Decayed Plasma Ascorbate Levels in Stage IV Melanoma Patients

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Key words
vitamin C
ascorbic acid
cancer
chemotherapy
ipilimumab
scurvy

Abstract

It has been reported that cancer patients frequently express low ascorbate (ascorbic acid, vitamin C) blood levels. However, so far this was not shown for melanoma patients. Total ascorbate (TAA) levels were determined in plasma of healthy control individuals (n = 31, mean age: 47.3 years, TAA: 64.86 μM) and in 126 melanoma patients (stage I: n = 30, mean age: 51 years, TAA: 59.95 μM; stage II: n = 30, mean age: 46.8 years, TAA: 58.85 μM; stage III: n = 32, mean age: 48.6 years, TAA: 57.27 μM; stage IV: n = 34, mean age: 51.1 years, TAA: 47.16 μM). Plasma TAA levels in stage IV patients were significantly reduced by 27.3 % when compared to healthy individuals (p = 0.0001, t-test). The reduced plasma TAA levels in stage IV patients negatively correlated with increased S100 and lactate dehydrogenase (LDH) levels. Further, plasma TAA levels were determined in additional 9 stage IV patients directly before and 24 h after intravenous polychemotherapy (carboplatin + paclitaxel, n = 5) or immunotherapy (ipilimumab, n = 4). TAA levels significantly decreased 24 h after therapy (mean TAA before therapy: 48.7 μM; mean TAA after therapy: 43.0 μM; 11.4 % reduction, p < 0.05, t-test).

Ascorbate levels in the plasma of 126 melanoma patients were significantly decreased in the cohort of stage IV patients and were further decreased by polychemo- or immunotherapy in stage IV patients. Considering the importance of adequate ascorbate supply, ascorbate substitution in physiological doses could be considered for late-stage melanoma patients.

Introduction

An estimated 3.2 million new cases of cancer and 1.7 million deaths from cancer occurred in Europe in 2008 [1]. The fact that only 5–10 % of all cancer cases are due to genetic defects and that the remaining 90–95 % are due to lifestyle factors (including smoking, alcohol, physical inactivity, obesity and sun exposure), nutrition, infections and environmental pollutants provide major opportunities for preventing cancer [1]. Malignant melanoma is a tumor derived from melanocytes, manifesting mainly on the skin, and in rare cases, from mucous membranes. The tumor-specific 10-year survival for patients with primary melanomas without any evidence of metastasis is 75–85 %, decreasing to 20–40 % for those patients with clinically apparent regional lymph node metastases. Distant organ metastases (stage IV disease) have a poor prognosis with a median survival in untreated patients of 6–9 months [2].

Several studies have shown that plasma ascorbate levels are decreased in patients with malignancies, such as hepatocellular carcinoma [3], lung cancer [4], colorectal cancer [5] and prostate cancer [6]. A basic biochemical role for ascorbate is to accelerate hydroxylation reactions in a number of biosynthetic pathways. In many of these reactions, ascorbate directly or indirectly provides electrons to enzymes that require prosthetic metal ions in a reduced form to achieve full enzymatic activity. Ascorbate is required in the synthesis of carnitine from lysine, collagen and neurotransmitter synthesis, cytochrome P-450 activity, cholesterol metabolism, detoxification of exogenous compounds, and as an antioxidant [7]. Moreover, ascorbate is essential for immunoglobulin synthesis. In cell-mediated immunity, immunocompetence is exercised overwhelmingly by lymphocytes, which contain high concentrations of ascorbate relative to other cells. In addition, ascorbate is required for active phagocytosis and
enhances interferon production [7]. In line, impairment of lymphocyte activation, and macrophage and neutrophil mobilization were reported in scorbutic animals [9].

**Aim of the study:** Considering the importance of adequate ascorbate supply for hundreds of biochemical processes and a functional immune system, we asked whether plasma ascorbate levels correlated with disease stage in patients afflicted with malignant melanoma and whether intravenous systemic therapy (polychemotherapy or immunotherapy) in stage IV melanoma patients influenced the plasma ascorbate level.

