

Application of Plant Metabonomics in Quality Assessment for Large-Scale Production of Traditional Chinese Medicine

Authors

Zhangchi Ning¹, Cheng Lu², Yuxin Zhang¹, Siyu Zhao¹, Baoqin Liu³, Xuegong Xu³, Yuanyan Liu¹

Affiliations

¹ School of Chinese Materia Medica, Beijing University of Chinese Medicine, Beijing, China

² Institute of Basic Research in Clinical Medicine, China Academy of Chinese Medical Sciences, Beijing, China

³ Zhengzhou Hospital of Traditional Chinese Medicine, Zhengzhou, China

Key words

- plant metabonomics
- large-scale production
- discrimination
- processing
- pharmaceuticals

received October 13, 2012
revised May 6, 2013
accepted May 12, 2013

Bibliography

DOI <http://dx.doi.org/10.1055/s-0032-1328656>
Published online June 27, 2013
Planta Med 2013; 79: 897–908
© Georg Thieme Verlag KG
Stuttgart · New York ·
ISSN 0032-0943

Correspondence

Dr. Yuanyan Liu
School of Chinese Materia Medica
Beijing University of Chinese Medicine
China
No. 6 Middle ring South road
Wangjing
Beijing 100029
China
Phone: + 86 10 84 73 86 58
Fax: + 86 10 84 73 86 11
yyliu_1980@163.com

Correspondence

Dr. Cheng Lu
Institute of Basic Research in Clinical Medicine
China Academy of Chinese Medical Sciences
Nonxiaojie 16# Dong zhimen
Beijing 100700
China
Phone: + 86 10 64 01 44 11 34 03
Fax: + 86 10 84 03 28 81
lv_cheng0816@163.com

Abstract

▼
The curative effects of traditional Chinese medicines are principally based on the synergic effect of their multi-targeting, multi-ingredient preparations, in contrast to modern pharmacology and drug development that often focus on a single chemical entity. Therefore, the method employing a few markers or pharmacologically active constituents to assess the quality and authenticity of the complex preparations has a number of severe challenges. Metabonomics can provide an effective platform for complex sample analysis. It is also reported to be applied to the quality analysis of the traditional Chinese medicine. Metabonomics enables comprehensive assessment of complex traditional Chinese medicines or herbal remedies and sample classification of diverse biological statuses, origins, or qualities in samples, by means of chemometrics. Identification, processing, and pharmaceutical preparation are the main procedures in the large-scale production of Chinese medicinal preparations. Through complete scans, plants metabonomics addresses some of the shortfalls of single analyses and presents a considerable potential to become a sharp tool for traditional Chinese medicine quality assessment.

Abbreviations

▼
TCM: traditional Chinese medicine
CE: capillary electrophoresis
MVDA: multivariate data analysis
PCA: principal component analysis
HCA: hierarchical cluster analysis
PLS: partial least squares
OSC: orthogonal signal correction
OPLS: orthogonal partial least squares

O2PLS: bidirectional orthogonal partial least squares
SIMCA: soft independent modeling of class analogy
PLS-DA: partial least squares discriminant analysis
kNN: k nearest neighbors
ANN: artificial neural networks
MEND: matched filtration with experimental noise determination
AFLP: amplified fragment length polymorphism
TOF: time of flight
UPLC-QTOF-MS: ultra performance liquid chromatography quadrupole time of flight high definition mass spectrometry
BP-ANN: back propagation artificial neural network
ELSD: evaporative light scattering detector
HPTLC: high performance thin layer chromatography
DART: direct analysis in real time photodiode array detector
PAD: least squares support vector machine
LS-SVM: least squares support vector machine
RBF: radial basis function
NIRS: near-infrared spectroscopy
WG: white ginseng
RG: red ginseng
LTQ: linear trap quadrupole
LDA: linear discriminant analysis
SA: similarity analysis
COW: correlation optimized warping
UPGMA: unweighted pair group method with arithmetic mean

FSMWEFA:	fixed size moving window-evolving factor analysis	NMR:	nuclear magnetic resonance spectroscopy
HELP:	heuristic evolving latent projection	MS:	mass spectrometry
LA:	licochoalcone A	TOF:	time of flight
RRLC:	rapid resolution liquid chromatography	UPLC-QTOFMS:	ultra-performance liquid chromatography-quadrupole time-of-flight mass spectrometry
UFLC:	ultrafast liquid chromatography	UPLC-QTOF-HDMS:	ultra-performance liquid chromatography-quadrupole time-of-flight high-definition mass spectrometry
MAS-NMR:	magic angle spinning nuclear magnetic resonance	UPLC-PAD:	ultra-performance liquid chromatography with photodiode array detector
IOP:	iterative optimization procedure	LC-LTQ-Orbitrap:	LC coupled with ESI hybrid linear trap quadrupole orbitrap
SFA:	subwindow factor analysis		
OPR:	orthogonal projection resolution		
EWOP:	evolving window orthogonal projection		
5-GGMF:	5-(α -D-glucopyranosyl-(1-6)- α -D-glucopyranosyloxymethyl)-2-furancarboxaldehyde		

Introduction

Metabonomics is an emerging subject of the post-genome era, which, together with genomics, transcriptomics, and proteomics, jointly constitutes the “Systems Biology” [1]. It is the branch of science concerned with the quantitative understanding of the metabolite components of integrated living systems and their dynamic responses to changes in both endogenous (such as those associated with physiology and development) and exogenous factors (such as environmental factors and xenobiotics) [2]. The success of the application of metabonomics has been illustrated in the literature from the perspective of the diagnosis of diseases such as diabetes [3], hypertension [4], and cancers [5, 6]. In recent years, a wide range of analytical metabonomic techniques have been implemented in research addressing TCM whose qualitative analysis is difficult because of the complexity and diversity of its components. In general, one or two biomarkers are used for identification and authentication of the herbal products. However, this approach does not provide information on the overall chemical composition of the plant extract, which is known to vary widely according to geographical origin, source, cultivar condition, harvesting and processing methods, and storage. Metabonomics, through achieving complete scans, addresses the shortfalls of single-component analysis. The rapid development of analytical instruments is accelerating research on TCM [7]. Additionally, multivariate statistical methods are increasingly improving, allowing the implementation of robust solutions [8]. Within the TCM practice, the majority of species used are plants. The multi-varieties employed in TCM are the main cause of confusion in the herbal medicine market. The identification of these varieties, as the first step in the production of Chinese medicinal preparations, is of great significance for ensuring the safety and effectiveness of clinical treatment. The quality and contents of the active components of herbs are highly variable depending on the species, parts of the herbs, cultivated geographic region, and planting period involved. Adulterants should be distinguishable from plant material and play the role of challenging substances. The processing of a characteristic portion, as the second step in production, appears to be of significance in clinical applications and has been proven to satisfy the requirements of therapeutics. It is essential to unify the degree of processing. In pharmaceutical production, extracts are commonly used. Metabonomics can be effectively applied for the quality control of plant extracts.

Identifying the plant material, processing, and pharmaceutical production is the sequence of manufacture for Chinese medicinal preparations. Here we demonstrate the application of metabonomics in the discrimination of TCM species, TCM production processes, and quality control. To avoid ambiguities, we also illustrated the factors affecting the identification step, the methods used in processing, and the forms of the pharmaceuticals.

Analytical Techniques

In recent years, many metabonomic-based methods have been implemented to facilitate research in the field of TCM. In the pharmacopeia, single-component analyses are employed in most research addressing TCM. Nevertheless, the lack of representativeness of single-component analyses seems to account for a deficiency of convincing data. Metabonomics, through achieving complete scans, addresses the shortfalls of single-component analysis.

The rapid development of analytical instruments is accelerating research on TCM [7]. Metabonomics measures the multi-parametric response of biological systems to a stimulus, typically employing analytical technologies such as NMR or MS to obtain comprehensive profiling and comparison of metabolic “fingerprints” [9]. In addition, other chemical analytical equipments and techniques, such as UV and IR spectroscopy were also employed. For biomarker identification, it is also possible to separate out substances of interest on a larger scale from a complex biological system using techniques such as LC, multidimensional liquid separation systems, GC, and CE. Especially multidimensional liquid separation systems have the potential to become a powerful approach for enriching, separating, and quantifying a large variety of exogenous and endogenous compounds in complex biological samples and TCM preparations, with a powerful separation ability, high resolution and sensitivity, high-peak capacity, and excellent detection in comparison with one-dimensional HPLC. However, every analytical technique has its advantages and drawbacks, as shown in **Table 1**. Multi-analysis techniques can partially overcome the shortcomings of individual analytical techniques. It is believed that with the further development of metabonomics analysis techniques, especially those employing multi-analysis, metabonomics will strongly promote TCM research and be beneficial to its modernization in terms of extending the application of modern methods in the assessment of TCM

Table 1 Comparison of analytical techniques.

