Correlation of the Apparent Diffusion Coefficient (ADC) with the Standardized Uptake Value (SUV) in Hybrid 18F-FDG PET/MRI in Non-Small Cell Lung Cancer (NSCLC) Lesions: Initial Results

Korrelation des scheinbaren Diffusionskoeffizienten (ADC) mit dem „standardized uptake values“ (SUV) bei nichtkleinzelligen Bronchialkarzinomen (NSCLC) in einem hybriden 18F-FDG-PET/MR

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Zusammenfassung

Ziel: Vergleich der in der kombinierten 18F-FDG-PET/MR bzw. 18F-FDG-PET/CT ermittelten, scheinbaren Diffusionskoeffizienten (ADC) und der „standardized uptake values“ (SUV) bei nichtkleinzelligen Bronchialkarzinomen (NSCLC).


Ergebnisse: Der SUV max der NSCLC betrug 12,3 ± 4,8 [Mittelwert ± Standardabweichung], der SUVmean betrug 7,2 ± 2,8, gemessen in der FDG-PET/MR. Die mit Hilfe der FDG-PET/CT und FDG-PET/MR gemessenen SUVmax- und SUVmean-Werte korrelierten jeweils sehr gut (R = 0,93; p < 0,001 und R = 0,92; p < 0,001). Der mittlere ADC (ADCmean) der Lungentumore war 187,9 ± 88,8 × 10⁻⁵ mm²/s [Mittelwert ± Standardabweichung]. Der mit Hilfe der FDG-PET/MR gemessene ADCmean wies eine signifikante, inverse Korrelation mit dem SUVmax (R = -0,72; p < 0,001) als auch mit dem SUVmean (R = -0,71; p < 0,001) auf.

Schlussfolgerung: Es besteht sowohl eine exzellente Korrelation zwischen dem SUVmax und SUVmean ermittelt in der FDG-PET/CT und der darauf folgenden FDG-PET/MR, als auch eine signifikante, inverse Korrelation zwischen dem

Abstract

Purpose: To compare the apparent diffusion coefficient (ADC) in non-small cell lung cancer lesions with standardized uptake values (SUV) derived from combined 18F-fluoro-deoxy-glucose-positron emission tomography/magnetic resonance imaging (FDG-PET/MRI) and those derived from FDG-PET/CT.

Materials and Methods: In 18 consecutive patients with histologically proven NSCLC (17 men, 1 woman; mean age, 61 ± 12 years), whole-body FDG-PET/MRI was performed after whole-body FDG-PET/CT. Regions of interest (ROI) encompassing the entire primary tumor were drawn into FDG-PET/CT and FDG-PET/MRI images to determine the maximum and mean standardized uptake value (SUVmax; SUVmean) and into ADC parameter maps to assess mean ADC values. Pearson’s correlation coefficients were calculated to compare SUV and ADC values.

Results: The SUVmax of NSCLC was 12.3 ± 4.8 [mean ±SD], and the SUVmean was 7.2 ± 2.8 as assessed by FDG-PET/MRI. The SUVmax and SUVmean derived from FDG-PET/CT and FDG-PET/MRI correlated well (R = 0.93; p < 0.001 and R = 0.92; p < 0.001, respectively). The ADCmean of the pulmonary tumors was 187.9 ± 88.8 × 10⁻⁵ mm²/s [mean ± SD]. The ADCmean exhibited a significant inverse correlation with the SUVmax (R = -0.72; p < 0.001) as well as with the SUVmean assessed by FDG-PET/MRI (R = -0.71; p < 0.001).

Conclusion: This simultaneous PET/MRI study corroborates the assumed significant inverse correlation between increased metabolic activity on FDG-PET and restricted diffusion on DWI in NSCLC.

Citation Format:

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SUV\text{\textsubscript{max}} dem SUV\text{\textsubscript{mean}} und dem ADC\text{\textsubscript{mean}} ermittelt mit Hilfe der FDG-PET/MR.

