Preparation and Characterization of Chitosan/β-cyclodextrin Nanoparticles Containing Plasmid DNA Encoding Interleukin-12

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Key words
- chitosan
- β-cyclodextrin
- plasmid
- nanoparticle
- interleukin-12

Abstract

Background: Interleukin-12 (IL-12) as a cytokine has been proved to possess antitumor effects via stimulating the immune system. Non-viral gene delivery systems offer several advantages, including easiness in production, low cost, safety; low immunogenicity and can carry higher amounts of genetic material without limitation on their sizes.

Methods: pUMVC3-hIL12 loaded Low Molecular Weight chitosan/β-cyclodextrin (LMW CS/CD) nanoparticles were prepared using ionotropic gelation method and characterized in terms of size, zeta potential, polydispersity index, morphology, loading efficiency and cytotoxicity against the CT-26 colon carcinoma cell line.

Results: All prepared particles were spherical in shape and nano-sized (171.3 ± 2.165 nm, PdI: 0.231 ± 0.014) and exhibited a positive zeta potential (34.3 ± 1.55). The nanoparticles demonstrated good DNA encapsulation efficiencies (83.315% ± 2.067). Prepared pUMVC3-hIL12 loaded LMW CS/CD nanoparticles showed no cell toxicity in murine CT-26 colon carcinoma cells. At the concentration of 0.1 μg/ml of nanoparticles, the transfection ability was obviously higher than that of the naked DNA.

Conclusion: LMW CS/CD-plasmid DNA nanoparticles encoding IL-12 prepared using ionotropic gelation method with no toxic effect on the tested cells can be considered as a basis for further gene delivery studies both in vitro and in vivo to enhance the expression of IL-12.

Introduction

Nanoparticle-based drug delivery systems offer numerous privileges including targeting the drug into the potential active site, thus decreasing its side effect, controlling the release of the drug at the active site and increasing the drug uptake. Also they produce some excellent pharmacokinetic properties leading to enhance the drug absorption and distribution. Application of nanoparticles in the delivery of nucleotides into their active sites has been the subject of several researches in the past few decades [1]. Gene therapy, as a new and promising way to treat some of most complicated health disorders, is defined as the transfer of a genetic material into a cell for therapeutic benefits [2]. Generally, there are 2 types of gene delivery systems including viral and non-viral. Although viral gene delivery systems possess high efficiency in various human tissues, they have some limitations like immunogenicity, toxicity and possible viral gene integration into the human genome. On the other hand, non-viral systems (including physical and chemical approaches), have lower toxicity and immunogenicity. In addition, manipulation and preparation of non-viral delivery systems are easier in comparison with those of viral [3,4]. The most important non-viral vectors are cationic lipids and cationic polymers which produce condensed complexes with negatively charged DNA (lipoplexes and polyplexes). The complexes protect DNA and facilitate cell uptake and intracellular delivery [5,6]. Of the cationic polymers investigated so far, chitosan has been introduced as a potential gene carrier. Chitosan is a linear biodegradable, biocompatible, non-allergenic and nucleic acid bindingable polysaccharide composed of randomly distributed N-acetyl-D-glucosamine and β-(1,4)-linked D-glucosamine. Chitosan generates a positively charged complex in interaction with DNA leading to enhanced cellular interaction and uptake. Chitosan can swirl across the membrane bilayer and facilitate the cellular uptake of the polyplex [1]. In spite of the main advantages of chitosan polyplexes, their
capacity to efficiently deliver the gene of interest into cells is still limited. It can be due to some reasons such as undefined
physical shapes, dissociation of the complexes in the presence of
anions and a limited capacity to co-associate other functional
molecules [7]. Cyclodextrins are a family of cyclic oligosaccha-
rides which consists of (α-1,4)-linked α-D-glucopyranose unit
with a hydrophilic outer surface and a lipophilic central cavity
[8]. Cyclodextrins present many advantages as potential core
molecules for the development of nonviral vectors [9–13].
Indeed new generations of polysaccharide nanocarriers consist-
ing of chitosan and cyclodextrins have been developed in recent
years to use the promising behavior of chitosan nanoparticles
with the excellent biopharmaceutical properties of cyclodextrin
[7]. The objective of the present study was to prepare chitosan-
cyclodextrin-plasmid DNA nanoparticles encoding interleukin-12 that has been proved to possess antitumor effects via stimulating the immune system. Also the physicochemical char-
acteristic as well as cytotoxicity of the prepared formulations were evaluated against the CT-26 colon carcinoma cell line.