	Advantages and problems
LC [7, 96–99]	<ul style="list-style-type: none"> ▶ Low cost ▶ Easy to use ▶ Highly sensitive ▶ Not limited by sample volatility and stability ▶ Favorable separating power
NMR [100]	<ul style="list-style-type: none"> ▶ Noninvasive and nondestructive for samples ▶ Quantitative and simultaneous detection unbiased for any molecules ▶ High throughput ▶ Produces rich, dynamic molecular information ▶ Requires little or no sample preparation ▶ Good resolution and reproducibility
GC [7, 101]	<ul style="list-style-type: none"> ▶ High sensitive detection for almost both volatile chemical and nonvolatile compounds ▶ Has more peak capacity and can accommodate more complex mixtures ▶ Unsuitable for nonvolatile and thermally unstable compounds
CE and HPCE [7, 102]	<ul style="list-style-type: none"> ▶ High speed and short analysis time ▶ Less sample and solvent consumption ▶ Appropriate for complex samples ▶ Lower operating cost ▶ Lower sensitivity than HPLC
MS [15, 103, 104]	<ul style="list-style-type: none"> ▶ Realize identification and quantification of volatile and thermally stable components ▶ Used for ionization of polar to nonpolar components ▶ Associates with LC overcoming problems ▶ Being destructive ▶ Requiring preknowledge about samples ▶ High recurrent expenditures
UV [105]	<ul style="list-style-type: none"> ▶ Easy to be applied ▶ Limited use for compounds without UV absorption ▶ Lacks specificity
IR [106–108]	<ul style="list-style-type: none"> ▶ Potential to use vibrational spectroscopy ▶ Lacks specificity ▶ Signal overlapping

safety, assisting in the formulation of TCM safety norms, and establishing international standards [7].

Data Processing Methods

The progress of metabonomics research will be illustrated. First, the proposed TCM component should be extracted. Second, analytical tools should be applied. Third, the chemical profile should be obtained. Combining data under a multivariate data analysis, validating models, molding, and applying diagnostic tools are the consecutive steps [10] supported by chemometrics and mathematical statistics.

Chemometrics are basically classified into two main categories: pattern recognition methods (unsupervised and supervised), when a qualitative evaluation is involved, and multivariate calibration for quantitative purposes. Data resulting from metabonomics-based work are typically high-dimensional data, requiring MVDA methods for interpretation. Most metabonomics data analysis methods are based on the classification of samples into different groups (e.g., by treatment or genotype), both via supervised (e.g., discriminant function analysis or artificial neural networks) and unsupervised data analysis methods (e.g., PCA or HCA) [11]. It is also possible to use MVDA to conduct regression

modeling between two blocks of data, usually denoted as X and Y. In metabonomics-based NP studies, X may represent signals from different metabolites present in plant extracts sampled at regular time intervals, while Y represents responses (e.g., the quality of product, bioactivity, or yield). The model then can be used to predict Y from X, which is achieved through new observations. The most common MVDA method employed for this type of modeling is the PLS method [12]. Recently, OSC, OPLS, and O2PLS were utilized [13, 14]. The specific methods include SIM-CA, PLS-DA, kNN, and ANN. Moreover, PCA and HCA are widely used in metabonomics research.

Model validation consists of cross validation, permutation testing, and external validation [10]. There are two significant procedures: noise filtration and peak matching [15]. Nonlinear noise filtration is extensively employed, substituting a point with the average of the surrounding points so as to filter noise successfully [16]. Matched filtration is another method setting up a standard mode of a peak and comparing its width; a narrower peak is regarded as noise [17]. Andreev et al. [18] developed MEND, improving the identification function. As for peak matching, identifying the retention time of the internal standard substance under the same conditions is the main method employed.

The main diagnostic tools applied include score plots, loading plots, VIP, DModX, and regression coefficients [10]. MetExtract, a new software tool for the automated comprehensive extraction of metabolite-derived LC/MS signals in metabonomics research, was recently reported in the literature [19].

Bioactivity and Profiles

The main research methods of metabonomics are metabolomic fingerprinting and metabolomic profiling analysis. Metabolomic profiling can be divided into two parts, targeted and nontargeted metabolic profiling analysis.

The targeted metabolic profiling aims to search one biomarker. Several components were usually chosen as marker compounds to assess the quality. These biomarkers were proved to be constituents that discriminate the different species, different parts, different cultivated geographic regions, different planting periods, and the processing products. In the present paper, the bioactivities of the biomarkers were obtained via PubChem (<http://www.ncbi.nlm.nih.gov/pccompound>) and related literatures.

However, these few selected markers sometimes are not unique to a particular herb since they might be present in many plants belonging to various families. In addition, the selection of suitable markers is sometimes difficult and subjective. Furthermore, adulterators are continuously trying to develop ways to make their products' chemical profile similar to the authentic medicinal herbal product. Under these specific circumstances, the marker approach, on the one hand, is unable to confirm the identity of a specific plant. On the other hand, the influences of the other inner chemicals present may be ignored. Therefore, in some cases, its use may be inappropriate for quality control purposes [20, 21]. The objective of "nontargeted" analysis is to describe metabolic events by determining all detectable metabolites [22]. Of the various profiling techniques, nontargeted analysis using UPLC–MS is a promising tool for investigating the diversity of phytochemicals [23]. Thus, it is believed that nontargeted metabolic profiling analysis will play an important role as an effective tool in terms of high-throughput elucidation of metabolic phenotypes.

Identification of Traditional Chinese Medicine Components

The identification of traditional Chinese medicine components plays a key role in ensuring the safety and effectiveness of clinical treatments. The quality and contents of the active constituents in herbs are highly variable depending on the species, parts of the plant, cultivated geographic region, and planting period involved. Adulterants are assumed to be distinguishable from plant material and play the role of challenging substances. Therefore, during large-scale production, the identification of these components is of vital significance. Moreover, many applications employed in the development of metabolic fingerprinting, which will be explained below, using appropriate analysis methods coupled with multivariate analysis, have been investigated and applied to discriminate between closely related plant species in performing quality control assessments of herbal drugs and to identify their different geographic origins. In addition, analyzing components is a robust way to control plant quality.

Here, the identification of TCM components can be divided into four categories: the various species and adulterants, the different parts of herbs, the planting period and the cultivated geographic region.

Identification of species and adulterants

Different species may contain approximately the same components, while the contents of these components vary, which influence the therapeutic effect. Two leguminous plants, *Astragalus membranaceus* (Fisch.) Bge. var. *mongholicus* (bge.) Hsiao and *Astragalus membranaceus* (Fisch.) Bge, are important medicinal herbs that share great similarities regarding their morphology, chemical constituents, and genomic DNA sequences. The identification of different medicinal species directly affects their pharmacological and clinical effects. Amplified AFLP-based genetic fingerprinting and GC-TOF/MS-based metabolic fingerprinting were used to successfully discriminate between the two species. The differences in some soluble sugars, fatty acids, proline, and polyamine reflected the plants' adaptation to different growth environments. Using multivariate and univariate statistical analyses, three AFLP markers and eight metabolites were identified as candidate DNA and metabolic markers to distinguish between the two herb materials [24]. In another study, metabolite profiling of five medicinal *Panax* herbs, which included *P. ginseng* (Chinese ginseng), *P. notoginseng*, *P. japonicus*, *P. quinquefolium* L., and *P. ginseng* (Korean ginseng), was performed using UPLC-QTOFMS and multivariate statistical analysis techniques. PCA of the analytical data showed that the five *Panax* herbs could be separated into five different groups of phytochemicals [25]. HPLC fingerprinting was used for comparison of three closely related species of Pericarpium Citri (*Citrus reticulata* 'Chachi', *Citrus reticulata* 'Dahongpao', and *Citrus erythrosa* Tanaka), and PLS-DA identified hesperidin, tangeretin, and nobiletin as potential biomarkers for their classification [26]. In a similar case, PCA and HCA as well as SIMCA and a BP-ANN were applied to identify and distinguish *Epimedium wushanense* and *Epimedium koreanum* based on their secondary metabolites. The SIMCA method failed to identify one sample, whereas BP-ANN precisely predicted the whole test set [27]. PCA was able to discriminate between ten *Aristolochia* species on the basis of their essential oil profiles, showing that 2 h of hydrodistillation produce the best outcome when the oils are used for discriminating between species [28]. Coincidentally, Sun et al. [29] drew on the same plant. In the Chinese Pharmacopoeia 2010, only two *Aconitum* species are recorded. One is the root of *Aconitum kusnezoffii* Reichb., namely "Caowu" in Chinese. The other species was *Aconitum carmichaelii* Debx. Two herbal drugs are derived from this species. The two species were distinguished successfully using UPLC-QTOF-HDMS, combining with PCA and S-plot. Moreover, a PCA score plot clearly demonstrated discrimination between *Artemisia annua* and *Artemisia afra* on the basis of phenylpropanoids (caffeic acid, chlorogenic acid, dicaffeoyl quinic acid, and ferulic acid) [30]. Spectral fingerprinting via NIR has been utilized for the rapid identification and counterfeit detection of *Eleutherococcus senticosus*, and PCA, DA, SIMCA, and PLS-DA were found to allow good discrimination between *E. senticosus* and other herbs both related to and not related to the Araliaceae family [31]. PCA has been successfully applied for distinguishing *Angelica sinensis* from related Apiaceae (syn. Umbelliferae) herbs based on complete HPLC fingerprints [32]. The same biomarkers were recognized by PLS-DA for the discrimination of authentic Pericarpium Citri from commercial samples, mixed peel samples, and other citrus peels [33]. ¹H-NMR spectroscopy and multivariate data analyses were applied to discriminate two *Bupleurum* species (*B. chinense* and *B. scorzonifolium*) and to explore the influences of habitat and culture methods on the quality of Radix Bupleuri plants based on their metabonomic profiles [34]. The quality of Radix Bupleuri plants was evaluated via HPLC-ELSD analysis and HPTLC based on analysis of their principal bioactive components (saikosaponins). The acquired data were processed using ANNs and kNN to distinguish between different species of the genus [35]. DART-MS provides a novel mass spectrometric ion source by producing [M + H]⁺ molecular ion species. In analyses of *Glycyrrhiza inflata* Batalin, the peak at *m/z* 339 originates mainly from the [M + H]⁺ of LA, a species-specific compound. These results indicate that *G. inflata* can be differentiated from the other two species based on detection of LA peaks using DART-MS analysis [36]. In addition, chromatographic fingerprinting via GC-MS coupled with SA and PCA has been undertaken for discriminating *Scutellaria barbata* D. Don from adulterants. The results showed that the samples could be identified based on differences between the samples and various adulterants [37]. Similarity analysis and HCA were applied for the first time to identify and distinguish genuine *Aconitum kusnezoffii* from its adulterants, which demonstrated the feasibility of linking the HCA approach to chemotaxonomic analysis on the basis of the presence of alkaloids [38]. To discriminate and assess the quality of *Curcuma phaeocaulis*, *C. kwangsiensis*, and *C. wenyujin* from different ecotypes, a metabonomics analysis was carried out via GC-MS coupled with multivariate statistical analysis. Characterization of phytochemicals in essential oils was performed by automated matching to the MS library and comparison of their mass spectra, which discriminated among the different plant parts [39]. *Curcuma* plants, such as *Curcuma wenyujin* Y.H. Chen et C. Ling and *Curcuma longa* L., were also distinguished successfully via HPLC-DAD-MS [40].