**Introduction**

Lung cancer is the most frequently diagnosed cancer with one of the highest cancer mortalities globally [1]. Non-small cell lung cancer (NSCLC) accounts for approximately 80% of all bronchogenic malignancies [1]. For a stage-adapted therapy and thus an individual therapeutic concept, 18F-fluorodeoxyglucose-pet tracer uptake value (SUV\text{\textsubscript{max}}) of NSCLC assessed by FDG-PET/CT is established as an independent marker of tumor characteristics [3]. Apart from PET/CT, magnetic resonance imaging (MRI) offers great potential in the detection and interpretation of malignancies [4]. Especially diffusion-weighted MR imaging (DWI) is a sensitive tool for lesion detection (primary tumors, nodal and distant metastases) in various malignancies and yields comparable results to FDG-PET/CT in some tumor subtypes [5-7]. Due to the bright appearance of malignant lesions on high-b-value DWI images, which provide a high lesion-to-background ratio, DWI commonly serves as a “search-sequence” with a PET-like appearance. Therefore, DWI is included in whole-body MRI staging protocols in many centers. DWI may also increase the specificity when it comes to lesion interpretation by quantification of the apparent diffusion coefficient (ADC). In malignant lesions the increased cellular density restricts water diffusion in the interstitial space, resulting in lower ADC values than in inflammatory reactive tissue or scar tissue [8, 9]. In a previous study the potential of DWI in the differentiation of malignant from benign pulmonary lesions has been demonstrated [10]. Matoba et al. reported a significant correlation between ADC values and the degree of cell differentiation in patients with lung cancer [11].

Recently introduced whole-body integrated PET/MRI scanners enable truly simultaneous acquisition and highly accurate spatial co-registration of PET and MRI data sets [12, 13]. Apart from and in addition to the lack of ionizing radiation as compared with CT, MRI in combination with PET is expected to provide a new quality in functional cancer imaging [14], mainly due to the combination of functional MR and PET information. Already published clinical FDG-PET/CT (60 min after injection of 300 ± 40 MBq 18F-FDG). All patient histories revealed no prior malignant disease and no chemotherapy or radiotherapy. Written informed consent was obtained from all participants. This study was performed in accordance with the regulations of the local institutional ethics committee.

**PET/CT Imaging**

Whole-body (WB) FDG-PET/CT scans were obtained on a mCT™ PET/CT scanner (Siemens Molecular Imaging, Hoffmann, Estates, IL, USA), containing 32.440 4 × 4 mm LSO crystals for PET imaging. Before imaging, patients fasted for at least 6 h. All patients had blood glucose levels below 150 mg/dL at the time of 18F-FDG injection. 18F-FDG (300 ± 40 MBq) was intravenously injected 60 min before the scan. The contrast-enhanced CT scan was taken with the following parameters: caudocranial scan direction, field of view: skull base to upper thighs, 120 kV, automatic mA/s adjustment (Care Dose 4D™, preset: 210 mAs), 5 mm slice thickness, 5 mm increment, pitch 1. PET scan: 3D mode, 2 min emission time per bed position (45% overlap), reconstruction according to the ordered subsets expectation maximization (OSEM) algorithm with 4 iterations and 8 subsets, 3D Gaussian filter, 4.0 mm, full width at half maximum (FWHM), scatter correction. The attenuation correction was based on the portal venous phase of whole-body CT.

