Milz dar und verspricht damit genauere MRE-Untersuchungen als bisherige Systeme.

Abstract

Purpose: To introduce a novel in-vivo three-dimensional multifrequency magnetic resonance elastography (3D-MMRE) method for high-resolution mechanical characterization of the liver and spleen.

Materials and Methods: Ten healthy volunteers were examined by abdominal single-shot 3D-MMRE using a novel patient-activated trigger system with respiratory control. 10 contiguous slices with 2.5 mm cubic voxel resolution, 3 wave components, 8 time steps, and 2 averages were acquired at 7 mechanical excitation frequencies from 30 to 60 Hz. The total imaging time was approximately 15 min. For postprocessing, multifrequency dual elasto-visco (MDEV) inversion was used to calculate high-resolution mechanical parameter maps of the abdomen including the liver and spleen.

Results: Two parameters maps were generated from each image slice to capture the magnitude and the phase angle of the complex shear modulus. Both parameters depicted the mechanical structures of the abdomen with unprecedented high spatial resolution. Spatially averaged group mean values of the liver and spleen are 1.27 ± 0.17 kPa and 2.01 ± 0.69 kPa, indicating a significantly higher asymptomatic stiffness of the spleen compared to the liver.

Conclusion: Patient-activated respiratory-gated 3D-MMRE combined with MDEV inversion provides highly resolved mechanical maps of the liver and spleen that are superior to previous elastograms measured by abdominal MRE.

Citation Format:

Introduction

The noninvasive mechanical examination of the liver provides a well-established marker of hepatic fibrosis [1–4]. Techniques of medical ultrasound and magnetic resonance imaging (MRI) have been developed for the measurement of the mechanical response of the liver to mechanical stress [5]. Most of these modalities, subsumed under the term elastography, use static compression [6], externally induced shear waves [7,8] or focused ultrasound pulses [9] to produce a controlled elastic deformation of the tissue.

MRI elastography (MRE) relies on time-harmonic shear waves [10] transmitted into the body from the surface, where they are generated by on-site vibrating coils or by passive transducers attached to a loudspeaker or to a piezoelectric driver [11–14]. To date, MRE is the only modality capable of measuring vector fields of shear waves in soft tissue with isotropic image resolution, which promotes a stable numerical solution of the inverse problem in elastography and therewith ultimately determines the diagnostic precision of elastography [15]. For this reason, the accuracy of MRE potentially outperforms that of other modalities with anisotropic motion estimation.

However, MRE still suffers from uncertainty regions in elastograms, which have to be masked in order to avoid biased stiffness estimates [16,17]. Such artifacts are mainly due to the strong impact of boundaries and tissue interfaces on the appearance of wave fields including non-evanescent waves, wave nodes, and amplitude modulations deviating from ideal wave models implied by the majority of inversion routines. To reduce such wave voids, multifrequency inversion was recently introduced, which adds wave fields of different drive frequencies to the solution of an over-determined system of wave equations [18]. As a result, the accuracy of elastograms can be greatly improved, depending on the amount of information combined into a single inversion step [14]. In this approach, the major limiting factor for improving the quality of elastograms is the duration of image acquisition. For example, single-shot MRE requires repetitive scans for each image slice, wave component, time increment, drive frequency, and, if required, signal averages [19]. Abdominal applications are particularly challenging as they require additional respiratory gating.

Previous hepatic MRE studies were run in 3D at single drive frequency [20–22] or at multiple frequencies in 2D [13,23] with voxel volumes of 64 mm³ or larger. Recently, we introduced 3D multifrequency MRE (3D-MMRE) with four vibration frequencies and an image resolution of 2.7 x 2.7 x 5 mm³ for better resolved and less biased MRE of the liver and spleen [14]. In order to further improve the resolution of abdominal MRE, the quality of the wave images has to be increased (for improved detail resolution of anatomical structures) and more vibration frequencies should be applied (in order to stabilize the inversion by independent wave fields) [18]. To achieve this in a clinically feasible protocol, this study introduces a patient-activated respiratory gating system for abdominal 3D-MMRE. Using this novel gating method, the patient can actively control the time intervals for image acquisition and breathing, which effectively reduces the total examination time and enables us to acquire dynamic wave field data in 10 slices at a resolution of 2.5 x 2.5 x 2.5 mm³ and at 7 vibration frequencies within a clinically acceptable time frame.

