Application of Plant Metabonomics in Quality Assessment for Large-Scale Production of Traditional Chinese Medicine

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Abstract

The curative effects of traditional Chinese medicines are principally based on the synergic effect of their multi-targeting, multi-ingredient preparations, in contrast to modern pharmacology and drug development that often focus on a single chemical entity. Therefore, the method employing a few markers or pharmacologically active constituents to assess the quality and authenticity of the complex preparations has a number of severe challenges. Metabonomics can provide an effective platform for complex sample analysis. It is also reported to be applied to the quality analysis of the traditional Chinese medicine. Metabonomics enables comprehensive assessment of complex traditional Chinese medicines or herbal remedies and sample classification of diverse biological statuses, origins, or qualities in samples, by means of chemometrics. Identification, processing, and pharmaceutical preparation are the main procedures in the large-scale production of Chinese medicinal preparations. Through complete scans, plants metabonomics addresses some of the shortfalls of single analyses and presents a considerable potential to become a sharp tool for traditional Chinese medicine quality assessment.

Abbreviations

TCM: traditional Chinese medicine
CE: capillary electrophoresis
MVDA: multivariate data analysis
PCA: principal component analysis
HCA: hierarchical cluster analysis
PLS: partial least squares
OPLS: orthogonal partial least squares

O2PLS: bidirectional orthogonal partial least squares
SIMCA: soft independent modeling of class analogy
PLS-DA: partial least squares discriminant analysis
kNN: k nearest neighbors
ANN: artificial neural networks
MEND: matched filtration with experimental noise determination
AFLP: amplified fragment length polymorphism
TOF: time of flight
UPLC-QTOF-MS: ultra performance liquid chromatography quadrupole time of flight high definition mass spectrometry
BP-ANN: back propagation artificial neural network
ELSD: evaporative light scattering detector
HPTLC: high performance thin layer chromatography
DART: direct analysis in real time
PAD: photodiode array detector
LS-SVM: least squares support vector machine
RBF: radial basis function
NIRS: near-infrared spectroscopy
WG: white ginseng
RG: red ginseng
LTQ: linear trap quadrupole
LDA: linear discriminant analysis
SA: similarity analysis
COW: correlation optimized warping
UPGMA: unweighted pair group method with arithmetic mean

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**Introduction**

Metabonomics is an emerging subject of the post-genome era, which, together with genomics, transcriptomics, and proteomics, jointly constitutes the “Systems Biology” [1]. It is the branch of science concerned with the quantitative understanding of the metabolite components of integrated living systems and their dynamic responses to changes in both endogenous (such as those associated with physiology and development) and exogenous factors (such as environmental factors and xenobiotics) [2]. The success of the application of metabonomics has been illustrated in the literature from the perspective of the diagnosis of diseases such as diabetes [3], hypertension [4], and cancers [5, 6]. In recent years, a wide range of analytical metabonomic techniques have been implemented in research addressing TCM whose qualitative analysis is difficult because of the complexity and diversity of its components. In general, one or two biomarkers are used for identification and authentication of the herbal products. However, this approach does not provide information on the overall chemical composition of the plant extract, which is known to vary widely according to geographical origin, source, cultivar condition, harvesting and processing methods, and storage. Metabonomics, through achieving complete scans, addresses the shortfalls of single-component analysis. The rapid development of analytical instruments is accelerating research on TCM [7]. Additionally, multivariate statistical methods are increasingly improving, allowing the implementation of robust solutions [8]. Within the TCM practice, the majority of species used are plants. The multi-varieties employed in TCM are the main cause of confusion in the herbal medicine market. The identification of these varieties, as the first step in the production of Chinese medicinal preparations, is of great significance for ensuring the safety and effectiveness of clinical treatment. The quality and contents of the active components of herbs are highly variable depending on the species, parts of the herbs, cultivated geographic region, and planting period involved. Adulterants should be distinguishable from plant material and play the role of challenging substances. The processing of a characteristic portion, as the second step in production, appears to be of significance in clinical applications and has been proven to satisfy the requirements of therapeutics. It is essential to unify the degree of processing. In pharmaceutical production, extracts are commonly used. Metabonomics can be effectively applied for the quality control of plant extracts.

Identifying the plant material, processing, and pharmaceutical production is the sequence of manufacture for Chinese medicinal preparations. Here we demonstrate the application of metabonomics in the discrimination of TCM species, TCM production processes, and quality control. To avoid ambiguities, we also illustrated the factors affecting the identification step, the methods used in processing, and the forms of the pharmaceuticals.

**Analytical Techniques**

In recent years, many metabonomic-based methods have been implemented to facilitate research in the field of TCM. In the pharmacopeia, single-component analyses are employed in most research addressing TCM. Nevertheless, the lack of representativeness of single-component analyses seems to account for a deficiency of convincing data. Metabonomics, through achieving complete scans, addresses the shortfalls of single-component analysis. The rapid development of analytical instruments is accelerating research on TCM [7]. Metabonomics measures the multi-parametric response of biological systems to a stimulus, typically employing analytical technologies such as NMR or MS to obtain comprehensive profiling and comparison of metabolic “fingerprints” [9]. In addition, other chemical analytical equipments and techniques, such as UV and IR spectroscopy were also employed. For biomarker identification, it is also possible to separate out substances of interest on a larger scale from a complex biological system using techniques such as LC, multidimensional liquid separation systems, GC, and CE. Especially multidimensional liquid separation systems have the potential to become a powerful approach for enriching, separating, and quantifying a large variety of exogenous and endogenous compounds in complex biological samples and TCM preparations, with a powerful separation ability, high resolution and sensitivity, high-peak capacity, and excellent detection in comparison with one-dimensional HPLC. However, every analytical technique has its advantages and drawbacks, as shown in Table 1. Multi-analysis techniques can partially overcome the shortcomings of individual analytical techniques. It is believed that with the further development of metabonomics analysis techniques, especially those employing multi-analysis, metabonomics will strongly promote TCM research and be beneficial to its modernization in terms of extending the application of modern methods in the assessment of TCM.