The choice of different parts of herbs determines the resulting curative effect, which is the purpose of therapy. The contents of the active components of diverse plant parts were identified. In the long history of the use of medicinal plant preparations, different plant parts have been regarded as different drugs. R. Jurišić Grubešić et al. [41] identified the variation in total polyphenol contents, employing Folin-Ciocalteu's reagent, between different parts of *Plantago* plants (leaves: up to 10.15%; stems: up to

Identification of different medicinal parts of herbs

The choice of different parts of herbs determines the resulting curative effect, which is the purpose of therapy. The contents of the active components of diverse plant parts were identified. In the long history of the use of medicinal plant preparations, different plant parts have been regarded as different drugs. R. Jurišić Grubešić et al. [41] identified the variation in total polyphenol contents, employing Folin-Ciocalteu's reagent, between different parts of *Plantago* plants (leaves: up to 10.15%; stems: up to

4.34%; and flowers: up to 5.56%). The content of tannins in stems ranged from 0.28% to 1.00%, while leaves and flowers contained tannins at concentrations of 2.26% and 2.21% based on UV-Vis spectrophotometry.

Metabolite profiling of different parts of *Panax notoginseng* was carried out using UPLC-ESI-MS and multivariate statistical analysis. PCA of the UPLC-ESI-MS data showed a clear separation of the compositions among the flower buds, roots, and rhizomes of *P. notoginseng*. The saponins accounting for these variations were identified based on corresponding loading weights and were further verified based on the accurate mass, tandem mass, and retention time of available standard saponins using UPLC-QTOF-MS [42]. Moreover, each extract from 24 mulberry leaf samples, divided into six locations from the tip of the stem in each of four strains, was analyzed via pattern recognition methods, including PCA and SIMCA. The 24 extracts from mulberry leaves showed independent spectra in $^1\text{H-NMR}$ analyses [43]. *Aconitum carmichaelii* Debx., another example for the application of plant metabolomics in the discrimination of different parts of herb plants, was studied by Sun et al. [29]. The mother root is named “Chuanwu”, while the daughter or lateral root of *Aconitum carmichaelii* Debx. is known as “Shengfuzi”. Shengfuzi has been prescribed more frequently than Chuanwu to treat rheumatic diseases. The analytical techniques, UPLC-Q-TOF-HDMS, as well as the data processing methods, PCA, and S-plot, were the main measures in this study.

Differentiation of distinct cultivated geographic regions

The environments of the cultivated geographic regions of medicinal plants, including their temperature, humidity, soil, and climate, are determinant factors. Therefore, the cultivated geographic region influences the growth of herbs. Wei-Jun Kong et al. [44] utilized UPLC-PAD analysis to examine the five active alkaloids in *Rhizoma Coptidis Chinensis*, successfully grouping the plants in accordance with their province of origin. Moreover, the LS-SVM, RBF-ANN, PLS-DA, and kNN methods were applied for the classification of *Rhizoma Corydalis*, and in general, no statistically significant differences were found between these four methods. NIRS was used to identify *Rhizoma Corydalis* plants from two different geographical origins [45]. Another example of the application of the HCA technique was its use for the classification of *Isatis indigotica* roots collected from different regions based on HPLC fingerprinting [46]. *Ganoderma lucidum* samples from different cultivated geographic regions were evaluated using HPLC fingerprinting. The HCA, PCA, PLS-DA, and SIMCA techniques were employed to classify samples in accordance with their province of origin [47]. In addition, the essential oils of the Cinnamon Cortex specimens obtained from different localities have been analyzed via GC-MS [48]. Furthermore, the volatile oils of *Artemisia capillaris* Herba from different locations were investigated through GC-MS to develop a characteristic fingerprint of this raw herb [49]. The discrimination of *Schizonepeta tenuifolia* Briq. from different origins has also been achieved via PCA and HCA, which classified the samples into two main groups on the basis of five marker compounds [50]. It is also worth noting that the combination of NIR spectroscopy with DA and PLS-DA analysis was applied in geographical origin discrimination for *Radix Scutellaria Baicalensis* [51]. A nontargeted procedure was applied for $^1\text{H-NMR}$ spectroscopic fingerprinting of extracts from *Rhodiola rosea* rhizomes for pattern recognition analysis and identification of secondary metabolites responsible for differences in sample composition. For this purpose, plants from three different

geographic areas (the Swiss Alps, Finland, and the Altai region in Siberia) were investigated [52]. Furthermore, quantitative estimates of the major isoflavones in *Pueraria lobata* were produced, and the studied samples were classified through PCA based on the amounts of puerarin, daidzin, daidzein, and genistin present [53]. Metabolite fingerprinting was applied in an attempt to evaluate the quality of dried *Angelica acutiloba* roots. An enhanced understanding of the dominance of the relationship of the cultivation area with the evaluated quality was conceptualized and applied to the construction of a PLS-DA classification model, which provided the basis for accurate and reliable predictivity [54]. Additionally, PCA was performed using the data generated through HPLC-DAD-ELSD analysis for quality control of *Polygala japonica* from different localities in China [55]. Recently, Suzuki et al. [56] classified *Sophora flavescens* grown in Japan and China via NMR.

Differentiation of distinct planting periods

The planting period is also a vital factor in the quality of a crude drug due to the duration over which a plant obtains nutrition from the soil.

An UPLC-Q-TOF-MS-based metabolomic technique was applied for metabolite profiling in 60 *Panax ginseng* samples aged from 1 to 6 years [57]. Ginseng is an important herbal resource worldwide, and adulteration or falsification of the cultivation age has been a serious problem for ginseng in the commercial market. In this study, ginseng roots cultivated for 2–6 years under good agricultural practices standard guidelines were analyzed via NMR-based metabolomics techniques using two solvents [58]. Moreover, it has been demonstrated that July might be the best harvest time for *Pericarpium Citri Reticulatae Viride*, while November and December are better for *Pericarpium Citri Reticulatae*. Furthermore, hesperidin, nobiletin, and tangeretin were screened as chemical markers based on PCA loadings. The HPLC-HELP-PCA strategy has shown potential in the optimization of harvest times [26]. Recently, Xue et al. [59] utilized GC-MS to investigate the flower buds of *Tussilago farfara* in different development stages. Collectively, medicinal herbs from different species and the different parts of the same plant usually exhibit different efficacy, pharmacological actions, and clinical indications due to the significant differences in the types and quantity of the constituents. The species diversity seems to be a significant factor to influence the quality assessment. In addition, chemical constituents of the same plant may be various due to different cultivation areas, climatic conditions, and cultivation ages. For example, ginseng of cultivation ages from 4 to 6 years is the most demanded ginseng in the market. However, age and cultivation areas can hardly be determined by the herb's physical appearance alone. Accordingly, confused clinical application led to the consumption of incorrect forms of plant material, improper use, and undesirable effects. Hence, an effective method applied in quality control is urgently demanded for the identification step of medicinal herbs. Since some samples share similarities in morphology but with subtle variations in certain ingredients, metabolomics can provide a platform to use analytical techniques coupled with multivariate statistics for the differentiation of these complex samples. Metabolomics information not only assist in the establishment of a deeper understanding of the complex interactive nature of plant metabolic networks and their responses to environmental change but also provide unique insights into the fundamental nature of plant phenotypes in relation to development, physiology, tissue identity, resistance, biodiversity, and so on. To make them

clear, the plant material, analysis technique, chemometrics methods, biomarkers, and bioactivity aspects are summarized in **Table 2**.

Processing

Processing is the second step in the production of Chinese medicinal preparations. TCM-specific production steps include storing, washing, rinsing, drying, remoistening, and cutting, and eventually, unique processing techniques, such as stir frying, steaming, or calcining are performed to satisfy different clinical therapeutic requirements. First, the effect of processing is thought to enhance the therapeutic efficiency in *Sophora japonica* L. [60]. Drug processing can also weaken the structure of plants so that the active components can be extracted easily. Additionally, additives react with the compounds present in plants generating new components dissolved in solvents. For instance, alkaloids dissolve in acidic solvents, so vinegar is widely applied to crude drugs to enrich alkaline substances. Second, processing has been reported to reduce the toxicity of the crude drugs, as described with Fuzi [61]. As evidence has accumulated, it has been shown that poisonous protein is one of the causes of accidental side effects. As proteins are thermo-sensitive, crude drugs should be subjected to heat treatment. Third, the expansion of applications is another important unexpected impact. Finally, after processing, the generation of several new compounds has been reported [62]. However, the dosage of additives and the time of heat treatment should be considered during processing, as they are the major factors that affect quality control.

