HPLC in Natural Product Analysis: The Detection Issue

Jean-Luc Wolfender
Laboratory of Pharmacognosy and Phytochemistry, School of Pharmaceutical Sciences, University of Geneva, University of Lausanne, Geneva, Switzerland

Abstract
High-performance liquid chromatography (HPLC) is a very powerful and versatile chromatographic technique for the separation of natural products (NPs) in complex matrices, such as crude extracts for selective detection and quantification or general profiling. The method is widespread and has been adapted to the analysis of a broad range of NPs generally without the need for complex sample preparation. The choice of the appropriate detection method in HPLC is crucial because of the diversity of NPs and the fact that there is no single technique for their efficient detection. In this review both qualitative and quantitative applications of HPLC with UV, DAD, FLD, ECD, RID, CAD, CL, ELSD, CAD, MS, MS-MS, and NMR are covered to provide a general, rather than an exhaustive, overview. The potential and limitations as well as some new trends in HPLC hyphenation are discussed.

Abbreviations
- APCI: atmospheric pressure chemical ionisation
- API: atmospheric pressure ionisation
- APPI: atmospheric pressure photoionisation
- CAD: charged aerosol detection
- CapNMR: capillary NMR
- CID: collision-induced dissociation
- CL: chemiluminescence
- DAD: photodiode array detection
- ECD: electrochemical detection
- EI: electron impact
- ELSD: evaporative light scattering detection
- ESI: electrospray ionisation
- FD: fluorescence detection
- FID: flame ionisation detection
- FT: Fourier transform
- HCA: hierarchical clustering analysis
- HPLC: high-performance liquid chromatography
- HR: high resolution
- LOD: limit of detection
- LOQ: limit of quantification
- LR: low resolution
- MS: mass spectrometry
- MS-MS: tandem mass spectrometry
- MSn: multiple-stage mass spectrometry
- MVDA: multivariate data analysis
- NI: negative ion
- NMR: nuclear magnetic resonance
- PCA: principal component analysis
- PI: positive ion
- Q: quadrupole
- QQ: triple quadrupole
- RID: refractive index detection
- SPE: solid-phase extraction
- TOF: time of flight
- UHPLC: ultra high-pressure liquid chromatography
- UV: ultraviolet

Introduction
Natural products (NPs) have provided the inspiration for most of the active ingredients in medicines. The reason for this success in drug discovery can probably be explained by their high chemical diversity, the effects of evolutionary pressure to create biologically active molecules, and/or the structural similarity of protein targets across many species. This large chemical diversity [1] is also directly linked to a high variability of their intrinsic physicochemical properties that render the universal detection challenging. None of the available HPLC detectors are able to detect all the NPs in a given extract within a single analysis. This limitation is well-known in me-
tabolomics [2], which aims at measuring all of the metabolites in an organism qualitatively and quantitatively. The complex and convoluted nature of the crude extracts analysed (from plants, marine organisms, fermentation broths, etc.) also contributes to the difficulty of NP detection. Furthermore, the analytes can be present in large or very small amounts, and, according to the type of study (quantification, standardisation, fingerprinting, screening, trace analysis, etc.), very sensitive and selective methods may be needed for their detection.

The analysis of individual NPs in complex crude extracts requires efficient separation methods prior to their detection. In this respect, high-performance liquid chromatography (HPLC) has been recognised since the early 1980s as the most versatile technique for the efficient separation of NPs directly in crude mixtures without the need for complex sample preparation [3]. HPLC has greatly developed through the years in terms of convenience, speed, choice of column stationary phases, high sensitivity, applicability to a broad variety of sample matrices, and ability to hyphenate the chromatographic method to spectroscopic detectors [4]. From the chromatography viewpoint, the development of columns with different phase chemistry (especially reversed phase) enabled the separation of almost any type of NPs. The latest developments of HPLC, including the recent introduction of very pH-stable phases, sub-2-μm particles [5], and monolith columns, have considerably improved the performances of HPLC systems in terms of resolution, speed, and reproducibility. Efficiencies exceeding 100,000 plates and peak capacities over 900 can be attained by coupling columns together [6].