Materials and Methods

Materials

PUMVC3-hIL12 (Aldevron, Fargo, ND, USA), Chitosan (CS) MW 190–310kDa 75–85 % degree of deacetylation, Beta cyclodextrin
(CD), 3-(4, 5-dimethylthiazol-2-yl)-2, 5-diphenyl tetrazolium bromide (MTT), Lipofectamine™ 2000 Transfection Reagent
(Invitrogen, Carlsbad, CA, USA), Kanamycin, Glycin, Trypan blue
solution, Penicillin and streptomycin (Sigma-Aldrich, Munich,
Germany), Ethylene diamine tetraacetic acid (EDTA), Tris base,
Luria-Bertani broth (LB), Acetic acid (HAc), Sodium Acetate
(NaAc), Triphosphophate (TPP) (Merck, Darmstadt, Germany),
Endotoxin-free plasmid gigaprep Kit (Qiagen, Santa Clarita, CA,
USA), Fetal bovine serum (FBS), RPMI-1640, (Gibco, Invitrogen
Inc., Burlington, Ontario, Canada), Murine CT-26 colon carcino-

Preparation and characterization of low molecular
weight chitosan

The low molecular weight CS (LMW CS) was prepared by oxida-
tive degradation with NaN02 at room temperature. Briefly, 1 %
(w/v) CS was dissolved in 1 % HAc solution, consequently an
appropriate amount of 0.1 M NaNO2 was added dropwise and
the reaction was performed at room temperature for 3 h and
24h, respectively. The reaction mixture was subsequently neu-
tralized with 1 N NaOH to pH 8.0 to precipitate CS. The precipi-
tated CS was recovered by centrifugation, washed several times
with deionized water, and dried [14]. Viscosities of prepared CS
samples in 0.2 M HAc/0.1 M NaAc were measured using an Ubbe-
lohe capillary viscometer (Model Schott mainz, Germany) in a
constant-temperature of 30±0.01°C. Solution concentrations were adjusted based on the viscosity of the samples and the flow
through time was kept in the range of 100–150 s. 5 different con-
centrations were tested for each sample. The intrinsic viscosity was
determined by the intercept of Huggins (ηsp/C = C) plot on
the ordinate at C = 0. Finally the molecular weight of each sample
was calculated based on the classical Mark-Houwink equation:
\[ \eta \text{sp} = K M_n^a \text{c} \], Where \( \eta \text{sp} \) is the intrinsic viscosity of the depoly-
merized CS, K and a are constants for given solute-solvent system
and temperature. For CS, they are influenced by the degree of
deacetylation, pH, and ionic strength of the solvent [14]. Accord-
ing to our solvent system (0.2 M HAc/0.1 M NaAc), k and a were
0.000104, 0.001424 and 1.12, 0.96 for 69 and 84 degree of
deacetylation, respectively [15].

Preparation of LMW CS/CD nanoparticles

Nanoparticles were obtained by ionotropic gelation Method
according to Alonso et al. [7]. 2 aqueous phases containing: (1)
LMW CS solution (2 mg/ml), and (2) CD solution (20 mg/ml) con-
taining 3 mg/ml TPP as cross-linker, were mixed under vortexing
for 10 min. For nanoparticles encapsulating the pDNA, the
required amount of the plasmid pUMVC3-hIL12 was incorpo-
rated directly into the CD/TPP phase [7]. To this end, plasmid
DNA was firstly extracted from transformed E.coli (DH5-α),
then the concentration of purified plasmid DNA was determined at
260 nm using nanodrop spectrophotometer.