In the large-scale production, it is difficult to guarantee the purity of all products. Researchers spend long periods finding solutions to quality control. The development of metabonomics has provided a necessary way to understand cellular responses to mutations at all levels of gene products [63]. In recent years, a wide range of metabonomic analytical techniques have been implemented in research on TCM [64]. Several cases illustrate the possibilities of the application of metabonomics in quality control during the processing of TCM materials. Ginseng has been employed in TCM for over two thousand years and is now widely used around the world as an elixir [63]. In Asia, there are two types of ginseng that are commonly found in the herbal medicine market: WG and RG. In the practice of traditional Chinese medicine, WG and RG have been used for different purposes. WG is traditionally produced via sun drying of fresh ginseng, and RG is manufactured by steaming fresh ginseng at 95–100 °C for 2–3 h and then drying it. WG is used to “supply qi and promote the production of body fluids” as well as enhance physical fitness and disease resistance, while RG has a “warming effect” and is used for “boosting yang” and replenishing vital essence [64]. Ginsenosides Rb1, Rb2, Rc, Rd, Rg1, and Re are the major constituents of both WG and RG, while ginsenosides Rg3, Rg5, Rg6, Rh1, Rh2, Rk1, Rs3, and F4 are known to be unique constituents of RG [65–72]. These unique ginsenosides found in RG have been reported to be converted from the ginsenosides found in fresh ginseng after steaming [70, 73, 74]. In one study, ginseng was processed under temperatures of 100, 140, and 180 °C, with or without vinegar; the duration of exposure to each temperature was 10, 30, and 50 min, respectively, and there was a clear separation in the score plots obtained for the various treatment conditions. The major compounds contributing to the separation of 50% methanol extracts of vinegar-treated ginseng subjected to various pro-

cessing conditions were valine, lactate, alanine, arginine, glucose, fructose, and sucrose. As the temperature increased, the valine, arginine, glucose, fructose, and sucrose concentrations decreased, whereas lactate, glucose, and fructose increased in the vinegar-treated samples compared to non-vinegar-treated samples [62]. Moreover, UPLC/TOFMS had been demonstrated to be a powerful tool for use in herbal metabonomics to discriminate differentially processed herbs, such as raw and steamed *P. notoginseng* [75]. An UHPLC-TOF-MS-based metabonomics platform coupled with PCA and PLS-DA was developed for *Panax notoginseng* to establish a correlation between the duration of steaming and the maximum production of bioactive ginsenosides [76]. A similar study was performed to determine chemical markers for discriminating between raw and processed *Radix Rehmanniae* samples [77]. In addition, the three types of products obtained from the processing of *Ligustrum lucidum* fruits have been distinguished, which correspond to steam treatment processing products, vinegar treatment processing products, and the fruits processed with wine. There are differences in metabolite profiles among the crude and different types of processed fruits of *L. lucidum*. Ligustaloside B was identified as a chemical marker for such variations, and its contents in crude *L. lucidum* specimens were found to be significantly higher than in processed samples. This study indicated that UPLC-QTOF-MS coupled with multivariate statistics is able to provide quality control for the crude and processed fruits of *L. lucidum*, and these results provide the basis for determining the appropriate mechanism of processing [78]. The products of the processing of *Polygala Radix* were also successfully distinguished [79]. One study was designed to perform a comprehensive metabonomics analysis of Fuzi and its processed products, Yanfuzi, Heishunpian, and Baifupian, via UPLC-Q-TOF-HDMS combined with pattern recognition methods. Differences in the metabolic profiles of Fuzi and its processed preparations were clearly observed based on PCA of the obtained MS spectra. Significant changes in 19 metabolite biomarkers were detected in the Fuzi samples and the three preparations [61]. Similarity analysis and PCA were applied to address the issue of the various quality changes that occur during the process of toasting *Fructus Xanthii* supplied by different producing areas. A high similarity was observed between different samples, which indicates that the proportion and distribution of the components in most extracts of *F. xanthii* show a high level of consistency [80]. HPLC fingerprints together with metal profiles were employed to assess the quality control procedures applied to *Atractylis chinensis*. A separate data matrix and combined data matrices were analyzed via PCA, kNN, and LDA. The PCA results from the combined data matrices indicated that the samples were discriminated on the basis of the applied processing methods. Within each group, the samples were reasonably well grouped according to their geographical origin and classification using kNN, and LDA results supported the PCA results [81].

As the raw and processed forms of herbs have different pharmacological actions, it is pertinent to administer the correct form of herb to avoid any undesirable consequences. Even the duration of the processing procedure, the processing adjuvants, and its dosage arouse the subtle changes in the contents of compounds. Therefore, it is of paramount importance to characterize the specific forms. The collection of quoted literature data is shown in

Table 2.

Table 2 Application of plant metabonomics in TCM.

TCM materials	Purpose	Analysis techniques	Chemometrics methods	Biomarkers	Bioactivity	Ref.
Two <i>Astragalus</i> plants	A1	GC-TOF/MS	AFLP	Three AFLP markers, eight metabolites	Antiperspirant and antidiuretic	[24]
Five <i>Panax</i> plants	A1	UPLC-QTOF-MS	PCA	Ginsenoside Rf, Rb, Rb2, 20(s)-pseudoginsenoside, F11	Antineoplastic, hypolipidemic agents*	[25]
<i>Panax notoginseng</i>	A2	UPLC-QTOF-MS	PCA	Saponins	Prevention and treatment of cerebrovascular diseases, immune regulation, hepato protection, anticarcinogenesis, neuroprotective effect	[42]
<i>Panax ginseng</i>	A4	UPLC-QTOF-MS	PCA-HCA RF PAM PLS-DA	\	\	[57]
<i>Panax ginseng</i>	B	UPLC-QTOF-MS	PCA	\	\	[62]
<i>Panax notoginseng</i>	B	UPLC-QTOF-MS	PCA, PLS-DA	\	\	[76]
<i>Panax ginseng</i>	A4	NMR	PCA, PLS-DA	Amino acids, organic acids, sugars	Cardiovascular control of blood pressure	[58]
Three tangerine peels	A1, 4	HPLC-DAD	PCA, HELP	Hesperidin, tangeretin, nobiletin	Protective effect on myocardial ischemia	[26]
<i>Mallotus</i> plants	A1, 2, 3	LC-MS	PLS-DA	Senkyunolide A	Antioxidants	[33, 109]
Two <i>Epimedium</i> plants	A1	HPLC	PCA, HCA, SIMCA, BP-ANN	Flavonoids	Influence on sexual function, anti-aging, effect on immune system, anti-inflammatory, antitussive, expectorant, antiasthma	[27]
Rhizoma <i>Coptidis</i>	A3	UPLC	SA, HCA, PCA	Berberine, coptisine, palmatine, jateorrhizine, epiberberine	Efficacy of suppressing fever, dispelling dampness, removing toxicosis and anti-microbes*	[44]
Five <i>Bupleurum</i> plants	A1	¹ H-NMR HPLC-ELSD HPTLC	ANNs, kNN	Saikosaponins	Anti-inflammatory, antineoplastic, immunosuppressive agents*	[34] [35]
Ten <i>Aristolochia</i> plants	A1	GC-MS	PCA	Essential oils	Abortifacients, stomachics, antiotheriacs, antiasthmatics, expectorants, slimming therapies	[28]
Two <i>Artemisia</i> plants	A1	NMR	PCA	Polar components	Antiplasmodial	[30]
<i>Artemisia capillaris</i> herba	A3	GC-MS	EWOP, FSMWEFA	Essential oils	Choleretic, anti-inflammatory and diuretic agent in the treatment of epidemic hepatitis	[49]
Three <i>Curcuma</i> plants	A1	GC-MS	PCA, PLS-DA	Essential oils	Against skin diseases, colic inflammatory disorders, insect repellants, antimicrobial	[39]
Two <i>Curcuma</i> plants	A1	HPLC-DAD-MS, GC-MS	PCA	Curcumin, demethoxycurcumin, bisdemethoxycurcumin, dihydrocurcumin, ar-turmerone, α , β -turmerone, zingiberene	Against skin diseases, colic inflammatory disorders, insect repellants, antimicrobial, antidiabetic medications	[40]
Rhizoma <i>Corydalis</i>	A3	NIRS	WT, LS-SVM, PLS-DA, KNN	\	\	[45]
<i>Eleutherococcus senticosus</i>	A1	NIRS	PCA, DA, SIMCA and PLS-DA	\	\	[31]
<i>Angelica sinensis</i>	A1	HPLC	PCA	Senkyunolide A	Treatment of gynecological diseases	[32]
<i>Plantago</i> L.	A2	UV	UPGMA, PCA	Polyphenols, tannins	Diuretic	[41]
Three <i>Glycyrrhiza</i> plants	A1	DART-MS	\	Licochalcone A	Antimicrobial activity, antiplasmodial activity, antileishmanial activity*	[36]
Mulberry leaf	A3	¹ H-NMR	PCA, SIMCA	\	\	[43]
<i>Isatis indigotica</i>	A3	RP-HPLC	HCA	Indirubin, indigotin	Anti-inflammatory, inhibition of leucine-rich repeat kinase-2, proliferative and androgenic effects*	[46]
Cortex <i>Cinnamomi</i>	A3	GC-MS	IOP, HELP, SFA, OPR	Essential oils	Antimicrobial activities	[48, 110]
<i>Ganoderma lucidum</i>	A3	NIRS	HCA, PCA, PLS-DA, SIMCA	Triterpenoidsaponins, polysaccharides	Inhibitors of the <i>in vitro</i> human recombinant aldose reductase*	[47]