<table>
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<tr>
<th>Abbreviation</th>
<th>Method Description</th>
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<tr>
<td>LSM</td>
<td>linear subspace model</td>
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<tr>
<td>PLS</td>
<td>partial least squares</td>
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<td>PCA</td>
<td>principal component analysis</td>
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<td>OPLS</td>
<td>orthogonal partial least squares</td>
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<td>SIMCA</td>
<td>soft independent model classification analysis</td>
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<td>LDA</td>
<td>linear discriminant analysis</td>
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<td>SVM</td>
<td>support vector machine</td>
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<td>RF</td>
<td>random forest</td>
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<td>DT</td>
<td>decision tree</td>
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<td>ANN</td>
<td>artificial neural network</td>
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<td>KNN</td>
<td>k-nearest neighbors</td>
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<td>CBS</td>
<td>chemical brain storm</td>
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<td>GA</td>
<td>genetic algorithm</td>
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<td>DE</td>
<td>differential evolution</td>
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Table 1. Multi-analysis techniques

More specifically, metabonomics can be applied in various aspects of TCM, such as diagnosis, treatment, and drug development. Particularly, metabonomics can provide useful information to understand the pharmacological effects of TCM, leading to a better understanding of the disease and its treatment. This approach can be utilized to identify the active components of TCM and provide insights into their biological activities. Furthermore, metabonomics can be used to study the efficacy of TCM in clinical trials, allowing for the optimization of treatment strategies. By integrating metabonomics with other omics approaches, researchers can gain a comprehensive understanding of TCM, ultimately leading to the development of more effective and personalized therapies.
It is also possible to use MVDA to conduct regression analysis methods based on the classification of samples into different groups (e.g., by treatment or genotype), both via supervised (e.g., discriminant function analysis or artificial neural networks) and unsupervised data analysis methods (e.g., PCA or HCA) [11]. It is also possible to use MVDA to conduct regression modeling between two blocks of data, usually denoted as X and Y. In metabolomics-based NP studies, X may represent signals from different metabolites present in plant extracts sampled at regular time intervals, while Y represents responses (e.g., the quality of product, bioactivity, or yield). The model then can be used to predict Y from X, which is achieved through new observations. The most common MVDA method employed for this type of modeling is the PLS method [12]. Recently, OSC, OPLS, and O2PLS were utilized [13, 14]. The specific methods include SIMCA, PLS-DA, kNN, and ANN. Moreover, PCA and HCA are widely used in metabolomics research.

Model validation consists of cross validation, permutation testing, and external validation [10]. There are two significant procedures: noise filtration and peak matching [15]. Nonlinear noise filtration is extensively employed, substituting a point with the average of the surrounding points so as to filter noise successfully [16]. Matched filtration is another method setting up a standard mode of a peak and comparing its width; a narrower peak is regarded as noise [17]. Andreev et al. [18] developed MEND, improving the identification function. As for peak matching, identifying the retention time of the internal standard substance under the same conditions is the main method employed. The main diagnostic tools applied include score plots, loading plots, VIP, DModX, and regression coefficients [10]. MetExtract, a new software tool for the automated comprehensive extraction of metabolite-derived LC/MS signals in metabolomics research, was recently reported in the literature [19].

### Bioactivity and Profiles

The main research methods of metabolomics are metabolomic fingerprinting and metabolomic profiling analysis. Metabolomic profiling can be divided into two parts, targeted and nontargeted metabolic profiling analysis. The targeted metabolic profiling aims to search one biomarker. Several components were usually chosen as marker compounds to assess the quality. These biomarkers were proved to be constituents that discriminate the different species, different parts, different cultivated geographic regions, different planting periods, and the processing products. In the present paper, the bioactivities of the biomarkers were obtained via PubChem (http://www.ncbi.nlm.nih.gov/pccompound) and related literatures. However, these few selected markers sometimes are not unique to a particular herb since they might be present in many plants belonging to various families. In addition, the selection of suitable markers is sometimes difficult and subjective. Furthermore, adulterators are continuously trying to develop ways to make their products’ chemical profile similar to the authentic medicinal herbal product. Under these specific circumstances, the marker approach, on the one hand, is unable to confirm the identity of a specific plant. On the other hand, the influences of the other inner chemicals present may be ignored. Therefore, in some cases, its use may be inappropriate for quality control purposes [20, 21]. The objective of “nontargeted” analysis is to describe metabolic events by determining all detectable metabolites [22]. Of the various profiling techniques, nontargeted analysis using UPLC–MS is a promising tool for investigating the diversity of phytochemicals [23]. Thus, it is believed that nontargeted metabolic profiling analysis will play an important role as an effective tool in terms of high-throughput elucidation of metabolic phenotypes.

### Data Processing Methods

The progress of metabolomics research will be illustrated. First, the proposed TCM component should be extracted. Second, analytical tools should be applied. Third, the chemical profile should be obtained. Combining data under a multivariate data analysis, validating models, molding, and applying diagnostic tools are the consecutive steps [10] supported by chemometrics and mathematical statistics.

Chemometrics are basically classified into two main categories: pattern recognition methods (unsupervised and supervised), when a qualitative evaluation is involved, and multivariate calibration for quantitative purposes. Data resulting from metabolomics-based work are typically high-dimensional data, requiring MVDA methods for interpretation. Most metabolomics data analysis methods are based on the classification of samples into different groups (e.g., by treatment or genotype), both via supervised (e.g., discriminant function analysis or artificial neural networks) and unsupervised data analysis methods (e.g., PCA or HCA) [11].