Physicochemical characterization of the pUMVC3-hIL12
loaded LMW CS/CD nanoparticles

Evaluation of particle size, polydispersity index, and zeta potential of the pUMVC3-hIL12 loaded LMW CS/CD nanoparticles were performed by zetasizer nano particle analyzer (Nano ZS, Red badge, ZEN3600). Particle morphology of the selected
pUMVC3-hIL12 loaded LMW CS/CD nanoparticles was deter-
mined by transmission electron microscopy (TEM). One drop of
LMW CS/CD nanoparticles was placed on a copper grid and
stained with 1 % uranyl acetate solution for 5 s. The grid was
allowed to dry further for 10 min and was then examined with
the electron microscope (LEO 906, Germany). The amount of
plasmid DNA encapsulated in the LMW CS/CD nanoparticles was calculated by considering the difference between the total
amount of DNA added into the preparation medium through the
coacervation process and the amount of non-entrapped DNA
remaining in the supernatant after centrifugation (12000 RPM,
for 30 min, at 4 °C) of prepared nanoparticles. For this purpose,
the supernatant was spectrophotometrically analyzed at 260 nm
for DNA concentration [16].

Evaluation of cytotoxicity

Cytotoxicity of the pUMVC3-hIL12 loaded LMW CS/CD nanoparticles was evaluated by the MTT based cytotoxicity assay in murine
CT-26 colon carcinoma cells. MTT assay is based on the ability of
living cells to reduce MTT, a water-soluble yellow dye, to a purple
colored formazan precipitates product by mitochondrial reduct-
ase enzymes. The cells were seeded in a 96-well plate at a density of
5×104 cells/cm2 in100μl of RPMI-1640 medium supplemented
with 10 % FBS and incubated for 24h at 37°C under 5 % CO2. After-
ward, the medium was removed and supplied with 200 μl of fresh
culture medium. Subsequently, pDNA loaded LMW CS/CD nanoparticles were added to the cells at a serial dose of 1.6, 0.8, 0.4, 0.2,
0.1, 0.05, 0.025μg pUMVC3-mIL12/μl. Non-treated cells as well as
and cells treated with lipofectamine™ were used as controls and
incubated for the same duration of time. Following 24h of incuba-
tion at 37°C under 5 % CO2, the medium was removed and fresh
cell culture medium was added, then the cells were incubated with
20μl MTT (0.5 mg/ml MTT in medium) for 4h. Subsequently
the medium was removed, and the formazan precipitates formed
in living cells were dissolved in 200μl DMSO and 25μl Sorenson
buffer per well. Relative viability (%) was calculated based on
absorbance at 570nm using an ELISA plate reader. Viability of
non-treated control cells was arbitrarily defined as 100 %. The
relative cell viability was calculated as [Absample/Abscontrol]×100.
Data were presented as mean±SD (n=4).
In vitro transfection studies
Murine CT-26 colon carcinoma cells were seeded into 24-well plates at a density of 80,000 cells/well in 1 ml of RPMI-1640 medium supplemented with 10% FBS and incubated for 24 h at 37 °C under 5% CO₂. Prior to transfection, the medium was removed, and the cells were rinsed with PBS (pH 7.4), then supplied with 900 μl of RPMI-1640 medium without FBS. The cells were incubated with pDNA loaded LMW CS/CD nanoparticles at concentrations range of 0.025 to 0.8 μg/ml for 6 h at 37 °C under 5% CO₂. Subsequently, the serum-free medium was removed and the wells were washed with PBS. Fresh medium (RPMI-1640 containing 10% FBS) was then added to each well. The cell culture plate was incubated for another 48 h at 37 °C under 5% CO₂. Non-treated cells and cells transfected with naked plasmid (as negative controls) were used and incubated for the same duration of time. All experiments were done in triplicates, with 3 separate experiments to demonstrate reproducibility.

For quantitative analysis of the pUMVC3-hIL12 expression, the human IL-12p70 enzyme-linked immunosorbent assay (ELISA) Kit was used according to the manufacturer’s instructions. For this purpose, the collected culture supernatants were analyzed for the measurement of hIL-12 p70. The amount of the protein was determined as picogram per ml.

Results

Preparation and characterization of low molecular weight chitosan
The LMW CS was prepared by oxidative degradation with 0.1 M NaNO₂ at room temperature using 1% (w/w) CS dissolved in 1% HAc solution. The molecular weight of 24 h degraded CS was calculated based on the classical Mark-Houwink equation as 12–32 KDa. Results related to 24 h degraded samples are shown in Table 1 and Fig. 1.