continued

Table 2 Continued

TCM materials	Purpose	Analysis techniques	Chemometrics methods	Biomarkers	Bioactivity	Ref.
<i>Schizonepeta tenuifolia</i> Briq.	A3	GC-MS	PCA, HCA	2-Hydroxy-2-isopropenyl-5-methyl-, <i>cis</i> -pulegone oxide, menthone, pulegone, cyclohexanone, schizonal	Antifungal properties, decrease in ambulation, and increase in pentobarbital-induced sleeping time	[50, 111]
<i>Scutellaria barbata</i> D. Don	A1	GC-MS	SA, PCA	86 Compounds	Antimicrobial, protecting liver and biliary	[37]
Radix <i>Scutellaria Baicalensis</i>	A3	NIR	DA, S-DA	\	\	[51]
<i>Rhodiola rosea</i> rhizomes	A3	¹ H-NMR	PCA	Salidroside, rosavin	Protective effects on LPS-induced acute lung injury*	[52]
<i>Pueraria lobata</i>	A3	RRLC	PCA	Isoflavonoids	Antioxidant, estrogen-like effect, and treatment of osteoporosis	[53, 112] [113]
<i>Angelica acutiloba</i>	A3	Pyrolyser-coupled (PY-GC-MS)	PCA, PLS-DA	\	\	[54]
<i>Polygala japonica</i>	A3	HPLC-DAD-ELSD	PCA, COW	\	\	[55]
<i>Sophora flavescens</i>	A3	NMR	PCA	Kurarinol	Relative inhibition or inhibition of phosphodiesterase 3,4,5*	[56]
<i>Tussilago farfara</i>	A4	GC-MS	PCA	Fifty-four metabolites were identified, including 35 polar metabolites and 19 nonpolar compounds	\	[59]
<i>Atractylis chinensis</i> DC	B	HPLC	PCA, kNN, LDA	\	\	[81]
<i>Aconitum kusnezoffii</i>	A1	HPLC	HCA	Mesaconitine, aconitine, hyaconitine	Voltage-gated sodium channel agonists, immunologic*	[38]
Fuzi	B	UPLC-Q-TOF-HDMS	PCA, PLS-DA, OPLS-DA	Aconitine, mesaconitine, hyaconitine, deoxyaconitine, 10-OH-mesaconitine	Voltage-gated sodium channel agonists, immunologic*	[61]
Two <i>Carmichaelii</i> plants	A1, B	UPLC-Q-TOF-HDMS	PCA, S-plot	22 Types of alkaloids	Voltage-gated sodium channel agonists, immunologic*	[29]
Radix <i>Rehmanniae</i>	B	UHPLC-TOFMS	PCA, OPLS-DA	Leonuride, its isomer, 5-GGMF	Antitumor, antidiabetic, neuro-protective*	[77]
<i>Ligustrum lucidum</i>	B	UPLC-QTOF-MS	PCA	Ligustaloid B	Antidiabetic, antioxidant	[78, 114]
Fructus <i>Xanthii</i>	B	GC-MS	PCA	14 Polar compounds	Treatment of cramping and numbness of the limbs, ulcer, sinusitis, catarrhs and pruritus	[80]

* Activity obtained from PubChem (<http://www.ncbi.nlm.nih.gov/pccompound>). A1: Identification of species and adulterants; A2: identification of different medicinal parts of herbs; A3: differentiation of distinct cultivated geographic regions; A4: differentiation of distinct planting periods; B: processing

Pharmaceuticals

The last step in the production of Chinese medicinal preparations, obtaining plant extracts, can also be subjected to quality control using metabonomic profiles. The extract is a contributing factor to the quality and toxicity of the drug produced. However, a prerequisite is that the extracts of intermediate products that are to be analyzed are well documented with regard to the production steps they have been subjected to. Bioactive compounds may be identified if it is possible to obtain or generate extracts of different materials from the same plant species that are highly variable in bioactivity. PCA may then be used to discriminate the chemical fingerprints of the extracts in a way that separates them by their activity or by spatial origin, and relevant chemical compounds can subsequently be deduced from their contributions to the respective fingerprints [82, 83]. The majority of TCM products for oral use are applied as water decoctions [84]. Other oral prepa-

arations include macerates in aqueous ethanol and powdered drugs suspended in water or prepared in pills, with honey, water, or rice gruel as an excipient [85, 86]. An HPLC fingerprinting analysis was developed to assess the quality and comparative contents of cinnamon bark and cinnamon twig components. PCA and PLS-DA allowed good discrimination of these samples, and cinnamaldehyde was found to be the most abundant marker component [87]. Several examples are found in the literature of utilizing HPLC together with different chemometric methods for the analysis of complex mixtures, including the resolution of HPLC fingerprints of complex, many-component substances found in Huoxiang Zhengqi tincture samples from a batch from a given manufacturer, or from different producers [88]. Another example of investigating complex mixtures is the analysis of nine bioactive compounds from a Yiqing preparation which is composed of three TCMs, to assess the consistency of the quality among 12 manufacturers based on SA. The results showed that

HPLC fingerprinting could serve as the first tool for revealing the consistency of the quality of Yiqing via similarity comparisons [89]. LC-LTQ-Orbitrap MS was applied for the simultaneous identification and quantification of multi-constituent Xin-Ke-Shu, a TCM preparation [90].

It is complicated during the pharmaceutical process. On the one hand, the pharmaceutical excipients hold back the analysis. On the other hand, a Chinese patent drug usually consists of many sorts of herbs; the constituents' analysis is a bottleneck in quality control. Plant metabonomics, a platform aimed at the complex ingredients, can help to better understand the nature of these problems.

Conclusions and Perspectives

Plant metabonomics can be applied in the discrimination, processing, and pharmaceutical preparation steps of TCM products, which represent the entire production process (Table 2). As the first step in the production of Chinese medicinal preparations, the identification of plant varieties used, involving species, parts of the herbs, cultivated geographic region, and planting period, is of great significance for ensuring the safety and effectiveness of clinical treatment since the quality and contents of the active constituents depend on these factors. Adulterants are assumed to be distinguishable from plant materials and play the role of challenging substances. The processing, as the second step in production, appears to be of significance in clinical applications and has been proven to satisfy the requirements of therapeutics. As different forms of TCMs show different pharmacological actions, it is pertinent to administer the correct form of herbs to avoid any undesirable consequences. Therefore, it is of paramount importance to characterize the specific form of TCM. Metabonomics can also be applied to control the content of extracts, which is crucial to the pharmaceutical production. The complexity and diversity of the components of TCM preparations make qualitative analyses difficult. In the pharmacopeia, single-component analysis is used on TCM mostly. Nevertheless, it lacks representativeness. In contrast to single-component analyses, metabonomics achieves comprehensive scans, addressing some of the shortfalls of single-component analysis. As shown by the three points we have just illustrated, plant metabonomics can play a vital role in quality assessments during the large-scale production of TCM preparations.

To facilitate the application of plant metabonomics in the quality assessment, on the one hand, we are supposed to utilize the new and effective techniques so that they will support metabonomics studies adequately. On the other hand, exploring potential research points of metabonomics in TCM seems necessary. With the appearance of RRLC and UFLC, the methods of analysis come to a new era. Shorter analysis time and the more efficient separation are the advantages of these methods [91]. MAS-NMR, another sharp technique, enhances the resolution of solid samples and plays an important role in the overlap of peaks [92]. Cristina Dao-lio et al. [93] applied MAS-NMR classifying commercial catuaba successfully. An LC-MS-NMR platform was demonstrated, which combines two innovations in microscale analysis, nanoSplitter LC-MS and microdroplet NMR, for the identification of unknown compounds found at low concentrations in complex sample matrices as frequently encountered in metabonomics or natural product discovery [94]. Therefore, in the future, employing new and effective chromatographic or spectroscopic techniques for

metabonomics studies seems to be an important tendency. What concerns exploring potential research points, in recent years, metabonomics was reported to the study of pharmacology in some terms. Employing a metabonomics platform, Yiqing Lin et al. [94] identified the active cyanobacterial metabolite. Kashif Ali et al. [95] also managed to screen the anti-TNF α activity in crude extracts of grapes and other berries by NMR spectroscopy and chemometric. Using this approach, compounds related to activity can be identified without extensive and elaborate chromatographic separation, and it thus allows rapid identification of extracts with biological activity. Moreover, screening the active compounds and effective parts seems to grow into a vital process in future studies. As a consequence, the results of quantitative assays will become more instructive and convincing. In a word, with the development of analysis methods and the exploration of potential research points, the application of metabonomics in TCM quality assessment tends to become more prevalent and considered in the future.

Acknowledgements

This study was financially supported by the National Natural Science Foundation of China (Projects No.81001623 and No.30902000).

Conflict of Interest

No conflicts of interest exist.