### Materials and Methods

**Ethics statement**

The use of human plasma samples was approved by the medical ethical committee of the University of Tübingen, Germany, and was performed in accordance with the Declaration of Helsinki Principles. The plasma samples were gathered during routine blood withdrawals. Every patient signed an informed consent form prior to blood withdrawal allowing the determination of plasma ascorbate level for research purposes.

**Recruitment of patients and controls**

Patients aged 18 years and older included in this study. Patients with artificial ascorbate supplementation (e.g. taking ascorbate- or multi-vitamin pills) were excluded from this study. The control cohort consisted of healthy volunteering employees (nurses, dermatologists and laboratory staff) of the Department of Dermatology, University of Tübingen, Germany, aged 18 years and older and was stratified by the exclusion criteria stated above. The control cohort and the stage I and stage II melanoma patients had an ECOG Performance Status grade 0. The stage III melanoma patients had an ECOG Performance Status grade 0 or 1, and the stage IV melanoma patients grade 3 or 4.

**HPLC ascorbate measurements**

For the quantification of ascorbate in EDTA-plasma, samples were immediately mixed with an equal amount of 5% perchloric acid (Sigma, Munich, Germany) and stored at −80°C. The time between blood withdrawal and snap-freezing was less than 10 min. For analysis of plasma ascorbate content, samples were thawed and centrifuged at 13 000 × g for 5 min at 4°C. For reduction of DHA and determination of TAA, the supernatant was mixed with 0.15 M aqueous Tris(2-carboxyethyl)phosphine (TCEP, ABCR, Karlsruhe, Germany) solution (v/v: 2/1) and incubated for 5 min. Afterwards, the mixture was centrifuged at 13 000 × g for 5 min at 4°C and transferred into light-protected micro-vials for chromatographic analysis. As external standards, 2 aqueous ascorbic acid solutions (50μM and 100μM) were prepared, diluted with an equal amount of perchloric acid (5%), mixed with 0.15 M aqueous TCEP solution (v/v: 2/1) incubated for 5 min. The mixture was centrifuged at 13 000 × g for 5 min at 4°C and transferred into light-protected micro-vials for chromatographic analysis. All analytical procedures were performed on ice and under dimmed light to avoid ascorbic acid degradation. For HPLC analysis, samples were placed in a cooled autosampler (Varian, Darmstadt, Germany). Chromatography was carried out on a reversed-phase column (Reprosil-Pur 120 C18 AQ 5μM; Trentec, Gerlingen, Germany) with a mobile phase consisting of 5 mM aqueous sodium phosphate buffer, pH 2.5 at a flow rate of 1 ml/min. Detection was performed by using an electrochemical detector (ESA Detektor Coulochem II, Chelmsford, United Kingdom) and a Model 5011 high-sensitivity analytical cell (ESA) at −300 mV (E1, upstream) and +300 mV (E2, downstream). Chromatograms were recorded and analyzed using Star chromatography workstation software version 5.31 (Varian, Darmstadt, Germany).

**Statistical analysis**

Statistical analysis of plasma ascorbate level was performed by using Student’s t-test. P-values < 0.05 were considered as statistically significant.

### Results

**Plasma ascorbate is decreased in more advanced stages of melanoma and negatively correlates with plasma S100 and LDH levels**

We determined total ascorbate (TAA) levels in plasma of healthy control individuals (n = 31, mean age: 47.3 years, TAA: 64.86μM [26.68μM–113.64μM; SD: 1.11μM]) and in 126 melanoma patients (stage I: n = 30, mean age: 46.8 years, TAA: 59.95μM [33.80μM–106.25μM; SD: 1.0μM]; stage II: n = 30, mean age: 46.8 years, TAA: 58.85μM [28.83μM–88.06μM; SD: 1.20μM]; stage III: n = 32, mean age: 48.6 years, TAA: 57.27μM [27.63μM–80.11μM; SD: 1.17μM]; stage IV: n = 34, mean age: 51.1 years, TAA: 47.16μM [4.82μM–73.99μM; SD: 1.0μM]) by HPLC (Table 1). Plasma TAA levels constantly decreased from stage I to stage III, but the difference was not significant when compared to the healthy controls. In stage IV patients, the ascorbate plasma levels were significantly reduced by 27.3% when compared to healthy individuals (p = 0.0001, t-test). The ascorbate plasma levels of stage IV patients were also signifi-