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<td></td>
<td>Low cost</td>
<td>Noninvasive and nondestructive for samples</td>
<td>High sensitive detection for almost both volatile and nonvolatile compounds</td>
<td>High speed and short analysis time</td>
<td>Realize identification and quantification of volatile and thermally stable components</td>
<td>Easy to be applied</td>
<td>Potential to use vibrational spectroscopy</td>
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<td></td>
<td>Easy to use</td>
<td>Quantitative and simultaneous detection unbiased for any molecules</td>
<td>Has more peak capacity and can accommodate more complex mixtures</td>
<td>Less sample and solvent consumption</td>
<td>Used for ionization of polar to nonpolar components</td>
<td>Limited use for compounds without UV absorption</td>
<td>Lacks specificity</td>
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<td></td>
<td>Highly sensitive</td>
<td>High throughput</td>
<td>Unsuitable for nonvolatile and thermally unstable compounds</td>
<td>Appropriate for complex samples</td>
<td>Associates with LC overcoming problems</td>
<td>Lower operating cost</td>
<td>Lacks specificity</td>
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<td>Not limited by sample volatility and stability</td>
<td>Produces rich, dynamic molecular information</td>
<td>Being destructive</td>
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<td>Requires little or no sample preparation</td>
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<td>Favorable separating power</td>
<td>Requires little or no sample preparation</td>
<td>Requiring preknowledge about samples</td>
<td>Lower sensitivity than HPLC</td>
<td>Requires little or no sample preparation</td>
<td>Good resolution and reproducibility</td>
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The targeted metabolic profiling aims to search one biomarker. Several components were usually chosen as marker compounds to assess the quality. These biomarkers were proved to be constituents that discriminate the different species, different parts, different cultivated geographic regions, different planting periods, and the processing products. In the present paper, the bioactivities of the biomarkers were obtained via PubChem (http://www.ncbi.nlm.nih.gov/pccompound) and related literatures. However, these few selected markers sometimes are not unique to a particular herb since they might be present in many plants belonging to various families. In addition, the selection of suitable markers is sometimes difficult and subjective. Furthermore, adulterators are continuously trying to develop ways to make their products’ chemical profile similar to the authentic medicinal herbal product. Under these specific circumstances, the marker approach, on the one hand, is unable to confirm the identity of a specific plant. On the other hand, the influences of the other inner chemicals present may be ignored. Therefore, in some cases, its use may be inappropriate for quality control purposes [20, 21]. The objective of “nontargeted” analysis is to describe metabolic events by determining all detectable metabolites [22]. Of the various profiling techniques, nontargeted analysis using UPLC–MS is a promising tool for investigating the diversity of phytochemicals [23]. Thus, it is believed that nontargeted metabolic profiling analysis will play an important role as an effective tool in terms of high-throughput elucidation of metabolic phenotypes.
Identification of Traditional Chinese Medicine Components

The identification of traditional Chinese medicine components plays a key role in ensuring the safety and effectiveness of clinical treatments. The quality and contents of the active constituents in herbs are highly variable depending on the species, parts of the plant, cultivated geographic region, and planting period involved. Adulterants are assumed to be distinguishable from plant material and play the role of challenging substances. Therefore, during large-scale production, the identification of these components is of vital significance. Moreover, many applications employed in the development of metabolic fingerprinting, which will be explained below, using appropriate analysis methods coupled with multivariate analysis, have been investigated and applied to discriminate between closely related plant species in performing quality control assessments of herbal drugs and to identify their different geographic origins. In addition, analyzing components is a robust way to control plant quality.

Here, the identification of TCM components can be divided into four categories: the various species and adulterants, the different parts of herbs, the planting period and the cultivated geographic region.

Identification of species and adulterants

Different species may contain approximately the same components, while the contents of these components vary, which influence the therapeutic effect. Two leguminous plants, Astragalus membranaceus (Fisch.) Bge. var. mongholicus (bge.) Hsiao and Astragalus membranaceus (Fisch.) Bge, are important medical herbs that share great similarities regarding their morphology, chemical constituents, and genomic DNA sequences. The identification of different medicinal species directly affects their pharmacological and clinical effects. Amplified AFLP-based genetic fingerprinting and GC–TOF/MS-based metabolic fingerprinting were used to successfully discriminate between the two species. The differences in some soluble sugars, fatty acids, proline, and polyamine reflected the plants’ adaptation to different growth environments. Using multivariate and univariate statistical analyses, three AFLP markers and eight metabolites were identified as candidate DNA and metabolic markers to distinguish between the two herb materials [24]. In another study, metabolite profiling of five medicinal Panax herbs, which included P. ginseng (Chinese ginseng), P. notoginseng, P. japonicus, P. quinquefolium L., and P. ginseng (Korean ginseng), was performed using UPLC-QTOFMS and multivariate statistical analysis techniques. PCA of the analytical data showed that the five Panax herbs could be separated into five different groups of phytochemicals [25]. HPLC fingerprinting was used for comparison of three closely related species of Pericarpium Citri (Citrus reticulata ‘Chachí’, Citrus reticulata ‘Dahongpao’, and Citrus erthrosa Tanaka), and PLS-DA identified hesperidin, tangeretin, and nobiletin as potential biomarkers for their classification [26]. In a similar case, PCA and HCA as well as SIMCA and a BP-ANN were applied to identify and distinguish Epimedium wushanense and Epimedium koreanum based on their secondary metabolites. The SIMCA method failed to identify one sample, whereas BP-ANN precisely predicted the whole test set [27]. PCA was able to discriminate between ten Aristolochia species on the basis of their essential oil profiles, showing that 2 h of hydrodistillation produce the best outcome when the oils are used for discriminating between species [28]. Coincidentally, Sun et al. [29] drew on the same plant. In the Chinese Pharmacopoeia 2010, only two Aconitum species are recorded. One is the root of Aconitum kusnezoffii Reichb., namely “Caowu” in Chinese. The other species was Aconitum carmichaelii Debx. Two herbal drugs are derived from this species. The two species were distinguished successfully using UPLC–QTOF–HDOMS, combining with PCA and S-plot. Moreover, a PCA score plot clearly demonstrated discrimination between Artemisia annua and Artemisia afra on the basis of phenylpropanoids (caffeic acid, chlorogenic acid, di-cafeoyl quinic acid, and ferulic acid) [30]. Spectral fingerprinting via NIR has been utilized for the rapid identification and counterfeit detection of Eleutherococcus senticosus, and PCA, DA, SIMCA, and PLS-DA were found to allow good discrimination between E. senticosus and other herbs both related to and not related to the Araliaceae family [31]. PCA has been successfully applied for distinguishing Angelica sinesis from related Apiaceae (syn. Umbelliferae) herbs based on complete HPLC fingerprints [32]. The same biomarkers were recognized by PLS-DA for the discrimination of authentic Pericarpium Citri from commercial samples, mixed peel samples, and other citrus peels [33]. 1H-NMR spectroscopy and multivariate data analyses were applied to discriminate two Bupleurum species (B. chinense and B. scorzoneraefolium) and to explore the influences of habitat and culture methods on the quality of Radix Bupleuri plants based on their metabolic profiles [34]. The quality of Radix Bupleuri plants was evaluated via HPLC-ELSD analysis and HPTLC based on analysis of their principal bioactive components (saikosaponins). The acquired data were processed using ANNs and KNN to distinguish between different species of the genus [35]. DART-MS provides a novel mass spectrometric ion source by producing [M + H]+ molecular ion species. In analyses of Glycyrrhiza inflata Batalin, the peak at m/z 339 originates mainly from the [M + H]+ of LA, a species-specific compound. These results indicate that G. inflata can be differentiated from the other two species based on detection of LA peaks using DART-MS analysis [36]. In addition, chromatographic fingerprinting via GC-MS coupled with SA and PCA has been undertaken for discriminating Scutellaria barbata D. Don from adulterants. The results showed that the samples could be identified based on differences between the samples and various adulterants [37]. Similarity analysis and HCA were applied for the first time to identify and distinguish genuine Aconitum kusnezoffii from its adulterants, which demonstrated the feasibility of linking the HCA approach to chemotaxonomic analysis on the basis of the presence of alkaloids [38]. To discriminate and assess the quality of Curcuma phaeocaulis, C. kwangsiensis, and C. wenyujin from different ecotypes, a metabolomics analysis was carried out via GC-MS coupled with multivariate statistical analysis. Characterization of phytochemicals in essential oils was performed by automated matching to the MS library and comparison of their mass spectra, which discriminated among the different plant parts [39]. Curcuma plants, such as Curcuma wenyujin Y.H. Chen et C. Ling and Curcuma longa L., were also distinguished successfully via HPLC-DAD-MS [40].