References

- Nicholson JK, Wilson ID. Opinion: understanding 'global' systems biology: metabonomics and the continuum of metabolism. *Nat Rev Drug Discov* 2003; 2: 668–676
- Hui-Ru T, Yu-Lan W. Metabonomics: a revolution in progress. *Prog Biochem Biophys* 2006; 33: 401–417
- Dong J, Xu L, Cao H, Dai X, Li X, Yang S, Chen Z. A new data processing method for metabonomic and its application in a study of diabetes. *Chin J Magn Reson* 2007; 24: 393
- Lu YH, Hao HP, Wang GJ, Chen XH, Zhu XX, Xiang BR, Huang Q, AJY. Metabolomics approach to the biochemical differentiation of Traditional Chinese Medicine syndrome types of hypertension. *Chin J Clin Pharmacol Ther* 2007; 12: 1144–1150
- Cheng LL, Chang IW, Louis DN, Gonzalez RG. Correlation of high-resolution magic angle spinning proton magnetic resonance spectroscopy with histopathology of intact human brain tumor specimens. *Cancer Res* 1998; 58: 1825–1832
- Cheng L, Ma M, Becerra L, Ptak T, Tracey I, Lackner A, Gonzalez R. Quantitative neuropathology by high resolution magic angle spinning proton magnetic resonance spectroscopy. *Proc Natl Acad Sci USA* 1997; 94: 6408–6413
- Lao YM, Jiang JG, Yan L. Application of metabonomic analytical techniques in the modernization and toxicology research of traditional Chinese medicine. *Br J Pharmacol* 2009; 157: 1128–1141
- Kim N, Kim K, Choi BY, Lee DH, Shin YS, Bang KH, Cha SW, Lee JW, Choi HK, Jang DS. Metabolomic approach for age discrimination of *Panax ginseng* using UPLC-Q-ToF MS. *J Agric Food Chem* 2011; 59: 10435–10441
- Holmes E, Wilson ID, Nicholson JK. Metabolic phenotyping in health and disease. *Cell* 2008; 134: 714–717
- Yuliana ND, Khatib A, Choi YH, Verpoorte R. Metabolomics for bioactivity assessment of natural products. *Phytother Res* 2010; 25: 157–169
- Nobeli I, Thornton JM. A bioinformatician's view of the metabolome. *Bioessays* 2006; 28: 534–545
- Wold S, Sjöström M, Eriksson L. PLS-regression: a basic tool of chemometrics. *Chemom Intell Lab Syst* 2001; 58: 109–130

- 13 Gabriellsson J, Jonsson H, Airiau C, Schmidt B, Escott R, Trygg J. OPLS methodology for analysis of pre-processing effects on spectroscopic data. *Chemom Intell Lab Syst* 2006; 84: 153–158
- 14 Wold S, Trygg J, Berglund A, Antti H. Some recent developments in PLS modeling. *Chemom Intell Lab Syst* 2001; 58: 131–150
- 15 Ceglarek U, Leichtle A, Brügel M, Kortz L, Brauer R, Bresler K, Thiery J, Fiedler GM. Challenges and developments in tandem mass spectrometry based clinical metabolomics. *Mol Cell Endocrinol* 2009; 301: 266–271
- 16 Hastings CA, Norton SM, Roy S. New algorithms for processing and peak detection in liquid chromatography/mass spectrometry data. *Rapid Commun Mass Spectrom* 2002; 16: 462–467
- 17 Danielsson R, Bylund D, Markides KE. Matched filtering with background suppression for improved quality of base peak chromatograms and mass spectra in liquid chromatography–mass spectrometry. *Anal Chim Acta* 2002; 454: 167–184
- 18 Andreev VP, Rejtar T, Chen HS, Moskovets EV, Ivanov AR, Karger BL. A universal denoising and peak picking algorithm for LC-MS based on matched filtration in the chromatographic time domain. *Anal Chem* 2003; 75: 6314–6326
- 19 Bueschl C, Kluger B, Berthiller F, Lirk G, Winkler S, Kraska R, Schuhmacher R. MetExtract: a new software tool for the automated comprehensive extraction of metabolite-derived LC/MS signals in metabolomics research. *Bioinformatics* 2012; 28: 736–738
- 20 Mok DKW, Chau FT. Chemical information of Chinese medicines: A challenge to chemist. *Chemom Intell Lab Syst* 2006; 82: 210–217
- 21 Xie P, Chen S, Liang YZ, Wang X, Tian R, Upton R. Chromatographic fingerprint analysis—a rational approach for quality assessment of traditional Chinese herbal medicine. *J Chromatogr A* 2006; 1112: 171–180
- 22 Li L, Sun B, Zhang Q, Fang J, Ma K, Li Y, Chen H, Dong F, Gao Y, Li F, Yan X. Metabonomic study on the toxicity of Hei-Shun-Pian, the processed lateral root of *Aconitum carmichaelii* Debx. (Ranunculaceae). *J Ethnopharmacol* 2008; 116: 561–568
- 23 Zhang A, Sun H, Wang P, Han Y, Wang X. Recent and potential developments of biofluid analyses in metabolomics. *J Proteom* 2012; 75: 1079–1088
- 24 Duan LX, Chen TL, Li M, Chen M, Zhou YQ, Cui GH, Zhao AH, Jia W, Huang LQ, Qi X. Use of the metabolomics approach to characterize Chinese medicinal material Huangqi. *Mol Plant* 2012; 5: 376–386
- 25 Xie G, Plumb R, Su M, Xu Z, Zhao A, Qiu M, Long X, Liu Z, Jia W. Ultra-performance LC/TOF MS analysis of medicinal *Panax* herbs for metabolomic research. *J Sep Sci* 2008; 31: 1015–1026
- 26 Yi L, Yuan D, Liang Y, Xie P, Zhao Y. Fingerprinting alterations of secondary metabolites of tangerine peels during growth by HPLC–DAD and chemometric methods. *Anal Chim Acta* 2009; 649: 43–51
- 27 Wang L, Wang X, Kong L. Automatic authentication and distinction of *Epimedium koreanum* and *Epimedium wushanense* with HPLC fingerprint analysis assisted by pattern recognition techniques. *Biochem Syst Ecol* 2012; 40: 138–145
- 28 Francisco CS, Messiano GB, Lopes LM, Tininis AG, de Oliveira JE, Capellari Jr. L. Classification of *Aristolochia* species based on GC-MS and chemometric analyses of essential oils. *Phytochemistry* 2008; 69: 168–175
- 29 Sun H, Wang M, Zhang A, Ni B, Dong H, Wang X. UPLC-Q-TOF-HDMS analysis of constituents in the root of two kinds of *Aconitum* using a metabolomics approach. *Phytochem Anal* 2013; 24: 263–276
- 30 Liu NQ, Cao M, Frederich M, Choi YH, Verpoorte R, van der Kooy F. Metabolomic investigation of the ethnopharmacological use of *Artemisia afra* with NMR spectroscopy and multivariate data analysis. *J Ethnopharmacol* 2010; 128: 230–235
- 31 Lucio-Gutiérrez JR, Coello J, Maspoch S. Application of near infrared spectral fingerprinting and pattern recognition techniques for fast identification of *Eleutherococcus senticosus*. *Food Res Int* 2011; 44: 557–565
- 32 Lu GH, Chan K, Liang YZ, Leung K, Chan CL, Jiang ZH, Zhao ZZ. Development of high performance liquid chromatographic fingerprints for distinguishing Chinese *Angelica* from related Umbelliferae herbs. *J Chromatogr A* 2005; 1073: 383–392
- 33 Tistaert C, Dejaegher B, Chataigne G, Van Minh C, Quetin-Leclercq J, Vander Heyden Y. Dissimilar chromatographic systems to indicate and identify antioxidants from *Mallotus* species. *Talanta* 2011; 83: 1198–1208
- 34 Qin X, Dai Y, Liu NQ, Li Z, Liu X, Hu J, Choi YH, Verpoorte R. Metabolic fingerprinting by (1)H-NMR for discrimination of the two species used as Radix Bupleuri. *Planta Med* 2012; 78: 926–933
- 35 Tian RT, Xie PS, Liu HP. Evaluation of traditional Chinese herbal medicine: Chaihu (Bupleuri Radix) by both high-performance liquid chromatographic and high-performance thin-layer chromatographic fingerprint and chemometric analysis. *J Chromatogr A* 2009; 1216: 2150–2155
- 36 Fukuda E, Baba M, Iwasaki N, Uesawa Y, Arifuku K, Kamoe O, Tsubono K, Okada Y. Identification of *Glycyrrhiza* species by direct analysis in real time mass spectrometry. *Nat Prod Commun* 2010; 5: 1755–1758
- 37 Pan R, Guo F, Lu H, Feng WW, Liang YZ. Development of the chromatographic fingerprint of *Scutellaria barbata* D. Don by GC-MS combined with chemometrics methods. *J Pharm Biomed Anal* 2011; 55: 391–396
- 38 Zhao YY, Zhang Y, Lin RC, Sun WJ. An expeditious HPLC method to distinguish *Aconitum kusnezoffii* from related species. *Fitoterapia* 2009; 80: 333–338
- 39 Xiang Z, Wang XQ, Cai XJ, Zeng S. Metabolomics study on quality control and discrimination of three *Curcuma* species based on gas chromatography-mass spectrometry. *Phytochem Anal* 2011; 22: 411–418
- 40 Wu HW. Studies on warm and cold nature of JiangHuang and YuJin based on metabonomics [dissertation]. Beijing: Chinese Academy of Chinese Medical Sciences; 2011
- 41 Grubesić RJ, Vuković J, Kremer D, Vladimir-Knezevic S. Spectrophotometric method for polyphenols analysis: prevalidation and application on *Plantago L.* species. *J Pharm Biomed Anal* 2005; 39: 837–842
- 42 Dan M, Su M, Gao X, Zhao T, Zhao A, Xie G, Qiu Y, Zhou M, Liu Z, Jia W. Metabolite profiling of *Panax notoginseng* using UPLC-ESI-MS. *Phytochemistry* 2008; 69: 2237–2244
- 43 Fukuda E, Yoshida M, Baba M, Uesawa Y, Suzuki R, Kamo O, Tsubono K, Arifuku K, Yatsunami K, Okada Y. Application to classification of mulberry leaves using multivariate analysis of proton NMR metabolomic data. *Nat Prod Commun* 2011; 6: 1621–1625
- 44 Kong WJ, Zhao YL, Xiao XH, Jin C, Li ZL. Quantitative and chemical fingerprint analysis for quality control of *rhizoma Coptidischinensis* based on UPLC-PAD combined with chemometrics methods. *Phytomedicine* 2009; 16: 950–959
- 45 Lai Y, Ni Y, Kokot S. Discrimination of *Rhizoma Corydalis* from two sources by near-infrared spectroscopy supported by the wavelet transform and least-squares support vector machine methods. *Vib Spectrosc* 2011; 56: 154–160
- 46 Zou P, Hong Y, Koh HL. Chemical fingerprinting of *Isatis indigotica* root by RP-HPLC and hierarchical clustering analysis. *J Pharm Biomed Anal* 2005; 38: 514–520
- 47 Chen Y, Xie MY, Yan Y, Zhu SB, Nie SP, Li C, Wang YX, Gong XF. Discrimination of *Ganoderma lucidum* according to geographical origin with near infrared diffuse reflectance spectroscopy and pattern recognition techniques. *Anal Chim Acta* 2008; 618: 121–130
- 48 Gong F, Liang YZ, Xu QS, Chau FT. Gas chromatography-mass spectrometry and chemometric resolution applied to the determination of essential oils in Cortex cinnamomi. *J Chromatogr A* 2001; 905: 193–205
- 49 Guo FQ, Liang YZ, Xu CJ, Li XN, Huang LF. Analyzing of the volatile chemical constituents in *Artemisia capillaris herba* by GC-MS and correlative chemometric resolution methods. *J Pharm Biomed Anal* 2004; 35: 469–478
- 50 Jung JH, Hong J. GC/MS combined with chemometrics methods for quality control of *Schizonepeta tenuifolia* Briq: Determination of essential oils. *Microchem J* 2011; 97: 274–281
- 51 Li W, Xing L, Cai Y, Qu H. Classification and quantification analysis of *Radix scutellariae* from different origins with near infrared diffuse reflection spectroscopy. *Vib Spectrosc* 2011; 55: 58–64
- 52 Ioset KN, Nyberg NT, Van Diermen D, Malnoe P, Hostettmann K, Shikov AN, Jaroszewski JW. Metabolic profiling of *Rhodiola rosea* rhizomes by (1)H-NMR spectroscopy. *Phytochem Anal* 2011; 22: 158–165
- 53 Zhao C, Chan HY, Yuan D, Liang Y, Lau TY, Chau FT. Rapid simultaneous determination of major isoflavones of *Pueraria lobata* and discriminative analysis of its geographical origins by principal component analysis. *Phytochem Anal* 2011; 22: 503–508
- 54 Tianniam S, Bamba T, Fukusaki E. Pyrolysis GC-MS-based metabolite fingerprinting for quality evaluation of commercial *Angelica acutiloba* roots. *J Biosci Bioeng* 2010; 109: 89–93
- 55 Wang HL, Yao WF, Zhu DN, Hu YZ. Chemical fingerprinting by HPLC-DAD-ELSD and principal component analysis of *Polygala japonica* from different locations in China. *Chin J Nat Med* 2010; 8: 343–348
- 56 Suzuki R, Ikeda Y, Yamamoto A, Saima T, Fujita T, Fukuda T, Fukuda E, Baba M, Okada Y, Shirataki Y. Classification using NMR-based metabolomics of *Sophora flavescens* grown in Japan and China. *Nat Prod Commun* 2012; 7: 1453–1455