**PET/MR Imaging**

WB FDG-PET/MRI was performed on a Magnetom Biograph mMR™ (Siemens Healthcare, Erlangen, Germany), containing a total of 28.672 LSO crystals for PET-measurement. FDG-PET/MRI was performed following FDG-PET/CT with a mean delay of 80 ± 13 min. The field of view (FoV) contained the body volume from the head to the thighs. The PET acquisition time was 20 min for the entire thorax. PET images were reconstructed using the iterative algorithm OSEM, 3 iterations and 21 subsets, Gaussian filter: FWHM 4.0 mm; scatter correction. A dedicated mMR head and neck coil and, depending on the patients’ height, up to 4 mMR body flex coils were used for MR imaging. MR imaging was performed simultaneously to PET imaging using the following sequence protocol for each bed position:

1. a coronal 3-dimensional volume interpolated gradient echo (VIBE) sequence (repetition time (TR) 3.6 ms, echo time 1 (TE1) 1.23 ms, TE2 2.46 ms, 3.12 mm slice thickness, FoV 500 mm for DIXON-based attenuation correction);  
2. a coronal T2-weighted steady state free precession (TrueFISP) sequence of the thorax (TR 3.75 ms, TE 1.64 ms, matrix size 5 mm increment, pitch 1. PET scan: 3D mode, 2 min emission time per bed position (45% overlap), reconstruction according to the ordered subsets expectation maximization (OSEM) algorithm with 4 iterations and 8 subsets, 3D Gaussian filter, 4.0 mm, full width at half maximum (FWHM), scatter correction. The attenuation correction was based on the portal venous phase of whole-body CT.

**Materials and Methods**

**Patients**

In 18 consecutive patients with histologically proven NSCLC (17 men, 1 woman; age 61 ± 12 years), whole-body FDG-PET/MRI was performed (no additional 18F-FDG injection) after routine clinical FDG-PET/CT (60 min after injection of 300 ± 40 MBq 18F-FDG). All patient histories revealed no prior malignant disease and no chemotherapy or radiotherapy. Written informed consent was obtained from all participants. This study was performed in accordance with the regulations of the local institutional ethics committee.

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320, 6 mm slices, FoV 330 mm, generalized auto calibrating partially parallel acquisition (GRAPPA); acceleration factor 2); 3. a transverse T2-weighted blade turbo spin echo sequence of the thorax in breath-hold technique (TR 4360 ms, TE 160 ms, matrix size 384, 5 mm slices, FoV 400 mm, GRAPPA, acceleration factor 2); 4. whole-body transverse echo planar imaging (EPI) DWI (TR 10 500 ms, TE 78 ms, diffusion weightings (b-values): 0, 500 and 1000 s/mm², matrix size 160, 5 mm slices, 40 slices, FoV 450 mm, GRAPPA, acceleration factor 2; 2 averages); 5. a transverse T1-weighted fast low angle shot (FLASH) gradient echo sequence (TR 1510 ms, TE 2.15 ms, inversion time (TI) 1200 ms, matrix size 320, 5 mm slices, FoV 400 mm, GRAPPA, acceleration factor 2); 6. a coronal half Fourier acquisition single shot turbo spin echo sequence (HASTE) of the thorax (TR 649 ms, TE 51 ms, matrix size 320, 6 mm slices, FoV 330 mm, GRAPPA, acceleration factor 2); 7. a coronal 3-dimensional VIBE sequence of the thorax (TR 3.66 ms, TE 1.29 ms, matrix size 192, 4 mm slice thickness, FoV 350 mm, GRAPPA, acceleration factor 2) 8. a coronal 3-dimensional VIBE sequence of the thorax after i. v. administration of gadolinium with a delay of 2 min (TR 3.67 ms, TE 1.29 ms, matrix size 192, 4 mm slice thickness, FoV 350 mm, GRAPPA, acceleration factor 2) 9. a coronal 2D turbo inversion recovery sequence with magnitude (TIRM) with short TI for fat suppression in free-breathing of the whole body (TR 3190 ms, TE 55 ms, matrix size 384, 5 mm slice thickness, FoV 450 mm, GRAPPA, acceleration factor 2) PET/MRI image fusion was performed for the post-contrast T1-weighted VIBE images and the T2-weighted blade images.