### Table 1 Plasma total ascorbate levels of 126 stage I–IV melanoma patients and 31 controls.

<table>
<thead>
<tr>
<th>cohorts</th>
<th>n</th>
<th>mean age</th>
<th>TAA [μM]</th>
<th>% reduction in TAA</th>
<th>p-value, t-test</th>
</tr>
</thead>
<tbody>
<tr>
<td>control</td>
<td>31</td>
<td>47.3</td>
<td>64.8</td>
<td>-</td>
<td>-</td>
</tr>
<tr>
<td>stage I</td>
<td>30</td>
<td>51</td>
<td>59.95</td>
<td>7.6</td>
<td>0.253</td>
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<tr>
<td>stage II</td>
<td>30</td>
<td>46.8</td>
<td>58.85</td>
<td>9.3</td>
<td>0.196</td>
</tr>
<tr>
<td>stage III</td>
<td>32</td>
<td>48.6</td>
<td>57.27</td>
<td>11.7</td>
<td>0.069</td>
</tr>
<tr>
<td>stage IV</td>
<td>34</td>
<td>51.1</td>
<td>47.16</td>
<td>27.3</td>
<td>0.0001</td>
</tr>
<tr>
<td>stage IV S100&lt;0.1μg/l</td>
<td>12</td>
<td>52.8</td>
<td>51.78</td>
<td>20.2</td>
<td>0.028</td>
</tr>
<tr>
<td>stage IV S100&gt;0.1μg/l</td>
<td>21</td>
<td>50.9</td>
<td>44.04</td>
<td>32.1</td>
<td>0.0002</td>
</tr>
<tr>
<td>stage IV LDH&lt;250 U/l</td>
<td>21</td>
<td>51.8</td>
<td>52.05</td>
<td>19.8</td>
<td>0.006</td>
</tr>
<tr>
<td>stage IV LDH&gt;250 U/l</td>
<td>12</td>
<td>51.1</td>
<td>37.76</td>
<td>41.8</td>
<td>0.0001</td>
</tr>
</tbody>
</table>

TAA: total ascorbic acid; LDH: lactate dehydrogenase
could not elevated (> 0.1 in stage IV patients: The lowest plasma ascorbate concentrations (LDH)-values (both prognostic markers for malignant melanoma) in stage IV patients: The lowest plasma ascorbate concentrations were measured in melanoma patients with pathologically elevated (> 0.1 µg/l) S100-values (both prognostic markers for malignant melanoma) in stage IV patients: The lowest plasma ascorbate concentrations were measured in melanoma patients with pathologically elevated (> 0.1 µg/l) S100-values (mean TAA: 44.04 µM [4.82 µM–73.99 µM; SD: 0.94 µM]; 32.1 % reduction compared to controls; p = 0.0002, t-test) and elevated (> 250 U/l) LDH-values (mean TAA: 37.76 µM [4.82 µM–64.28 µM; SD: 0.92 µM]; 41.8 % reduction compared to controls; p = 0.0001, t-test) (Table 1).

Plasma ascorbate decreases after poly-chemotherapy and immunotherapy
Second, we asked whether plasma ascorbate levels were influenced by systemic intravenous therapies performed on stage IV melanoma patients. To this end, plasma ascorbate levels were determined in 9 patients directly before and 24 h after either polychemotherapy (carboplatin + paclitaxel) or immunotherapy targeting CTLA4 (ipilimumab) [9,10] (Table 2). We detected significantly reduced plasma ascorbate levels 24 h after therapy (48.7 ± 17.4 µM vs. 43.0 ± 14.7 µM; p < 0.05, t-test) (Table 2). We could not find a significant difference of the reduction of plasma ascorbate levels caused by poly-chemotherapy when compared to immunotherapy.