We use this enhanced elastodynamic information to produce mechanical parameter maps which display details of the abdomen including the liver and the spleen with fewer artifacts than previous methods. This methodological improvement is intended to provide the methodological background for a precise mechanical characterization of abdominal tissue in different clinical applications such as the assessment of lesions, staging of heterogeneous hepatic fibrosis, or detection of a vascular pressure imbalance, which may influence the elasticity ratio between the liver and spleen [24].

Methods

The study was approved by the institutional review board. Ten healthy volunteers (2 females, mean age, 34.5 ± 8.7 years) were investigated using a clinical protocol including T1-, T2-, and proton-density-weighted pulse sequences. Furthermore, 3D-MMRE was applied as described in the following.

3D-MMRE

A custom-designed nonmagnetic driver based on piezoelectric ceramics as proposed by [25] was mounted at the end of the patient table (Fig. 1). The vibrations were transferred from the tip of the lever to the patient by a carbon fiber rod connected at its distal end to a rubber mat. The transducer mat was moderately pressed onto the body surface in the position just below the right costal arch using Velcro strips attached to the patient table [14]. All experiments were conducted on a 1.5-T MRI scanner (Magnetom Sonata; Siemens Erlangen, Germany) equipped with a 12-channel phased-array surface coil. A 2D single-shot spin-echo echo-planar imaging (EPI) sequence with a trapezoidal flow-compensated motion-encoding gradient (MEG), consecutively applied along all three axes of the scanner coordinate system, was used for rapid motion field acquisition. The motion was initiated by a trigger pulse from the scanner at least 100 ms before the start of the MEG to allow the waves to propagate into the tissue. Eight dynamics of a wave cycle were captured by delaying the trigger pulse in increments of 1/(8f) with f being the vibration frequency. The vibration frequencies were 30, 35, 40, 45, 50, 55, and 60 Hz. For each drive frequency, MEG direction, and wave dynamic, 10 adjacent transverse image slices with a resolution of 2.5 x 2.5 x 2.5 mm³ were recorded, yielding a total of 1680 images. For each image, the signal was averaged twice to improve the signal-to-noise ratio (SNR). Further imaging parameters were: repetition time (TR), 3790 ms; echo time (TE), 55 ms; field of view (FoV), 320 x 260 mm²; matrix size, 128 x 104; parallel imaging with a GRAPPA of factor 2; MEG frequency, 50 Hz (implying fractional motion-encoding for all drive frequencies ± 50 Hz [26] and providing a feasible compromise between motion sensitivity and short TE); MEG amplitude, 30 mT/m; net scan time (without respiratory gating) for each frequency, 1.5 min. The total examination time (for 7 frequencies) was approximately 13 min, with variations depending on the voluntarily adjusted breathing intervals.

Patient-activated respiratory gating

Each volunteer was asked to maintain a reproducible position of the chest and abdomen in expiration through the patient-activated 3D-MMRE acquisition. When activated, the trigger delayed the processing of wave image acquisition after accomplishing an entire TR-interval. Therefore, the halt of the sequence did not interfere with the k-space acquisition of single images, and the respiratory pauses could be introduced anytime during the acquisition of full wave images of different vibration phases or field components.

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For activating the trigger, a pneumatic ball was connected to a pressure-sensitive button which produced a continuous 4.5-V gating signal fed into a signal generator outside the scanner room. The signal generator produced a comb of equidistantly spaced rectangular TTL pulses with a frequency of 100 Hz and a 50% duty cycle. Thus, upon activation, the pneumatic ball shown in Fig. 1 generated the TTL-comb-shaped signal for starting the externally triggered 3D-MMRE sequence. The signal pathway is schematically shown in Fig. 1.

Data processing

In the first step of postprocessing, derivative-based unwrapping was performed. This method is a useful tool for treating phase wraps if the derivative of the displacement field is further considered (in our case when applying the curl-operator). The method was proposed several years ago in reference [27] and has been used since then as a standard procedure in MRE. First-order spatial derivatives of the m-th component of spin phase \( \varphi_m \) were calculated using the following equation:

\[
\frac{\partial \varphi_m(r, \omega_n)}{\partial x_k} = -i \cdot \exp(-i \varphi_m(r, \omega_n)) \cdot \frac{\partial}{\partial x_k} \exp(i \varphi_m(r, \omega_n))
\]

(1)

where \( \varphi_m(r, \omega_n) \) denotes the phase measured at n-th external angular drive frequencies and position \( r \). \( \varphi_m \) is wrapped (\( \varphi_m \in [0, 2\pi] \)), while its derivative is wrap-free and unbiased if the phase difference of two adjacent voxels does not exceed \( \pi \). \( r \) denotes the position vector and the n-th angular drive frequency. \( \partial \varphi_m / \partial x_k \) was scaled to \( \partial u_m / \partial x_k \) (i.e., spatial derivatives of the physical displacement field \( u \)) by the factor given in equation (4b) of [28].