The pUMVC3-hIL12 loaded LMW CS/CD nanoparticles formation and characterization
LMW CS/CD nanoparticles were prepared by ionotropic gelation in the presence of TPP, as described in the methodology section. The average size, polydispersity index and Zeta potential of pUMVC3-hIL12 loaded LMW CS/CD nanoparticles are given in Table 2 and depicted in Fig. 2. Also the amount of DNA loaded in the nanoparticles was calculated as encapsulation efficiency (%) which is shown in Table 2.

A typical transmission electron microscopy (TEM) image of F1 is shown in Fig. 3. As can be seen from this figure, the particles were nano-sized and spherical in shape.

Evaluation of cytotoxicity and viability
The cell toxicity of pUMVC3-hIL12 loaded LMW CS/CD nanoparticles was investigated by MTT-based cytotoxicity assay in murine CT-26 colon carcinoma cells. The prepared polyplexes showed no cell toxicity in murine CT-26 colon carcinoma cells. The average cell viability determined using MTT assay for

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**Table 1**  Viscosity measurement of 24 h degraded chitosan.

<table>
<thead>
<tr>
<th>Chitosan Conc. (g/ml)</th>
<th>0.001</th>
<th>0.0025</th>
<th>0.005</th>
<th>0.0075</th>
<th>0.01</th>
</tr>
</thead>
<tbody>
<tr>
<td>Average Flow throw Times (S) ± SD</td>
<td>98.6 ± 1.140</td>
<td>102 ± 0.707</td>
<td>109.6 ± 0.894</td>
<td>126.4 ± 4.505</td>
<td>136.4 ± 4.560</td>
</tr>
<tr>
<td>Viscosity (cp)</td>
<td>1.095</td>
<td>1.133</td>
<td>1.217</td>
<td>1.404</td>
<td>1.515</td>
</tr>
<tr>
<td>Specific Viscosity</td>
<td>0.014</td>
<td>0.049</td>
<td>0.127</td>
<td>0.300</td>
<td>0.403</td>
</tr>
<tr>
<td>Reduced Viscosity (ml/g)</td>
<td>14.403</td>
<td>19.753</td>
<td>25.514</td>
<td>40.054</td>
<td>40.329</td>
</tr>
</tbody>
</table>

**Table 2**  Formulation, particle size, polydispersity index, and zeta potential of pUMVC3-hIL12 loaded LMW CS/CD nanoparticles.

<table>
<thead>
<tr>
<th>Formulation</th>
<th>pDNA (μg/ml)</th>
<th>Chitosan (mg/ml)</th>
<th>cyclodextrin (mg/ml)</th>
<th>TPP (mg/ml)</th>
<th>Average Particle Size* (nm) ± SD</th>
<th>Average Polydispersity Index ± SD</th>
<th>Average Zeta Potential (mV) ± SD</th>
<th>Encapsulation Efficiency (%) ± SD</th>
</tr>
</thead>
<tbody>
<tr>
<td>F1</td>
<td>100</td>
<td>2</td>
<td>20</td>
<td>3</td>
<td>171.3 ± 2.165</td>
<td>0.231 ± 0.014</td>
<td>34.3 ± 1.55</td>
<td>83.315 ± 2.067</td>
</tr>
</tbody>
</table>

*Average particle size measured as Z-average after sonication

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different DNA concentrations ranged from 97.7 ± 5 to 126.8 ± 23 (Fig. 4). In contrast, Lipofectamine/IL-12 complexes as a control resulted in less than 60% cell viability.

In vitro transfection studies

Fig. 5 compares the transfection efficiencies of the pUMVC3-hIL12 loaded LMW CS/CD nanoparticles in different concentrations in CT-26 carcinoma cells. Untreated CT-26 cells showed no production of IL-12. At plasmid concentrations below or above 0.1 μg/ml, the transfection efficiencies of pUMVC3-hIL12 loaded LMW CS/CD nanoparticles were not remarkably different from that of the naked DNA. At the concentration of 0.1 μg/ml, the transfection ability was obviously higher than that of the naked DNA.