Discussion

Critical steps in the determination of plasma ascorbate levels
In this study we show for the first time (to the best of our knowledge) that plasma ascorbate levels are reduced in stage IV melanoma patients when compared to healthy age-matched controls. Comparing the mean plasma ascorbate level of our healthy control cohort (64.86 µM), we find a discrepancy in comparison to plasma ascorbate concentrations in the literature. In the European Prospective Investigation into Cancer and Nutrition (EPIC-Norfolk prospective study), the mean ascorbate serum concentrations in the different cohorts were 20.8–72.6 µM with a calculated mean of 47.3 µM (the exact mean was not communicated in the study) [11]. Secondly, in the more recent European Prospective Investigation into Cancer and Nutrition (EPIC-EURGAST), the mean ascorbate serum concentration in controls was 41.5 µM [12]. However, in the latter 2 studies the processing of the blood samples for the determination of ascorbate levels was not comparable to the conditions used in the present study. According to the methods section of the 2 studies, the authors did not enforce a quick handling and freezing of the blood samples. Considering the importance of a quick processing (stabilization and subsequent freezing in liquid nitrogen), which is indispensable due to the instability of ascorbic acid, and which we performed within the short time of less than 10 min after blood withdrawal for all samples, this obvious difference in sample handling most likely accounts for the difference of ascorbate concentrations on our control group when compared to the mean values of the latter 2 studies. In line with this assumption, in the second National Health and Nutrition Examination Survey, the median serum ascorbate concentrations were 49.4 µM for men and 64.2 µM for women [13]. In the latter study, the mean ascorbate concentration was comparable to our control group, and the blood samples were put on dry ice after stabilization in the mobile examination centers without further storage at 7 °C over night. Thus, the quick stabilization and constant freezing (on dry ice) is comparable to the handling of the samples in our study and most likely accounts for the similar results obtained in the control samples. The lack of significant reduction of ascorbate plasma levels in stage I–III patients was obviously related to tumor burden. Stage I and II melanoma patients are completely tumor-free after resection of the primary melanoma, and stage III patients after adequate surgical treatment most of the times as well. Only in stage IV there is a metastatic tumor burden with all its consequences.

Impact of high plasma ascorbate levels for cancer development
Ascorbate-rich diets have protective effects on cancer development and other age-related diseases [14]. A recent meta-analysis of 8 case-control studies found a significant preventive effect on cervical neoplasms for increased intake of ascorbate [15]. Further, large epidemiological studies demonstrated a significantly increased risk of dying from cancer for adults [13] and for men [11] with low serum ascorbate concentrations. The prospective “European Prospective Investigation into Cancer and Nutrition” study revealed that higher plasma vitamin C levels were associated with a decreased risk of gastric cancer, independent of the particular gastric cancer anatomical subsite or histological subtype [12].

Impact of low plasma ascorbate levels for cancer patients
Previous clinical studies demonstrated reduced ascorbate levels in non-melanoma cancer patients [4,16] sometimes even leading to scurvy [17]. In the latter case series the authors stressed the fact