For reducing noise, numerical derivatives were calculated by 3D gradients according to Anderssen and Hegland [29] using a two-pixel symmetric window in three dimensions [30]. The resulting strain components of the wave field (derivatives of the three Cartesian components of the displacement field) were used to calculate the curl field, as has been established in 3D MRE for suppressing volumetric strain [31]. The curl components \( C_1, C_2 \) and \( C_3 \) are defined as follows:

\[
C_1 = \frac{\partial u_2}{\partial x_3} - \frac{\partial u_3}{\partial x_2}, C_2 = \frac{\partial u_3}{\partial x_1} - \frac{\partial u_1}{\partial x_3}, C_3 = \frac{\partial u_1}{\partial x_2} - \frac{\partial u_2}{\partial x_1}
\]

(2)
where \( u_m(r, t) \) is the \( m \)-th component of the wave field with \( m = 1, 2, \) and \( 3 \) referring to the imaging gradient directions of read-out, phase-encoding, and slice selection, respectively. The curl components of equation (2) were smoothed by a 3D Gaussian kernel with an edge length of 3 pixels and were Fourier-transformed along the time axis (eight samples over one vibration cycle). The resulting complex 3D wave images, corresponding to the first harmonic \( n \)-th vibration frequency (\( \omega_n \)), were taken as components \( u'_n(r, \omega_n) \) of the wave field for multifrequency dual elastivisco (MDEV) parameter recovery as described in detail in [14]. In MDEV inversion, the complex-valued shear modulus \( \mathcal{G} \) is represented by its magnitude \( |\mathcal{G}| \) and phase angle \( \phi \). Both parameters are reconstructed separately using a modified Helmholtz inversion as briefly summarized in the appendix.

**Results**

The new patient-activated gating mechanism was easily handled by all volunteers and was reported to support better reproduction of the breath-hold position than conventional breathing commands.

**Fig. 2** portrays a set of complex waves \( u'_n = \text{Re}\{ u_m(r, \omega_n) \} \) obtained in a central slice of a volunteer at 7 vibration frequencies. The images were derived from the raw phase signal by gradient unwrapping (equation (1)), curl calculation (equation (2)), and temporal Fourier transformation. Application of the curl operator suppresses compression wave artifacts. The wave lengths decrease with increasing vibration frequency. It is well visible that the appearance of the wave patterns depends on the field component and frequency, which corroborates our approach of compensating for low wave amplitudes due to wave nodes (e.g., demarcated by the arrow in \( u_1 \) at 30 Hz) or attenuation (e.g., at 60 Hz) by combining all images in the inversions of equations (5) and (6).

**Fig. 3** highlights the appearance of MDEV parameter maps in a transverse slice of five volunteers. We arbitrarily chose 5 volunteers including the ones with the smallest and largest liver in order to cover a wide range of physiological variations in hepatic morphology. Other elastograms acquired in this study appear similar with regard to anatomical features and resolution. In contrast to the classic display of elastograms in MRE [12, 13, 32], the maps of mechanical parameters in **Fig. 3** are not masked for excluding noise or regions of unreliable reconstructed parameters. Instead, the full mechanical image can be evaluated including the liver and spleen. Visible heterogeneities are due to the blood vessel in the liver. The spleen appears to be stiffer and more heterogeneous than the liver as indicated by group mean values of \( |\mathcal{G}| = 1.27 \pm 0.17 \) kPa and \( |\mathcal{G}| = 2.01 \pm 0.69 \) kPa for hepatic and splenic stiffness and SD = 0.35 ± 0.08 kPa and SD = 0.69 ± 0.34 kPa for intrahepatic and intrasplenic heterogeneity (**Table 1**). Stiffness and heterogeneity are both significantly different with \( p \)-values of 0.0041 and 0.0066, respectively. Similar statistically significant differences were found for \( \phi \), which was higher in the spleen (0.607 ± 0.152) than in the liver (0.407 ± 0.044, \( p = 0.0008 \)), again with a higher intrasplenic heterogeneity (SD = 0.204 ± 0.034 [liver] vs. 0.282 ± 0.076 [spleen], \( p = 0.0084 \)). Similar to previous studies [24, 33], we found a significant correlation between \( |\mathcal{G}| \) of the liver and spleen (\( p = 0.046 \)), while \( \phi \) was not correlated in the two organs, as has been reported in [14].