Discussion

Immunotherapy-based strategies for cancer therapy try to increase the local and systemic immune response against the cancer cells and possibly provide a less toxic approach. IL-12 has been shown to have superior antitumor activity compared to other cytokines. To get optimal results, cytokines are required to be available over an extended period of time, which is not possible with IL-12 protein owing to its short half-life [17]. According to the several studies reported in this area, polymeric vectors like CS, release their payload in a sustained manner [1]. Hence, we hypothesized that gene therapy using pUMVC3-hIL12 loaded LMW CS/CD nanoparticles possibly provide noteworthy advantages over protein therapy due to the potential ability of polymeric based nanoparticles to produce sustained levels of IL-12 in the tumor location. Based on the previous studies in this regard, LMW CS showed the ability to efficiently interact with the pDNA [7]. Thus CS in the molecular weight of 12–32 KDa prepared by oxidative degradation method was used in this study. pUMVC3-hIL12 loaded LMW CS/CD nanoparticles were prepared via ionotropic gelation method which is simple, quick, economical, and requires no toxic solvents compared to the other approaches [1]. The electrostatic interaction between positively charged CS and negatively charged DNA and CD is involved in the nanoparticle formation. Furthermore, the ability of CS to undergo a liquid–gel transition due to its ionic interaction with TPP is of importance in the process. The maximum LMW CS/CD/TPP mass ratio to produce optimum nanoparticle characterization was chosen to be 6/1/1 (w/w) according to Teijeiro-Osorio et al. [7]. Following the preparation of the nanoparticles, they were characterized in terms of size, zeta potential, morphology, loading efficiency, cytotoxicity, and finally transfection efficiency. As shown in Fig. 2, 3, all particles were nano-sized (171.3 ± 2.165 nm, PdI: 0.231 ± 0.014) and exhibited positive zeta potential (34.3 ± 1.55). In general, zeta potential is one of the important particle characteristics, because more pronounced zeta potential values lead to stabilization of the particle suspension due to the electrostatic repulsion between particles with the same electric charge which prevents aggregation [18]. Furthermore, the positively charged particles can facilitate adherence to negatively charged cellular membranes which will induce and increase intracellular uptake [19,20]. Alonso et al. prepared ionically crosslinked nanoparticles based on LMW CS (10KDa) with plasmid DNA and TPP at the same ratio of CS/TPP (6/1/1). Based on their results, the size of prepared particles were in the range of 109±13 to 165±10 and zeta potential of

Nahaei M et al. Chitosan/β-cyclodextrin Nanoparticles for IL-12 Gene Delivery... Drug Res 2013; 63: 7–12
7.7 ± 0.4 to 10.8 ± 4.2. Also in cytotoxicity test, they were non toxic. In comparison with these results and ours, it is obvious that there are no considerable differences in the physicochemical properties and cytotoxicity of LMW CS/TPP (6/1) nanoparticles with LMW CS/CD/TPP (6/1/1) nanoparticles [21].