- 57 Kim N, Kim K, Choi BY, Lee D, Shin YS, Bang KH, Cha SW, Lee JW, Choi HK, Jang DS. Metabolomic approach for age discrimination of *Panax ginseng* using UPLC-Q-ToF MS. *J Agric Food Chem* 2011; 59: 10435–10441
- 58 Yang SO, Shin YS, Hyun SH, Cho S, Bang KH, Lee D, Choi SP, Choi HK. NMR-based metabolic profiling and differentiation of ginseng roots according to cultivation ages. *J Pharm Biomed Anal* 2012; 58: 19–26
- 59 Xue SY, Wang XJ, Sun HF, Zhang LZ, Qin XM, Li ZY. Metabolomic study of flower buds of *Tussilago farfara* in different development stages by GC-MS. *Zhongguo Zhong Yao Za Zhi* 2012; 37: 2863–2869
- 60 Wang JH, Wang ZH. Studies on enhancing therapeutic action of processing Chinese crude drugs. *Lishizhen J Tradit Chin Med Res* 1997; 8: 67–68
- 61 Sun H, Ni B, Zhang A, Wang M, Dong H, Wang X. Metabolomics study on Fuzi and its processed products using ultra-performance liquid-chromatography/electrospray-ionization synapt high-definition mass spectrometry coupled with pattern recognition analysis. *Analyst* 2012; 137: 170–185
- 62 Kim SH, Hyun SH, Yang SO, Choi HK, Lee BY. (1)H-NMR-based discrimination of thermal and vinegar treated ginseng roots. *J Food Sci* 2010; 75: C577–C581
- 63 Angelova N, Kong HW, van der Heijden R, Yang SY, Choi YH, Kim HK, Wang M, Hankemeier T, van der Greef J, Xu G, Verpoorte R. Recent methodology in the phytochemical analysis of ginseng. *Phytochem Anal* 2008; 19: 2–16
- 64 Zhang HM, Li SL, Zhang H, Wang Y, Zhao ZL, Chen SL, Xu HX. Holistic quality evaluation of commercial white and red ginseng using a UPLC-QTOF-MS/MS-based metabolomics approach. *J Pharm Biomed Anal* 2012; 62: 258–273
- 65 Lee JI, Ha YW, Choi TW, Kim HJ, Kim SM, Jang HJ, Choi JH, Choi MH, Chung BC, Sethi G, Kim SH, Ahn KS, Choi SH, Shim BS. Cellular uptake of ginsenosides in Korean white ginseng and red ginseng and their apoptotic activities in human breast cancer cells. *Planta Med* 2011; 77: 133–140
- 66 Kim SN, Ha YW, Shin H, Son SH, Wu SJ, Kim YS. Simultaneous quantification of 14 ginsenosides in *Panax ginseng* C.A. Meyer (Korean red ginseng) by HPLC-ELSD and its application to quality control. *J Pharm Biomed Anal* 2007; 45: 164–170
- 67 Leung KW, Wong AST. Pharmacology of ginsenosides: a literature review. *Chin Med* 2010; 5: 20
- 68 Yun TK, Lee YS, Lee YH, Kim SI, Yun HY. Anticarcinogenic effect of *Panax ginseng* CA Meyer and identification of active compounds. *J Korean Med Sci* 2001; 16: 6–18
- 69 Shibata S. Chemistry and cancer preventing activities of ginseng saponins and some related triterpenoid compounds. *J Korean Med Sci* 2001; 16: S28–S37
- 70 Kim WY, Kim JM, Han SB, Lee SK, Kim ND, Park MK, Kim CK, Park JH. Steaming of ginseng at high temperature enhances biological activity. *J Nat Prod* 2000; 63: 1702–1704
- 71 Kwon SW, Han SB, Park IH, Kim JM, Park MK, Park JH. Liquid chromatographic determination of less polar ginsenosides in processed ginseng. *J Chromatogr A* 2001; 921: 335–339
- 72 Park IH, Kim NY, Han SB, Kim JM, Kwon SW, Kim HJ, Park MK, Park JH. Three new dammarane glycosides from heat processed ginseng. *Arch Pharm Res* 2002; 25: 428–432
- 73 Lee SM, Shon HJ, Choi CS, Hung TM, Min BS, Bae KH. Ginsenosides from heat processed ginseng. *Chem Pharm Bull* 2009; 57: 92–94
- 74 Park IH, Han SB, Kim JM, Piao L, Kwon SW, Kim NY, Kang TL, Park MK, Park JH. Four new acetylated ginsenosides from processed ginseng (sun ginseng). *Arch Pharm Res* 2002; 25: 837–841
- 75 Chan EC, Yap SL, Lau AJ, Leow PC, Toh DF, Koh HL. Ultra-performance liquid chromatography/time-of-flight mass spectrometry based metabolomics of raw and steamed *Panax notoginseng*. *Rapid Commun Mass Spectrom* 2007; 21: 519–528
- 76 Toh DF, New LS, Koh HL, Chan EC. Ultra-high performance liquid chromatography/time-of-flight mass spectrometry (UHPLC/TOFMS) for time-dependent profiling of raw and steamed *Panax notoginseng*. *J Pharm Biomed Anal* 2010; 52: 43–50
- 77 Li SL, Song JZ, Qiao CF, Zhou Y, Qian K, Lee KH, Xu HX. A novel strategy to rapidly explore potential chemical markers for the discrimination between raw and processed *Radix Rehmanniae* by UHPLC-TOFMS with multivariate statistical analysis. *J Pharm Biomed Anal* 2010; 51: 812–823
- 78 Guo N, Fan B, Peng J, Yan H, Ma F, Yu Y. Ultra-performance LC/TOF MS analysis of fruits of *Ligustrum lucidum* for metabolomic research. *Chin J Exp Tradit Med F* 2010; 10: 131–133
- 79 Wang XJ. Based on NMR/UPLC plant metabolomics study on quality control of *Polygala tenuifolia* Willd [dissertation]. Shanxi: Shanxi University; 2012
- 80 Ruan GH, Li GK. The study on the chromatographic fingerprint of *Fucus xanthii* by microwave assisted extraction coupled with GC-MS. *J Chromatogr B Analyt Technol Biomed Life Sci* 2007; 850: 241–248
- 81 Ni Y, Peng Y, Kokot S. Fingerprinting of complex mixtures with the use of high performance liquid chromatography, inductively coupled plasma atomic emission spectroscopy and chemometrics. *Anal Chim Acta* 2008; 616: 19–27
- 82 Tanaka K, Tamura T, Fukuda S, Batkhuu J, Sanchir C, Komatsu K. Quality evaluation of *Astragalus Radix* using a multivariate statistical approach. *Phytochemistry* 2008; 69: 2081–2087
- 83 Okada T, Afendi FM, Altaf-Ul-Amin M, Takahashi H, Nakamura K, Kanaya S. Metabolomics of medicinal plants: the importance of multivariate analysis of analytical chemistry data. *Curr Comput Aided Drug Des* 2010; 6: 179–196
- 84 Tang JC, Zhang JN, Wu YT, Li ZX. Effect of the water extract and ethanol extract from traditional Chinese medicines *Angelica sinensis* (Oliv.) Diels, *Ligusticum chuanxiong* Hort. and *Rheum palmatum* L. on rat liver cytochrome P450 activity. *Phytother Res* 2006; 20: 1046–1051
- 85 Li HB, Jiang Y, Wong CC, Cheng KW, Chen F. Evaluation of two methods for the extraction of antioxidants from medicinal plants. *Anal Bioanal Chem* 2007; 388: 483–488
- 86 Martin J, Stöger EA, Wiebrecht A. Praxisleitfaden TCM-Drogen-Vorbehandlung-Zubereitung-Sondervorschriften. *Dtsch Zeitschrift Akupunktur (DZA)* 2009; 52: 72–74
- 87 Ding Y, Wu EQ, Liang C, Chen J, Tran MN, Hong CH, Jang Y, Park KL, Bae K, Kim YH, Kang JS. Discrimination of cinnamon bark and cinnamon twig samples sourced from various countries using HPLC-based fingerprint analysis. *Food Chem* 2011; 127: 755–760
- 88 Ni Y, Zhang L, Churchill J, Kokot S. Application of high performance liquid chromatography for the profiling of complex chemical mixtures with the aid of chemometrics. *Talanta* 2007; 72: 1533–1539
- 89 Li Y, Wu T, Zhu J, Wan L, Yu Q, Li X, Cheng Z, Guo C. Combinative method using HPLC fingerprint and quantitative analyses for quality consistency evaluation of an herbal medicinal preparation produced by different manufacturers. *Pharm Biomed Anal* 2010; 52: 597–602
- 90 Peng JB, Jia HM, Liu YT, Zhang HW, Dong S, Zou ZM. Qualitative and quantitative characterization of chemical constituents in Xin-Ke-Shu preparations by liquid chromatography coupled with a LTQ Orbitrap mass spectrometer. *J Pharm Biomed Anal* 2011; 55: 984–995
- 91 Yang CF. Application of UPLC/RRLC/UFLC in study on Chinese materia medica and its preparation. *Chin Tradit Herb Drugs* 2009; 39: 1259–1263
- 92 Zhang LJ. The development of NMR. *J Hebei Morm Univ* 2000; 24: 224–227
- 93 Daolio C, Beltrame FL, Ferreira AG, Cass QB, Cortez DA, Ferreira MM. Classification of commercial *Catuaba* samples by NMR, HPLC and chemometrics. *Phytochem Anal* 2008; 19: 218–228
- 94 Lin Y, Schiavo S, Orjala J, Vouros P, Kautz R. Microscale LC-MS-NMR platform applied to the identification of active cyanobacterial metabolites. *Anal Chem* 2008; 80: 8045–8054
- 95 Ali K, Iqbal M, Korthout HA, Maltese F, Fortes AM, Pais MS, Verpoorte R, Choi YH. NMR spectroscopy and chemometrics as a tool for anti-TNF α activity screening in crude extracts of grapes and other berries. *Metabolomics* 2012; 8: 1148–1161
- 96 Yang J, Xu G, Kong H, Zheng Y, Pang T, Yang Q. Artificial neural network classification based on high-performance liquid chromatography of urinary and serum nucleosides for the clinical diagnosis of cancer. *J Chromatogr B Analyt Technol Biomed Life Sci* 2002; 780: 27–33
- 97 Yang J, Xu G, Zheng Y, Kong H, Pang T, Lv S, Yang Q. Diagnosis of liver cancer using HPLC-based metabolomics avoiding false-positive result from hepatitis and hepatocirrhosis diseases. *J Chromatogr B Analyt Technol Biomed Life Sci* 2004; 813: 59–65
- 98 Yang J, Xu G, Zheng Y, Kong H, Wang C, Zhao X, Pang T. Strategy for metabolomics research based on high-performance liquid chromatography and liquid chromatography coupled with tandem mass spectrometry. *J Chromatogr A* 2005; 1084: 214–221
- 99 Pham-Tuan H, Kaskavelis L, Daykin CA, Janssen HG. Method development in high-performance liquid chromatography for high-throughput profiling and metabolomic studies of biofluid samples. *J Chromatogr B Analyt Technol Biomed Life Sci* 2003; 789: 283–301