Identification of different medicinal parts of herbs

The choice of different parts of herbs determines the resulting curative effect, which is the purpose of therapy. The contents of the active components of diverse plant parts were identified. In the long history of the use of medicinal plant preparations, different plant parts have been regarded as different drugs. R. Jurišić Grušetić et al. [41] identified the variation in total polyphenol contents, employing Folin–Ciocalteu’s reagent, between different parts of Plantago plants (leaves: up to 10.15%; stems: up to...
Metabolite profiling of different parts of *Panax notoginseng* was carried out using UPLC–ESI–MS and multivariate statistical analysis. PCA of the UPLC–ESI–MS data showed a clear separation of the compositions among the flower buds, roots, and rhizomes of *P. notoginseng*. The saponins accounting for these variations were identified based on corresponding loading weights and were further verified based on the accurate mass, tandem mass, and retention time of available standard saponins using UPLC–QTOF-MS [42]. Moreover, each extract from 24 mulberry leaf samples, divided into six locations from the tip of the stem in each of four strains, was analyzed via pattern recognition methods, including PCA and SIMCA. The 24 extracts from mulberry leaves showed independent spectra in ¹H-NMR analyses [43]. *Aconitum carmichaelii* Debx., another example for the application of plant metabolomics in the discrimination of different parts of herb plants, was studied by Sun et al. [29]. The mother root is named “Chuanwu,” while the daughter or lateral root of *Aconitum carmichaelii* Debx. is known as “Shengfuzi.” Shengfuzi has been prescribed more frequently than Chuanwu to treat rheumatic diseases. The analytical techniques, UPLC–Q–TOF–HDMS, as well as the data processing methods, PCA, and S-plot, were the main measures in this study.

Differentiation of distinct cultivated geographic regions

The environments of the cultivated geographic regions of medicinal plants, including their temperature, humidity, soil, and climate, are determinant factors. Therefore, the cultivated geographic region influences the growth of herbs. Wei-Jun Kong et al. [44] utilized UPLC-PAD analysis to examine the five active alkaloids in *Rhzoma Coptidis Chinensis*, successfully grouping the plants in accordance with their province of origin. Moreover, the LS-SVM, RBF-ANN, PLS-DA, and kNN methods were applied for the classification of *Rhzoma Corydalis* from two different geographical origins [45]. Another example of the application of the HCA technique was its use for the classification of *Isatis indigotica* roots collected from different regions based on HPLC fingerprinting [46]. *Canaderrma lucidum* samples from different cultivated geographic regions were evaluated using HPLC fingerprinting. The HCA, PCA, PLS-DA, and SIMCA techniques were employed to classify samples in accordance with their province of origin [47]. In addition, the essential oils of the *Cinnamomum* Cortex specimens obtained from different localities have been analyzed via GC-MS [48]. Furthermore, the volatile oils of *Artemisia capillaris* Herba from different locations were investigated through GC-MS to develop a characteristic fingerprint of this raw herb [49]. The discrimination of *Schizonepeta tenuifolia* Briq. from different origins has also been achieved via PCA and HCA, which classified the samples into two main groups on the basis of five marker compounds [50]. It is also worth noting that the combination of NIR spectroscopy with DA and PLS-DA analysis was applied in geographical origin discrimination for *Radix Scutellaria Baicalensis* [51]. A nontargeted procedure was applied for ¹H-NMR spectroscopic fingerprinting of extracts from *Rhodiola rosea* rhizomes with pattern recognition analysis and identification of secondary metabolites responsible for differences in sample composition. For this purpose, plants from three different geographic areas (the Swiss Alps, Finland, and the Altai region in Siberia) were investigated [52]. Furthermore, quantitative estimates of the major isoflavones in *Pueraria lobata* were produced, and the studied samples were classified through PCA based on the amounts of puerarin, daidzin, daidzein, and genistin present [53]. Metabolite fingerprinting was applied in an attempt to evaluate the quality of dried *Angelica acutiloba* roots. An enhanced understanding of the dominance of the relationship of the cultivation area with the evaluated quality was conceptualized and applied to the construction of a PLS-DA classification model, which provided the basis for accurate and reliable predictivity [54]. Additionally, PCA was performed using the data generated through HPLC-DAD-ELSD analysis for quality control of *Polygala japonica* from different localities in China [55]. Recently, Suzuki et al. [56] classified *Sophora flavescens* grown in Japan and China via NMR.