The mean encapsulation efficiency of our prepared nanoparticles for DNA was 83.31% as shown in Table 2. The high encapsulation efficiency is observed in the previous studies for other CS nanoparticles which might be due to the high affinity of CS to DNA [22]. It is important to evaluate the toxicity of the prepared nanoparticles in a cell line model. MTT assay is a quantitative colorimetric method to determine cell proliferation and it utilizes the yellow tetrazolium salt which is metabolized by mitochondrial succinic dehydrogenase activity of proliferating cells. The ability of cells to reduce MTT provides an indication of the mitochondrial integrity and activity, which, in turn, may be interpreted as a measure of viability. Comparison of results between control and test cultures provides an indication of the cytotoxic effect of test compounds [14]. The effect of different CS derivatives as well as CS/CD complexes on the viability of tumor cells has been extensively studied so far [26–29] and some controversial results have been reported. Some studies reported no toxicity against the tested cell lines at the used concentrations, both for CS or CS/CD complexes [7]; whereas, other studies reported the inhibition of the growth of tumor cell in the presence of CS or its derivatives [23,24]. Also, there are few reports about the stimulation effect of CS, for example, Dhiman and coworkers showed that the reduction percentage of MCF-7 cells brought by Tamoxifen in the presence and absence of CS are 15% and 50% respectively, indicating strong stimulation (inhibition of reduction) of the selected cell lines [25]. Also, in another study [26], the stimulation effect of CS on orchid meristem culture and orchid protocorm propagation was reported. Prepared pUMVC3-hIL12 loaded LMW CS/CD/TPP nanoparticles showed no cell toxicity with average cell viability of almost over 97% in murine CT-26 colon carcinoma cells (97.7 ± 5 to 126.8 ± 23) that was significantly higher than the average cell viability of lipofectamine treated cells (58.2 ± 9.5). Even the viability percentages of the tested cells were increased by about 120% in the case of 1.6 and 0.8 μg/ml. Having low toxicity will make it possible to increase the dose of these nanoparticles or use repeated administrations. Based on the studies reported for the in vitro release properties of polymeric vectors like CS, a sustained manner has been reported [1]. Nevertheless, the in vitro release studies can be a representative of the in vitro stability of the particles, rather than in situ release behavior. Since the intracellular release condition is completely different, so the in vitro release studies cannot mimic the real conditions inside the cells. Consequently, we decided to study the effect of our prepared nanoparticles on the expression of IL-12, as a correct way to find an idea about the intracellular release behavior as well as the transfection efficiency.

CT-26 colon carcinoma cells as cancer cells which are less immunogenic have been widely used for the study of the anti-tumor activity of IL-12. According to our results which is depicted in Fig. 5, CT-26 cell as untreated control showed approximately no production of IL-12 (3.3 pg/ml). On the other hand, following 48h treatment of the cells with IL-12 naked plasmid, 82.3 pg/ml of IL-12 protein was detected using ELISA Kit indicating the transfection of the cells. We tried to evaluate the transfection efficiencies in longer incubation times, but the best response was obtained in 48h. The expression study of our prepared nanoparticles was performed by 48h treatment of the cells using different plasmid concentrations. LMW CS/CD/TPP nanoparticles resulted in the highest IL-12 production in murine CT-26 at plasmid concentration of 0.1 μg/ml. This is in agreement with the reported studies using LMW CS/TPP chitosan nanoparticles indicating that the best transfection efficiency was at an optimum pDNA loading [7,21]. However, there was no significant difference between the transfection efficiency of our prepared nanoparticles and the naked DNA (p > 0.05). This finding may be due to several reasons, especially the strong electrostatic interaction between CS and CD, which may prevent prompt intracellular DNA release. Indeed, due to the technical difficulties like acidity of the medium, prolonged incubation of the cells (more than 48h) was not possible. Hence, the expression amounts of IL-12 in the cells treated with the sustained release nanoparticles and naked plasmid DNA were not properly differentiated in short incubation times. Alonso et al. found that the transfection efficiency of HMW CS/TPP was higher than some versions of HMW CS/CD/TPP and inclusion of CD into HMW CS/TPP had a negative effect in the transfection efficiency. On the other hand, in the case of nanoparticles prepared using LMW CS, addition of CD enhanced the transfection efficiency but it was not a massive change [7,21]. Taken together, mild preparation process of DNA complexes using LMW CS/CD/TPP yielded the non-toxic nanoparticles with suitable physicochemical properties. Therefore the prepared LMW CS/CD nanoparticles were selected for in vitro expression studies. In the expression studies in spite of the slight increase in IL-12 at plasmid concentration of 0.1 μg/ml, the differences between IL-12 concentration in nanoparticles and naked plasmid DNA were insignificant. Further in vivo studies should be performed using animal models to investigate the effect of the prepared LMW CS/CD nanoparticles on the IL-12 expression in real conditions.

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Conflict of Interest
The authors report no conflict of interest.

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Nahaei M et al. Chitosan/β-cyclodextrin Nanoparticles for IL-12 Gene Delivery... Drug Res 2013; 63: 7–12
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Nahael M et al. Chitosan/β-cyclodextrin Nanoparticles for IL-12 Gene Delivery... Drug Res 2013; 63: 7–12