**Discussion**

High-resolution mechanical imaging remains a major goal of method development in elastography. Our study contributes to this development by introducing a new respiratory gating method that enables us to acquire more wave fields at different frequencies in the abdomen than was feasible with earlier techniques (7 frequencies, 2 averages compared to 4 frequencies, no average in Ref. [14]). This multidimensional information was combined into two maps of viscoelastic parameters which are related to the elasticity and viscosity of the tissue. Our previous studies using MMRE in the liver [23, 34, 35] have shown that the liver is a highly viscous organ giving rise to signif-
significant dispersion of the shear modulus values within the applied range of drive frequencies (25 Hz to 62.5 Hz). Notably, MDEV-inversion used here sacrifices the dispersion-related information in order to stabilize the inversion and improve the certainty and precision of elastograms (i.e. \( |G^*| \) and \( \phi \) maps). This approach is fundamentally different from previous work in abdominal MRE where consistent shear modulus values were obtained by spatially averaging \( G^* \) values within larger regions of the liver. At the moment, ‘high-resolution’ MRE cannot provide information about the frequency dependence of the measured mechanical parameters. However, further work is warranted to address this issue.

\( |G^*| \) reflects the stiffness or softness of the tissue and is closely related to the shear wave speed measured by transient stimulation methods in ultrasound elastography. Our group mean value of \( |G^*| = 1.27 \pm 0.17 \text{ kPa} \) for the liver falls in the range of shear modulus values (1.1 to 1.8 kPa) measured by Fibroscan [36, 37] or ARFI [37, 38] ultrasound devices. The phase angle of the complex modulus \( \phi = \arctan(\text{Im}(G^*)/\text{Re}(G^*)) \) is a relatively unexplored parameter within the field of elastography. \( \phi \) is potentially sensitive to the geometry of the viscoelastic lattice [39].

### Table 1

<table>
<thead>
<tr>
<th>vol. #</th>
<th>sex</th>
<th>age (years)</th>
<th>liver</th>
<th>spleen</th>
<th>liver</th>
<th>spleen</th>
<th>liver</th>
<th>spleen</th>
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<tr>
<td>1</td>
<td>m</td>
<td>43</td>
<td>1.23</td>
<td>0.25</td>
<td>2.60</td>
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<td>0.172</td>
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<td>2</td>
<td>m</td>
<td>27</td>
<td>1.41</td>
<td>0.43</td>
<td>2.84</td>
<td>0.76</td>
<td>0.397</td>
<td>0.229</td>
</tr>
<tr>
<td>3</td>
<td>m</td>
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<td>1.47</td>
<td>0.39</td>
<td>1.52</td>
<td>0.58</td>
<td>0.450</td>
<td>0.291</td>
</tr>
<tr>
<td>4</td>
<td>m</td>
<td>30</td>
<td>1.30</td>
<td>0.41</td>
<td>1.99</td>
<td>0.51</td>
<td>0.411</td>
<td>0.198</td>
</tr>
<tr>
<td>5</td>
<td>m</td>
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<td>1.10</td>
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<td>0.188</td>
</tr>
<tr>
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<td>m</td>
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<td>1.20</td>
<td>0.35</td>
<td>1.50</td>
<td>0.44</td>
<td>0.486</td>
<td>0.189</td>
</tr>
<tr>
<td>7</td>
<td>m</td>
<td>41</td>
<td>1.03</td>
<td>0.28</td>
<td>0.87</td>
<td>0.25</td>
<td>0.465</td>
<td>0.187</td>
</tr>
<tr>
<td>8</td>
<td>f</td>
<td>29</td>
<td>1.55</td>
<td>0.48</td>
<td>3.16</td>
<td>1.40</td>
<td>0.357</td>
<td>0.206</td>
</tr>
<tr>
<td>9</td>
<td>m</td>
<td>29</td>
<td>1.18</td>
<td>0.31</td>
<td>1.58</td>
<td>0.48</td>
<td>0.375</td>
<td>0.203</td>
</tr>
<tr>
<td>10</td>
<td>f</td>
<td>27</td>
<td>1.18</td>
<td>0.37</td>
<td>1.95</td>
<td>0.71</td>
<td>0.380</td>
<td>0.179</td>
</tr>
<tr>
<td>Mean</td>
<td></td>
<td>34.50</td>
<td>1.27</td>
<td>0.35</td>
<td>2.01</td>
<td>0.69</td>
<td>0.407</td>
<td>0.204</td>
</tr>
<tr>
<td>SD*</td>
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<td>8.68</td>
<td>0.17</td>
<td>0.08</td>
<td>0.69</td>
<td>0.34</td>
<td>0.044</td>
<td>0.034</td>
</tr>
</tbody>
</table>

SD* denotes the intersubject standard deviation.