Differentiation of distinct planting periods

The planting period is also a vital factor in the quality of a crude drug due to the duration over which a plant obtains nutrition from the soil. An UPLC-Q-TOF-MS-based metabolomic technique was applied for metabolite profiling in 60 *Panax ginseng* samples aged from 1 to 6 years [57]. Ginseng is an important herbal resource worldwide, and adulteration or falsification of the cultivation age has been a serious problem for ginseng in the commercial market. In this study, ginseng roots cultivated for 2–6 years under good agricultural practices standard guidelines were analyzed via NMR-based metabolomics techniques using two solvents [58]. Moreover, it has been demonstrated that July might be the best harvest time for *Pericarpium Citri Reticulatae Viride*, while November and December are better for *Pericarpium Citri Reticulatae*. Furthermore, hesperidin, nobiletin, and tangeretin were screened as chemical markers based on PCA loadings. The HPLC–HELP–PCA strategy has shown potential in the optimization of harvest times [26]. Recently, Xue et al. [59] utilized GC-MS to investigate the flower buds of *Tussilago farfara* in different development stages. Collectively, medicinal herbs from different species and the different parts of the same plant usually exhibit different efficacy, pharmacological actions, and clinical indications due to the significant differences in the types and quantity of the constituents. The species diversity seems to be a significant factor to influence the quality assessment. In addition, chemical constituents of the same plant may be various due to different cultivation areas, climatic conditions, and cultivation ages. For example, ginseng of cultivation ages from 4 to 6 years is the most demanded ginseng in the market. However, age and cultivation areas can hardly be determined by the herb’s physical appearance alone. Accordingly, confused clinical application led to the consumption of incorrect forms of plant material, improper use, and undesirable effects. Hence, an effective method applied in quality control is urgently demanded for the identification step of medicinal herbs. Since some samples share similarities in morphology but with subtle variations in certain ingredients, metabolomics can provide a platform to use analytical techniques coupled with multivariate statistics for the differentiation of these complex samples. Metabolomics information not only assist in the establishment of a deeper understanding of the complex interactive nature of plant metabolic networks and their responses to environmental change but also provide unique insights into the fundamental nature of plant phenotypes in relation to development, physiology, tissue identity, resistance, biodiversity, and so on. To make them...
clear, the plant material, analysis technique, chemometrics methods, biomarkers, and bioactivity aspects are summarized in Table 2.

Processing

Processing is the second step in the production of Chinese medicinal preparations. TCM-specific production steps include storing, washing, rinsing, drying, remoistening, and cutting, and eventually, unique processing techniques, such as stir frying, steaming, or calcining are performed to satisfy different clinical therapeutic requirements. First, the effect of processing is thought to enhance the therapeutic efficiency in Sophora japonica L. [60]. Drug processing can also weaken the structure of plants so that the active components can be extracted easily. Additionally, additives react with the compounds present in plants generating new components dissolved in solvents. For instance, alkaloids dissolve in acidic solvents, so vinegar is widely applied to crude drugs to enrich alkaline substances. Second, processing has been reported to reduce the toxicity of the crude drugs, as described with Fuzi [61]. As evidence has accumulated, it has been shown that poisonous protein is one of the causes of accidental side effects. As proteins are thermo-sensitive, crude drugs should be subjected to heat treatment. Third, the expansion of applications is another important unexpected impact. Finally, after processing, the generation of several new compounds has been reported [62]. However, the dosage of additives and the time of heat treatment should be considered during processing, as they are the major factors that affect quality control.