Table 2

<table>
<thead>
<tr>
<th>Patient</th>
<th>Treatment</th>
<th>TAA [µM] pre</th>
<th>TAA [µM] post</th>
<th>% reduction in TAA</th>
<th>p-value, t-test</th>
</tr>
</thead>
<tbody>
<tr>
<td>1</td>
<td>Carboplat</td>
<td>41.1</td>
<td>36.2</td>
<td>11.9</td>
<td></td>
</tr>
<tr>
<td>2</td>
<td>Carboplat</td>
<td>8.5</td>
<td>6.9</td>
<td>18.1</td>
<td></td>
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<tr>
<td>3</td>
<td>Carboplat</td>
<td>53.9</td>
<td>46.0</td>
<td>14.7</td>
<td></td>
</tr>
<tr>
<td>4</td>
<td>Carboplat</td>
<td>64.3</td>
<td>45.4</td>
<td>29.4</td>
<td></td>
</tr>
<tr>
<td>5</td>
<td>Carboplat</td>
<td>46.9</td>
<td>42.2</td>
<td>10.1</td>
<td></td>
</tr>
<tr>
<td>1</td>
<td>Ipilimumab</td>
<td>74.0</td>
<td>64.4</td>
<td>13.0</td>
<td></td>
</tr>
<tr>
<td>2</td>
<td>Ipilimumab</td>
<td>56.4</td>
<td>51.6</td>
<td>8.4</td>
<td></td>
</tr>
<tr>
<td>3</td>
<td>Ipilimumab</td>
<td>52.3</td>
<td>48.8</td>
<td>6.6</td>
<td></td>
</tr>
<tr>
<td>4</td>
<td>Ipilimumab</td>
<td>41.1</td>
<td>45.2</td>
<td>-9.9</td>
<td></td>
</tr>
<tr>
<td>mean</td>
<td></td>
<td>48.7</td>
<td>43.0</td>
<td>11.4</td>
<td>&lt;0.05</td>
</tr>
</tbody>
</table>

TAA: total ascorbic acid; pre: before therapy; post: 24 h after therapy; Carbotax: carboplatin + paclitaxel.
that scurvy should be considered in patients with cancer because of the high incidence of malnutrition caused by chemotherapy, cachexia caused by the disease, and other factors that might lead to an unbalanced dietary intake such as exclusive parenteral nutrition, depression, impaired taste, dysphagia, and abdominal pain. Further, weakness, anorexia, and depression are common in scurvy but also in patients with cancer. The authors concluded that clinicians should suspect ascorbate deficiency when a patient has hemorrhagic features without a clear explanation [17]. The importance of ascorbate for innumerable biochemical processes in the human organism is beyond dispute. Since humans and other primates lack the enzyme L-gulonolactone oxidase required for ascorbate synthesis, an adequate oral uptake is vital [7]. Considering the dependency of the immune system on adequate ascorbate supply [8] and the importance of the immune system for phagocytosis and enhances interferon production [7]. In addition, scorbutic animals show an impairment of lymphocyte role in immunoglobulin synthesis. Moreover, lymphocytes contain high concentrations of ascorbate, and ascorbate is required for phagocytosis and enhances interferon production [7]. In addition, scurritic animals show an impairment of lymphocyte activation and macrophage and neutrophil mobilization [8]. Together, for the immune response, ascorbate seems to be a double-edged sword functioning both as pro- and anti-inflammatory stimulate that inhibits tumor growth. On the cellular level, growth inhibition of melanoma cells was driven by cell cycle arrest and caspase-3 activation with subsequent apoptosis induction [21]. Further, ascorbate reduced the expression of the transferrin receptor in melanoma cells, resulting in a down-regulation of cellular iron uptake followed by apoptosis induction [22]. Due to recent pharmacokinetic data showing that intravenous administration of ascorbate results in high plasma levels not achievable by oral uptake and in a significant flux of H2O2 to tumors, the use of i.v. ascorbate in pharmacological doses in the treatment of cancer seems plausible and is currently being investigated in clinical trials and in experimental studies [23].

Conclusions ▼

We were able to demonstrate that plasma ascorbate levels were significantly lower in stage IV melanoma patients. Systemic therapy on stage IV patients further reduced plasma ascorbate levels. Considering the importance of adequate ascorbate supply for biochemical processes and host defense, our results suggest that late-stage melanoma patients might benefit from physiological ascorbate supplementation.

Acknowledgements ▼

This project was supported by grants from the Deutsche Forschungsgemeinschaft SFB 773: “Understanding and overcoming drug resistance of solid tumors” to CG and CB. SV was supported by a research fellowship from the Deutsche Forschungsgemeinschaft (VE671/2-1). CB and SV received a research grant from the Wissenschaftsfoerderung der Deutschen Brauwirtschaft e.V.

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