**Image Analysis**

Two readers with 8 and 4 years of experience in MRI and 8 and 6 years of experience in hybrid PET/CT imaging analyzed the images in consensus and in random order using a picture archiving and communication system (Centricity; General Electric Medical Systems, Milwaukee, WI, USA) and a dedicated viewing software for hybrid imaging (Syngo.via; Siemens, Healthcare Sector, Erlangen, Germany). Tumor borders of the primary lung cancer were identified on axial slices of the CT scan or T1w images and a polygonal volume of interest (VOI) was placed on fused PET/CT or PET-MR images (Fig. 1), covering the entire tumor manifestation to determine the mean and maximum standardized uptake value (SUVmax; SUVmean).

For ADC analysis, DWI data were transferred to an external workstation equipped with the custom software STROKETOOL (http://www.digitalimagesolutions.de). ADC parameter maps were computed using the following formula:

\[ S(b) = S_0 e^{-\frac{b}{ADC_{mono}}} \]

with \( S(b) \) = signal intensity at b-value \( b \), \( S_0 \) = signal acquired without diffusion sensitizing gradients and \( ADC_{mono} \) = diffusion coefficient. For further analysis, a polygonal region of interest (ROI) was manually drawn on \( b = 0 \) images encompassing the entire area of the targeted lesions by one author with 8 years of experience in body MR imaging and then transferred to the corresponding parameter maps (Fig. 2) using the STROKETOOL software.

In order to ensure proper positioning of the ROI on the \( b = 0 \) images, the contrast-enhanced T1-weighted images were reviewed during the placement of the ROI. ADCmean values were determined for all lung tumors. The mean ADC was defined as the mean value of all tumor pixels in the assessed ADC slices.
Fig. 2  Increased signal intensity on the b1000 DWI image of a NSCLC located in the right upper lobe a and with the tumor appearing as an area of low signal intensity on the corresponding ADC map b.

Abb. 2  Gesteigerte Signalintensität eines NSCLC im rechten Oberlappen in der b1000 DWI a. Der Tumor erscheint signalabgesenkt in der korrespondierenden ADC Karte b.

Fig. 3  Scatter plot showing a significant inverse correlation between the ADCmean and the SUVmax (R = –0.72; p < 0.001).

Abb. 3  Das Streudiagramm zeigt eine signifikante, inverse Korrelation zwischen dem mittleren ADC und dem SUVmax (R = –0.72; p < 0.001).

Fig. 4  Scatter plot showing a significant inverse correlation between the ADCmean and the SUVmean (R = –0.71; p < 0.001).

Abb. 4  Das Streudiagramm zeigt eine signifikante, inverse Korrelation zwischen dem mittleren ADC und dem SUVmean (R = –0.71; p < 0.001).
Tumor histopathology was confirmed with percutaneous needle biopsy, endobronchial ultrasound biopsy, or thoracotomy in the 18 patients. Nine patients had adenocarcinomas, seven patients had squamous cell carcinomas, and two patients had large cell carcinomas.

**Statistics**

Statistical analysis was performed using SPSS 20™ (SPSS Inc., Chicago, IL, USA). Data are presented as means ± standard deviation (SD). Descriptive analysis was used for SUV<sub>max</sub> and SUV<sub>mean</sub> of the tumor tissue. Pearson’s correlation coefficients were calculated to compare SUV and ADC values. A p-value ≤ 0.05 was considered to indicate statistical significance.

**Results**

FDG-PET/CT and FDG-PET/MRI acquisitions were completed successfully in all 18 patients. The mean delay between FDG-PET/CT and FDG-PET/MRI was 80 ± 13 min.