In the large-scale production, it is difficult to guarantee the purity of all products. Researchers spend long periods finding solutions to quality control. The development of metabolomics has provided a necessary way to understand cellular responses to mutations at all levels of gene products [63]. In recent years, a wide range of metabolonomic analytical techniques have been implemented in research on TCM [64]. Several cases illustrate the possibilities of the application of metabolomics in quality control during the processing of TCM materials. Ginseng has been employed in TCM for over two thousand years and is now widely used around the world as an elixir [63]. In Asia, there are two types of ginseng that are commonly found in the herbal medicine market: WG and RG. In the practice of traditional Chinese medicine, WG and RG have been used for different purposes. WG is traditionally produced via sun drying of fresh ginseng, and RG is manufactured by steaming fresh ginseng at 95–100°C for 2–3 h and then drying it. WG is used to “supply qi and promote the production of body fluids” as well as enhance physical fitness and disease resistance, while RG has a “warming effect” and is used for “boosting yang” and replenishing vital essence [64]. Ginsenosides Rb1, Rb2, Rc, Rd, Rg1, and Re are the major constituents of both WG and RG, while ginsenosides Rg3, Rg5, Rg6, Rh1, Rh2, Rk1, Rs3, and F4 are known to be unique constituents of RG [65–72]. These unique ginsenosides found in RG have been reported to be converted from the ginsenosides found in fresh ginseng after steaming [70,73,74]. In one study, ginseng was processed under temperatures of 100, 140, and 180°C, with or without vinegar; the duration of exposure to each temperature was 10, 30, and 50 min, respectively, and there was a clear separation in the score plots obtained for the various treatment conditions. The major compounds contributing to the separation of 50% methanol extracts of vinegar-treated ginseng subjected to various processing conditions were valine, lactate, alanine, arginine, glucose, fructose, and sucrose. As the temperature increased, the valine, arginine, glucose, fructose, and sucrose concentrations decreased, whereas lactate, glucose, and fructose increased in the vinegar-treated samples compared to non-vinegar-treated samples [62]. Moreover, UPLC/TOFMS had been demonstrated to be a powerful tool for use in herbal metabolomics to discriminate differentially processed herbs, such as raw and steamed P. notoginseng [75]. An UHPLC-TOF-MS-based metabolomics platform coupled with PCA and PLS-DA was developed for Panax notoginseng to establish a correlation between the duration of steaming and the maximum production of bioactive ginsenosides [76]. A similar study was performed to determine chemical markers for discriminating between raw and processed Radix Rehmanniae samples [77]. In addition, the three types of products obtained from the processing of Ligusticum lucidum fruits have been distinguished, which correspond to steam treatment processing products, vinegar treatment processing products, and the fruits processed with wine. There are differences in metabolite profiles among the crude and different types of processed fruits of L. lucidum. Ligustilode B was identified as a chemical marker for such variations, and its contents in crude L. lucidum specimens were found to be significantly higher than in processed samples. This study indicated that UPLC-QTOF-MS coupled with multivariate statistics is able to provide quality control for the crude and processed fruits of L. lucidum, and these results provide the basis for determining the appropriate mechanism of processing [78]. The products of the processing of Polygala Radix were also successfully distinguished [79]. One study was designed to perform a comprehensive metabolomics analysis of Fuzi and its processed products, Yanfuzi, Heishunpian, and Baifu-pian, via UPLC-Q-TOF-HDMS combined with pattern recognition methods. Differences in the metabolic profiles of Fuzi and its processed preparations were clearly observed based on PCA of the obtained MS spectra. Significant changes in 19 metabolite biomarkers were detected in the Fuzi samples and the three preparations [61]. Similarity analysis and PCA were applied to address the issue of the various quality changes that occur during the process of toasting Fructus Xanthii supplied by different producing areas. A high similarity was observed between different samples, which indicates that the proportion and distribution of the components in most extracts of F. xanthii show a high level of consistency [80]. HPLC fingerprints together with metal profiles were employed to assess the quality control procedures applied to Atractylis chinensis. A separate data matrix and combined data matrices were analyzed via PCA, kNN, and LDA. The PCA results from the combined data matrices indicated that the samples were discriminated on the basis of the applied processing methods. Within each group, the samples were reasonably well grouped according to their geographical origin and classification using kNN, and LDA results supported the PCA results [81]. As the raw and processed forms of herbs have different pharmacological actions, it is pertinent to administer the correct form of herb to avoid any undesirable consequences. Even the duration of the processing procedure, the processing adjuvants, and its dosage arouse the subtle changes in the contents of compounds. Therefore, it is of paramount importance to characterize the specific forms. The collection of quoted literature data is shown in Table 2.
<table>
<thead>
<tr>
<th>TCM materials</th>
<th>Pur-</th>
<th>Analysis techniques</th>
<th>Chemometrics methods</th>
<th>Biomarkers</th>
<th>Bioactivity</th>
<th>Ref.</th>
</tr>
</thead>
<tbody>
<tr>
<td>Two Astragalus plants</td>
<td>A1</td>
<td>GC-TOF/MS</td>
<td>AFLP</td>
<td>Three AFLP markers, eight metabolites</td>
<td>Antiperspirant and antidiuretic</td>
<td>[24]</td>
</tr>
<tr>
<td>Five Panax plants</td>
<td>A1</td>
<td>UPLC-QTOF-MS</td>
<td>PCA</td>
<td>Ginsenoside Rf, Rb, Rb2, 20(s)-pseudo-sequinsenosid e, F11</td>
<td>Antineoplastic, hypolipidemic agents*</td>
<td>[25]</td>
</tr>
<tr>
<td>Panax notoginseng</td>
<td>A2</td>
<td>UPLC-QTOF-MS</td>
<td>PCA</td>
<td>Saponins</td>
<td>Prevention and treatment of cerebrovascular diseases, immune regulation, hep to protection, anticarcinogen, neuroprotective effect</td>
<td>[42]</td>
</tr>
<tr>
<td>Panax ginseng</td>
<td>A4</td>
<td>UPLC-QTOF-MS</td>
<td>PCAHCA RF PAM PLS-DA</td>
<td>\</td>
<td>\</td>
<td>[57]</td>
</tr>
<tr>
<td>Panax ginseng</td>
<td>B</td>
<td>UPLC-QTOF-MS</td>
<td>PCA</td>
<td>\</td>
<td>\</td>
<td>[62]</td>
</tr>
<tr>
<td>Panax ginseng</td>
<td>B</td>
<td>UPLC-QTOF-MS</td>
<td>PCA, PLS-DA</td>
<td>\</td>
<td>\</td>
<td>[76]</td>
</tr>
<tr>
<td>Panax ginseng</td>
<td>A4</td>
<td>NMR</td>
<td>PCA, PLS-DA</td>
<td>Amino acids, organic acids, sugars</td>
<td>Cardiovascular control of blood pressure</td>
<td>[58]</td>
</tr>
<tr>
<td>Three tangerine peels</td>
<td>A1, 4</td>
<td>HPLC-DAD</td>
<td>PCA, HELP</td>
<td>Hesperidin, tangeretin, nobilitein</td>
<td>Protective effect on myocardial ischemia</td>
<td>[26]</td>
</tr>
<tr>
<td>Mallotus plants</td>
<td>A1, 2, 3</td>
<td>LC-MS</td>
<td>PLS-DA</td>
<td>Senkyunolide A</td>
<td>Antioxidants</td>
<td>[33, 109]</td>
</tr>
<tr>
<td>Two Epimedium plants</td>
<td>A1</td>
<td>HPLC</td>
<td>PCA, HCA, SIMCA, BP-ANN</td>
<td>Flavonoids</td>
<td>Influence on sexual function, antiaging, effect on immune system, anti-inflammatory, antiinvasive, expectorant, antiasthma</td>
<td>[27]</td>
</tr>
<tr>
<td>Rhizoma Coptidis</td>
<td>A3</td>
<td>UPLC</td>
<td>SA, HCA, PCA</td>
<td>Berberine, coptisine, palmitine, jateorrhizine, epiberberine</td>
<td>Efficacy of suppressing fever, dispelling dampness, removing toxicosis and anti-microbes*</td>
<td>[44]</td>
</tr>
<tr>
<td>Five Bupleurum plants</td>
<td>A1</td>
<td>¹H-NMR HPLC-ELSD HPTLC</td>
<td>ANNs, kNN</td>
<td>Saikosaponins</td>
<td>Anti-inflammatory, antineoplastic, immunosuppressive agents*</td>
<td>[34] [35]</td>
</tr>
<tr>
<td>Ten Aristolochia plants</td>
<td>A1</td>
<td>GC-MS</td>
<td>PCA</td>
<td>Essential oils</td>
<td>Abortifacients, stomachics, antiophidians, antiasthmatics, expectorants, slimming therapies</td>
<td>[28]</td>
</tr>
<tr>
<td>Two Artemisia plants</td>
<td>A1</td>
<td>NMR</td>
<td>PCA</td>
<td>Polar components</td>
<td>Antiplasmodial</td>
<td>[30]</td>
</tr>
<tr>
<td>Artemisia capillaris herba</td>
<td>A3</td>
<td>GC-MS</td>
<td>EWOP, FSM/WEFA</td>
<td>Essential oils</td>
<td>Choleretic, anti-inflammatory and diuretic agent in the treatment of epidemic hepatitis</td>
<td>[49]</td>
</tr>
<tr>
<td>Three Curcuma plants</td>
<td>A1</td>
<td>GC-MS</td>
<td>PCA, PLS-DA</td>
<td>Essential oils</td>
<td>Against skin diseases, colic inflammatory disorders, insect repellants, antimicrobial</td>
<td>[39]</td>
</tr>
<tr>
<td>Two Curcuma plants</td>
<td>A1</td>
<td>HPLC-DAD-MS, GC-MS</td>
<td>PCA</td>
<td>Curcumin, demethoxycurcumin, bisdemethoxycurcumin, dihydrocurcumin, ar-turmerone, α,β-turmerone, zingiberene</td>
<td>Against skin diseases, colic inflammatory disorders, insect repellants, antimicrobial, antidiabetic medications</td>
<td>[40]</td>
</tr>
<tr>
<td>Rhizoma Corydalis</td>
<td>A3</td>
<td>NIRS</td>
<td>WT, LS-SVM, PLS-DA, KNN</td>
<td></td>
<td>\</td>
<td>[45]</td>
</tr>
<tr>
<td>Eleutherooccus senticosus</td>
<td>A1</td>
<td>NIRS</td>
<td>PCA, DA, SIMCA and PLS-DA</td>
<td>\</td>
<td>\</td>
<td>[31]</td>
</tr>
<tr>
<td>Angelica sinensis</td>
<td>A1</td>
<td>HPLC</td>
<td>PCA</td>
<td>Senkyunolide A</td>
<td>Treatment of gynecological diseases</td>
<td>[32]</td>
</tr>
<tr>
<td>Plantago L.</td>
<td>A2</td>
<td>UV</td>
<td>UPGMA, PCA</td>
<td>Polyphenols, tannins</td>
<td>Diuretic</td>
<td>[41]</td>
</tr>
<tr>
<td>Three Glycyrrhiza plants</td>
<td>A1</td>
<td>DART-MS</td>
<td>\</td>
<td>Licochalcone A</td>
<td>Antimicrobial activity, antiplasmodial activity, antiestimonal activity*</td>
<td>[36]</td>
</tr>
<tr>
<td>Mulberry leaf</td>
<td>A3</td>
<td>¹H-NMR</td>
<td>PCA, SIMCA</td>
<td>\</td>
<td>Anti-inflammatory, inhibition of leucine-rich repeat kinase-2, proliferative and androgenic effects*</td>
<td>[46]</td>
</tr>
<tr>
<td>Isatis indigotica</td>
<td>A3</td>
<td>RP-HPLC</td>
<td>HCA</td>
<td>Indirubin, indigotin</td>
<td>Anti-inflammatory, inhibition of leucine-rich repeat kinase-2, proliferative and androgenic effects*</td>
<td>[46]</td>
</tr>
<tr>
<td>Cortex Cinnamomi</td>
<td>A3</td>
<td>GC-MS</td>
<td>IOP, HELP, SFA, OPR</td>
<td>Essential oils</td>
<td>Antimicrobial activities</td>
<td>[48, 110]</td>
</tr>
<tr>
<td>Ganoderma lucidum</td>
<td>A3</td>
<td>NIRS</td>
<td>HCA, PCA, PLS-DA, SIMCA</td>
<td>Triterpenoidsaponins, polysaccharides</td>
<td>Inhibitors of the in vitro human recombinant aldose reductase*</td>
<td>[47]</td>
</tr>
</tbody>
</table>