- 100 Lenz EM, Wilson ID. Analytical strategies in metabolomics. *J Proteome Res* 2007; 6: 443–458
- 101 Chace DH, Kalas TA. A biochemical perspective on the use of tandem mass spectrometry for newborn screening and clinical testing. *Clin Biochem* 2005; 38: 296–309
- 102 Fan TW, Lorkiewicz PK, Sellers K, Moseley HN, Higashi RM, Lane AN. Stable isotope-resolved metabolomics and applications for drug development. *Pharmacol Ther* 2012; 133: 366–391
- 103 Gad HA, El-Ahmady SH, Abou-Shoer MI, Al-Azizi MM. Application of chemometrics in authentication of herbal medicines: a review. *Phytochem Anal* 2013; 24: 1–24
- 104 Dharmaraj S, Jamaludin AS, Razak HM, Valliappan R, Ahmad NA, Harn GL, Ismail Z. The classification of *Phyllanthus niruri* Linn. according to location by infrared spectroscopy. *Vib Spectrosc* 2006; 41: 68–72
- 105 Vlachos N, Skopelitis Y, Psaroudaki M, Konstantinidou V, Chatzilazarou A, Tegou E. Applications of Fourier transform-infrared spectroscopy to edible oils. *Anal Chim Acta* 2006; 573–574: 459–465
- 106 Kokalj M, Kolar J, Trafela T, Kreft S. Differences among *Epilobium* and *Hypericum* species revealed by four IR spectroscopy modes: transmission, KBr tablet, diffuse reflectance and ATR. *Phytochem Anal* 2011; 22: 541–546
- 107 Debonneville C, Thome MA, Chaintreau A. Hyphenation of quadrupole MS to GC and comprehensive two-dimensional GC for the analysis of suspected allergens: review and improvement. *J Chromatogr Sci* 2004; 42: 450–455
- 108 Ronda F, Rodríguez-Nogales JM, Sancho D. Multivariate optimisation of a capillary electrophoretic method for the separation of glutenins. Application to quantitative analysis of the endosperm storage proteins in wheat. *Food Chem* 2008; 108: 287–296
- 109 Ou LJ, Sun XP, Liu QD, Mi SQ, Wang NS. Effects of *Rhizoma Zingiberis* and *Pericarpium Citri reticulatae* extracts on myocardial ischemia in rats. *Zhong Yao Cai* 2009; 32: 1723–1726
- 110 Ooi LS, Li Y, Kam SL, Wang H, Wong EY, Ooi VE. Antimicrobial activities of cinnamon oil and cinnamaldehyde from the Chinese medicinal herb *Cinnamomum cassia* Blume. *Am J Chin Med* 2006; 34: 511–522
- 111 de Sousa DP, Nobrega FF, de Lima MR, de Almeida RN. Pharmacological activity of (R)-(+)-pulegone, a chemical constituent of essential oils. *Z Naturforsch C* 2011; 66: 353–359
- 112 Zhang GC, Fang SM. Antioxidation of *Pueraria lobata* isoflavones. *Zhong Yao Cai* 1997; 26: 340–343
- 113 Qi CF, Song SW, Liu DF, Du XF, Wan XC, Song LH. Effects of total isoflavones of *Pueraria* DC. on bone mineral density and Ca in ovariectomized rats. *J Biol* 2007; 24: 22–24
- 114 Gao D, Li Q, Li Y, Liu Z, Fan Y, Zhao H, Li J, Han Z. Antidiabetic and antioxidant effects of oleanolic acid from *Ligustrum lucidum* Ait in alloxan-induced diabetic rats. *Phytother Res* 2009; 23: 1257–1262