The SUV<sub>max</sub> of NSCLCs was 12.3 ± 4.8, and the SUV<sub>mean</sub> was 7.2 ± 2.8 assessed by FDG-PET/MRI. For FDG-PET/CT the SUV<sub>max</sub> of NSCLC was 12.7 ± 4.6, and the SUV<sub>mean</sub> was 7.5 ± 2.7. The SUV<sub>max</sub> measurements derived from FDG-PET/CT and from FDG-PET/MRI exhibited an excellent correlation (R = 0.93; p < 0.001). Also, the SUV<sub>mean</sub> for FDG-PET/CT and for FDG-PET/MRI had an excellent correlation (R = 0.92; p < 0.001).

The tumor ADC<sub>mean</sub> for all patients was 187.9 ± 88.8 × 10<sup>-5</sup> mm<sup>2</sup>/s and exhibited a significant inverse correlation with the SUV<sub>max</sub> (R = −0.72; p < 0.001) (Fig. 3) as well as with the SUV<sub>mean</sub> assessed by FDG-PET/MRI (R = −0.71; p < 0.001) (Fig. 4). Furthermore, the ADC<sub>mean</sub> exhibited a significant inverse correlation with the SUV<sub>max</sub> (R = −0.76; p < 0.001) as well as with the SUV<sub>mean</sub> assessed by FDG-PET/CT (R = −0.77 p < 0.001).

**Discussion**

For NSCLC a significant correlation between ADC values and SUV values derived from FDG-PET/CT has been reported [18]. Whole-body integrated PET/MRI scanners provide diagnostic image quality and comparably good lesion characterization and tumor staging in pulmonary masses as compared with PET/CT [16]. In this study we were able to demonstrate a significant inverse correlation of the SUV and ADC values assessed with simultaneous PET/MRI examination.

MR imaging combined with PET requires an accurate correction of detected annihilation quanta for the attenuation effect caused by different body tissues. The major difficulty of MRAC lies in the fact that the MR signal is not related to the radiodensity of the examined tissue and therefore cannot directly be used for attenuation correction. A discrepancy in MRAC and CTAC may result in differences in quantitative SUV measurements, potentially leading to a misinterpretation of the PET-MR data. However, SUV measurements were reported to be robust and provide a high reproducibility using 18F-FDG-PET/CT in NSCLC lesions [20]. In oncologic patients examined with PET/CT and PET/MRI, the SUV<sub>max</sub> and SUV<sub>mean</sub> values generally correlate well in normal organ tissues [21]. Furthermore, SUV measurements revealed a high correlation between mean SUVs measured with PET/MRI and PET/CT in lesions [15]. Concordantly, there was a significant correlation in this study between the SUV<sub>max</sub> and SUV<sub>mean</sub> of the NSCLCs when assessed by a simultaneous PET/MRI and PET/CT for NSCLC patients. Hybrid PET/MRI is often performed following clinically indicated PET/CT, leading to a longer uptake time from tracer injection in PET/MRI scans. This delay potentially causes different absolute organ distribution and physiological washout of 18F-FDG. In the present study we expected higher SUVs in PET/MRI due to tracer accumulation in malignant tissue. Surprisingly we experienced the opposite. One potential explanation is a difference regarding the type of PET attenuation correction. However, our findings are concordant with recently published PET/MRI studies by Drzezga et al. and Wiesmüller et al. [15, 22]. The potential of FDG-PET/CT for staging in patients with diagnosed NSCLC has been demonstrated [2, 23]. Based on SUV changes, 18F-FDG uptake can determine patient prognoses after treatment [24, 25] and predict tumor response [26]. On the other hand, DWI also has an established role in tumor detection [11] and response assessment. Ohno et al. evaluated the prognostic value of the ADC in NSCLC patients undergoing chemoradiotherapy, suggesting that DWI measurements may be superior to FDG-PET/CT in the prediction of tumor response [27]. Image acquisition significantly differs between DWI and PET as DWI yields information on motion of water molecules representing tissue cellularity, whereas PET measurements yield information about tissue metabolism. However, FDG uptake correlates with the number of viable tumor cells, thus high cellularity may also lead to an increase in FDG uptake. This may be the explanation for the relationship between both aspects (DWI and metabolism) reported in previous studies [25]. Our study demonstrated an inverse correlation between FDG uptake and ADC supporting the theory of increasing FDG uptake and decreasing ADC with increasing cellularity.