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Table 2 Continued

<table>
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<th>Analysis techniques</th>
<th>Chemometrics methods</th>
<th>Biomarkers</th>
<th>Bioactivity</th>
<th>Ref.</th>
</tr>
</thead>
<tbody>
<tr>
<td>Schizonepeta tenuifolia Briq.</td>
<td>A3</td>
<td>GC-MS</td>
<td>PCA, HCA</td>
<td>2-Hydroxy-2-isopropenyl-5-methylc, cis-pulegolone oxide, menthone, pulegolone, cyclo-hexanone, schizonal</td>
<td>Antifungal properties, decrease in ambulation, and increase in pentobarbital-induced sleeping time</td>
<td>[50, 111]</td>
</tr>
<tr>
<td>Scutellaria barbata D. Don</td>
<td>A1</td>
<td>GC-MS</td>
<td>SA, PCA</td>
<td>86 Compounds</td>
<td>Antimicrobial, protecting liver and biliary</td>
<td>[37]</td>
</tr>
<tr>
<td>Radix Scutellaria Baicalensis</td>
<td>A3</td>
<td>NiR</td>
<td>DA, S-DA</td>
<td>\</td>
<td>\</td>
<td>[51]</td>
</tr>
<tr>
<td>Rhodiolae roseae rhizomes</td>
<td>A3</td>
<td>$^3$H-NMR</td>
<td>PCA</td>
<td>Salidroside, rosavin</td>
<td>Protective effects on LPS-induced acute lung injury*</td>
<td>[52]</td>
</tr>
<tr>
<td>Puerariae lobata</td>
<td>A3</td>
<td>RRLC</td>
<td>PCA</td>
<td>Isoflavonoids</td>
<td>Antioxidant, estrogen-like effect, and treatment of osteoporosis</td>
<td>[53, 112] [113]</td>
</tr>
<tr>
<td>Angelicae acutiloba</td>
<td>A3</td>
<td>Pyrolyser-coupled (PY-GC-MS)</td>
<td>PCA, PLS-DA</td>
<td>\</td>
<td>\</td>
<td>[54]</td>
</tr>
<tr>
<td>Polygalae japonica</td>
<td>A3</td>
<td>HPLC-DAD-ELSD</td>
<td>PCA, COW</td>
<td>Kurarinel</td>
<td>Relative inhibition or inhibition of phosphodiesterase 3,4,5*</td>
<td>[55]</td>
</tr>
<tr>
<td>Sophora flavescens</td>
<td>A3</td>
<td>NMR</td>
<td>PCA</td>
<td>\</td>
<td>\</td>
<td>[56]</td>
</tr>
<tr>
<td>Tussilaghi farfara</td>
<td>A4</td>
<td>GC-MS</td>
<td>PCA</td>
<td>Fifty-four metabolites were identified, including 35 polar metabolites and 19 nonpolar compounds</td>
<td>\</td>
<td>[59]</td>
</tr>
<tr>
<td>Atractylis chinensis DC</td>
<td>B</td>
<td>HPLC</td>
<td>PCA, IONN, LDA</td>
<td>\</td>
<td>\</td>
<td>[81]</td>
</tr>
<tr>
<td>Aconiti kusnezoffii</td>
<td>A1</td>
<td>HPLC</td>
<td>HCA</td>
<td>Mesoacanthine, acanthine, hypaconitine</td>
<td>Voltage-gated sodium channel agonists, immunologic*</td>
<td>[38]</td>
</tr>
<tr>
<td>Fuzi</td>
<td>B</td>
<td>UPLC-Q-TOF-HDMS</td>
<td>PCA, PLS-DA, OPLS-DA</td>
<td>Aconitine, mesoacanthine, hypaconitine, deoxyacanthine, 10-OH-mesoacanthine</td>
<td>Voltage-gated sodium channel agonists, immunologic*</td>
<td>[61]</td>
</tr>
<tr>
<td>Two Carmichaeli plants</td>
<td>A1, B</td>
<td>UPLC-Q-TOF-HDMS</td>
<td>PCA, S-plot</td>
<td>22 Types of alkaloids</td>
<td>Voltage-gated sodium channel agonists, immunologic*</td>
<td>[29]</td>
</tr>
<tr>
<td>Radix Rehmanniae</td>
<td>B</td>
<td>UHPLC-TOFMS</td>
<td>PCA, OPLS-DA</td>
<td>Leunonride, its isomer, S-GCME</td>
<td>Antitumor, antiadipic, neuro-protection*</td>
<td>[77]</td>
</tr>
<tr>
<td>Ligustrum lucidum</td>
<td>B</td>
<td>UPLC-QTOF-MS</td>
<td>PCA</td>
<td>Ligustiliside B</td>
<td>Antidiabetic, antioxidant</td>
<td>[78, 114]</td>
</tr>
<tr>
<td>Fructus Xanthii</td>
<td>B</td>
<td>GC-MS</td>
<td>PCA</td>
<td>14 Polar compounds</td>
<td>Treatment of cramping and numb-ness of the limbs, ulcer, sinusitis, cataracts and pruritus</td>
<td>[80]</td>
</tr>
</tbody>
</table>