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surprising that $\varphi$ is higher in the spleen than in the liver, given the fact that nearly 80% of the spleen is made up of red pulp, which consists of fibrils and connective tissue cells with a strongly cross-linked mechanical structure. However, the interpretation of $\varphi$ as a loss tangent implies the validity of the so-called spring-pot model [35], which we ignore in the recovery of $G'$. Using equation (6). MDEV inversion does not account for viscoelastic models but provides numerically stable solutions of two independent parameters related to the elastic and viscous behavior of the tissue. Similar to the storage modulus $Re(G)$, we expect $G'$ to be sensitive to fibrosis [13], whereas the significance of $\varphi$ as a diagnostic marker remains to be determined.

Our feasibility study in a group of ten healthy volunteers paves the road for further in-depth evaluation of mechanical imaging in the abdomen. Potential applications of high clinical relevance include the kidney [40], prostate [41, 42], and cervix [43, 44]. A further perspective of abdominal 3D-MMRE is the noninvasive assessment of tissue pressure by evaluating volumetric strain [19] or poroelastic parameters [45]. Applications in patients are warranted for the ultimate determination of the clinical value of the proposed respiratory gating method.

A limitation of the respiratory gating technique presented here is the need for patient cooperation. Patients who are not able to follow instructions have to be excluded from the examination. However, a similar problem arises in all MRI examinations involving actively controlled breathing. Therefore, our active gating approach in 3D-MMRE may be replaced in the future by passive respiratory gating such as navigator-based gating. At any rate, patient-activated 3D-MMRE, as proposed in this study, represents an advance toward the goal of solving the gating problem in abdominal MRE.

In summary, we have introduced a clinically feasible respiratory gating method for abdominal MRE providing highly resolved 3D wave fields at multiple harmonic vibration frequencies for the further improvement of mechanical parameter maps. Respiratory gating was achieved by a patient-activated pneumatic trigger. Using this novel method, we were able to acquire an unprecedented amount of information of in vivo abdominal shear waves in 10 transverse slices of 2.5 mm isotropic resolution with 3 field components, 7 vibration frequencies, and 8 time steps. We used this information to generate mechanical parameter maps of the magnitude and the phase of the complex shear modulus capable of displaying the heterogeneity of the in vivo shear modulus in abdominal organs. The significance of the enhanced MRE contrast for diagnostic applications such as the noninvasive staging of hepatic fibrosis or tumor detection remains to be determined.

### Appendix

#### MDEV inversion

Dual elasto-visco inversion relies on two independent solutions of different representations of the Helmholtz wave equation. We start with representing complex shear modulus $G'$ by its magnitude $|G'|$ and phase $\varphi$:

$$G' = |G'| \exp(i \varphi).$$

(3)

Corresponding to this representation, the Helmholtz equation can be written in terms of $|G'|$ and $\varphi$ by

$$|G'| \left[ \begin{array}{l} \cos \varphi - \sin \varphi \\ \sin \varphi \cos \varphi \end{array} \right] \Delta x_m = -\rho c^2 x_m \text{ with } -x_m = \left[ u_m'(\omega_n) \right].$$

(4)

with $\Delta$ denoting the 3D-Laplacian operator and material density $\rho$ (assumed to be 1.1 g/cm$^3$ in the following [46]). $u_m(\omega_n)$ and $u_m'(\omega_n)$ denote the real and imaginary part of $u_m(r, \omega_n)$, respectively, acquired at the vibration frequency $\omega_n$. As demonstrated in [18], equation (4) can be solved by

$$\varphi = \arccos \left( \frac{1}{\rho c^2} \sum_{n=m}^{N} \sum_{m=1}^{m} \left| \Delta u_m'(\omega_n) L_{nm}(\omega_n) \right| \right).$$

(5)

A similar least-squares solution is obtained for $G'$ by considering the magnitude of the Helmholtz equation $|G'| \Delta u_m'(r, \omega_n) = \rho c^2 u_m(r, \omega_n)$

$$|G'| = \rho \left| \frac{1}{\sum_{n=m}^{N} \sum_{m=1}^{m} \Delta u_m'(\omega_n) L_{nm}(\omega_n)} \right|$$

(6)

### Acknowledgement

This manuscript is dedicated to Professor Bernd Hamm for his 60th birthday.

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