While in B-cell lymphomas and invasive ductal carcinomas no correlation was found between ADC and SUV values on PET/CT [28, 29], a significant inverse correlation between SUV and ADC values was reported for GIST, rectal cancers and head and neck squamous cell carcinomas [30–32]. This difference may be explained by tumor heterogeneity. For NSCLC, a significant inverse correlation was found between the SUV<sub>max</sub> and ADC<sub>min</sub> whereas no significant correlation was found when comparing the ADC<sub>mean</sub> vs. the SUV<sub>mean</sub> or the ADC<sub>mean</sub> vs. the SUV<sub>max</sub> [18]. In contrast, we were able to demonstrate a significant inverse correlation between the ADC<sub>mean</sub> and the SUV<sub>max</sub> and between the ADC<sub>mean</sub> and the SUV<sub>mean</sub>. This discrepancy may be due to the simultaneous coregistration of the PET and MRI datasets in our study, whereas Regier et al. performed PET/CT measurements within 30 days prior to the MR examination [18]. ROIs were drawn manually by a radiologist and might potentially include parts of the surrounding lung tissue. This might lead to a very low ADC<sub>min</sub> value, if pixels containing healthy lung tissue are erroneously included in the ROI measurement. In contrast, the ADC<sub>mean</sub> is generally accepted as a more reliable indicator of tumor cellularity and aggressiveness since the entire lesion is taken into account. Hence, we did not examine the ADC<sub>min</sub> in our study. It has to been taken into account that measured ADC values strongly depend on the exact choice of b-values, different field strengths and diffusion encoding techniques [33]. In the present study we chose a range of three b-values (b0-b500-b1000), because the whole-body PET/MRI system is foremost designed for and used in oncologic applications. With our choice of b-values, we met the consensus recommendations on the use of DWI as an image biomarker in cancer patients [34]. Furthermore, as
demonstrated previously in human brains, ADC values can be reliably calculated using only 2 b-values [35]. To the best of our knowledge, the robustness and repeatability of DWI has not been investigated in NSCLC lesions so far. Several factors may affect ADC quantification, for example the choice of b-values, tissue perfusion [36] or scanner geometry and field strengths [37]. Therefore, the ADC values reported for different cancer types might exhibit significant variations. However, looking at the reproducibility of ADC measurements, recent studies in transplanted kidneys and liver metastases of colorectal carcinoma have shown a high reproducibility with a coefficient of variation below 10 % [38, 39]. Up to now, it is unclear if this high reproducibility is transferable to DWI of lung cancer. Future studies are required to investigate this issue, as a high robustness is a prerequisite for the use of DWI in treatment monitoring. A recently published study using an ice water phantom has demonstrated that DWI data acquisition using a DWI protocol designed for comparability across scanners might compensate differences in scanner geometry leading to a high robustness of ADC measurements [40]. However, based on our data, studies on the combined use of ADC and SUV values derived from PET/MRI for the prediction of therapy response are rewarded.

This study has some limitations. First, due do the length of the examination, we did not use respiratory gating for DWI measurements. Second, because tumor grading was not available for all patients, we did not test for a potential correlation between tumor grading and the SUV or ADC. Overall, the number of included patients is relatively low. Therefore, these first results need further confirmation.

In conclusion, this simultaneous PET/MRI study corroborates the assumed significant inverse correlation between increased metabolic activity on FDG-PET and restricted diffusion on DWI in NSCLC. The simultaneous acquisition of the ADC and SUV results in complementary information of the assessed cancer tissue. Further studies are required to investigate the correlation of the SUV and ADC with regard to the underlying histology and tumor response after chemotherapy.

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