Pharmaceuticals

The last step in the production of Chinese medicinal preparations, obtaining plant extracts, can also be subjected to quality control using metabonomic profiles. The extract is a contributing factor to the quality and toxicity of the drug produced. However, a prerequisite is that the extracts of intermediate products that are to be analyzed are well documented with regard to the production steps they have been subjected to. Bioactive compounds may be identified if it is possible to obtain or generate extracts of different materials from the same plant species that are highly variable in bioactivity. PCA may then be used to discriminate the chemical fingerprints of the extracts in a way that separates them by their activity or by spatial origin, and relevant chemical compounds can subsequently be deduced from their contributions to the respective fingerprints [82, 83]. The majority of TCM products for oral use are applied as water decoctions [84]. Other oral preparations include macerates in aqueous ethanol and powdered drugs suspended in water or prepared in pills, with honey, water, or rice gruel as an excipient [85, 86]. An HPLC fingerprinting analysis was developed to assess the quality and comparative contents of cinnamon bark and cinnamon twig components. PCA and PLS-DA allowed good discrimination of these samples, and cinnamaldehyde was found to be the most abundant marker component [87]. Several examples are found in the literature of utilizing HPLC together with different chemometric methods for the analysis of complex mixtures, including the resolution of HPLC fingerprints of complex, many-component substances found in Huoxiang Zhengqi tincture samples from a batch from a given manufacturer, or from different producers [88]. Another example of investigating complex mixtures is the analysis of nine bioactive compounds from a Yiqing preparation which is composed of three TCMs, to assess the consistency of the quality among 12 manufacturers based on SA. The results showed that...
HPLC fingerprinting could serve as the first tool for revealing the consistency of the quality of Yiqing via similarity comparisons [89]. LC-LTQ-Orbitrap MS was applied for the simultaneous identification and quantification of multi-constituent Xin-Ke-Shu, a TCM preparation [90].

It is complicated during the pharmaceutical process. On the one hand, the pharmaceutical excipients hold back the analysis. On the other hand, a Chinese patent drug usually consists of many sorts of herbs; the constituents’ analysis is a bottleneck in quality control. Plant metabolomics, a platform aimed at the complex ingredients, can help to better understand the nature of these problems.

Conclusions and Perspectives

Plant metabolomics can be applied in the discrimination, processing, and pharmaceutical preparation steps of TCM products, which represent the entire production process (Table 2). As the first step in the production of Chinese medicinal preparations, the identification of plant varieties used, involving species, parts of the herbs, cultivated geographic region, and planting period, is of great significance for ensuring the safety and effectiveness of clinical treatment since the quality and contents of the active constituents depend on these factors. Adulterants are assumed to be distinguishable from plant materials and play the role of challenging substances. The processing, as the second step in production, appears to be of significance in clinical applications and has been proven to satisfy the requirements of therapeutics. As different forms of TCMs show different pharmacological actions, it is pertinent to administer the correct form of herbs to avoid any undesirable consequences. Therefore, it is of paramount importance to characterize the specific form of TCM. Metabolomics can also be applied to control the content of extracts, which is crucial to the pharmaceutical production. The complexity and diversity of the components of TCM preparations make qualitative analyses difficult. In the pharmacopoeia, single-component analysis is used on TCM mostly. Nevertheless, it lacks representativeness. In contrast to single-component analyses, metabolomics achieves comprehensive scans, addressing some of the shortfalls of single-component analysis. As shown by the three points we have just illustrated, plant metabolomics can play a vital role in quality assessments during the large-scale production of TCM preparations.

To facilitate the application of plant metabolomics in the quality assessment, on the one hand, we are supposed to utilize the new and effective techniques so that they will support metabolomics studies adequately. On the other hand, exploring potential research points in TCM seems necessary. With the appearance of RRLC and UFLC, the methods of analysis come to a new era. Shorter analysis time and the more efficient separation are the advantages of these methods [91]. MAS-NMR, another sharp technique, enhances the resolution of solid samples and plays an important role in the overlap of peaks [92]. Cristina Daolio et al. [93] applied MAS-NMR classifying commercial catuaba successfully. An LC-MS-NMR platform was demonstrated, which combines two innovations in microscale analysis, nanoSplitter LC-MS and microdroplet NMR, for the identification of unknown compounds found at low concentrations in complex sample matrices as frequently encountered in metabolomics or natural product discovery [94]. Therefore, in the future, employing new and effective chromatographic or spectroscopic techniques for metabolomics studies seems to be an important tendency. What concerns exploring potential research points, in recent years, metabolomics was reported to the study of pharmacology in some terms. Employing a metabolomics platform, Yiqing Lin et al. [94] identified the active cyanobacterial metabolite. Kashif Ali et al. [95] also managed to screen the anti-TNFα activity in crude extracts of grapes and other berries by NMR spectroscopy and chemometric. Using this approach, compounds related to activity can be identified without extensive and elaborate chromatographic separation, and it thus allows rapid identification of extracts with biological activity. Moreover, screening the active compounds and effective parts seems to grow into a vital process in future studies. As a consequence, the results of quantitative assays will become more instructive and convincing. In a word, with the development of analysis methods and the exploration of potential research points, the application of metabolomics in TCM quality assessment tends to become more prevalent and considered in the future.

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Conflict of Interest

No conflicts of interest exist.

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