In UV detection, the relationship between the intensity of light transmitted through the detector cell and the solute concentration of NPs with weak chromophores [10]. However, several mobile-phase constituents that exhibit high UV cut-offs should be avoided because they might blind the detection of NPs with weak chromophores [10]. In UV detection, the relationship between the intensity of light transmitted through the detector cell and the solute concentration.

### Table 1 Performance of the different detectors for HPLC

<table>
<thead>
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<th>Type</th>
<th>Dynamic range</th>
<th>Sensitivity</th>
<th>Specificity</th>
<th>Chromatographic compatibility</th>
<th>Structural information</th>
<th>Ease of use</th>
<th>Wide-spread use for NPs</th>
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Note: Uni.: universal; Spec.: specific.

- Non-linear response.
- **b** NMR provides absolute quantification but is not used in this way in the on-flow mode.
- **c** Requires solvents and buffers with low UV cut-offs.
- **d** Not compatible with gradients.
- **e** Usually not compatible with non-volatile buffers.
- **f** Need deuterated water and deuterated modifiers in the on-flow mode.
- **g** Relatively new technique.
tion is given by Beer's Law [11]. The two factors that control the detector sensitivity are the magnitude of the extinction coefficient of the analyte of interest at a given wavelength and the path length of the light passing through the UV cell. The sensitivity will increase with increasing path length, but a compromise must be found with the cell volume in order to avoid peak dispersion [8].

Three types of UV detectors are available: fixed wavelength, multiple wavelength, or photodiode array (DAD). The fixed-wavelength detector is the least expensive and has higher intrinsic sensitivity because the light is emitted at specific wavelengths with given lamps. However, the multiple-wavelength detector is more versatile and can often compensate for its lower sensitivity when a wavelength with the highest extinction coefficient for the solutes of interest is chosen. The photodiode array will be treated in the hyphenation section, as it can provide complete UV spectra of the constituent during separation. Limits of detection (LODs) can reach 10⁻⁸ g/ml, with a linear dynamic range of about three orders of magnitude.

Often, the separation of crude extracts requires a gradient elution which cause a shift of the baseline at low wavelengths that can be avoided when working with modifiers having low UV cutoffs such as phosphate buffers or trifluoroacetic acid (TFA). These modifiers, however, are not optimal in multi-hyphenated systems with HPLC-MS (not volatile, ion suppression) and should be avoided [12].

Many publications refer to the application of HPLC-UV at a single wavelength for the detection of nearly all NPs having a chromophore for both profiling and quantification purposes. An exhaustive survey would exceed the frame of this paper. Many applications for flavonoids, terpenes, alkaloids, coumarins, alkamides, and polyacetylene have been summarised in a review [13]. Some selected quantitative methods are discussed here.

Being simple and inexpensive, HPLC-UV is used in several pharmacopoeias for the quantification of individual NPs in the quality control of herbal drugs or phytopreparations. In the U.S. and European pharmacopoeias [14], [15], HPLC-UV (203 nm) methods are used for the determination of the total ginsenoside content of *Ginseng* roots and the measurement of the ratio of the ginsenosides, Rg1 and Rb1 [16]. This analysis is indeed important to assess pharmacological effects, as a low Rg1/Rb1 ratio is linked to the calming properties of American ginseng, while the other active ingredients in *Ginkgo* are the triterpen triacetates, which are standardised at 6% in the extracts. These terpenes exhibit rather weak chromophores (*Ginkgo*) and are difficult to detect in the presence of the strongly UV-absorbing flavonoids [23]. They have thus successfully been quantified by other methods such as ELSD [24] (see below). In *Ginkgo*, the level of highly allergenic alklyphenols (ginkgolic acids, cardanol, and cardols) also needs to be assessed, and a limit of 5 ppm has been proposed in the EU and U.S. pharmacopoeias. An in-depth analysis requires GC-MS [25] or HPLC-MS [26], but their total amount can be determined by a simple HPLC-UV (210 nm) analysis without enrichment procedures after peak purity assessment by HPLC-DAD-MS [27].

In other cases, for the same plant extract, quantification may require both HPLC-UV and HPLC-MS. This was necessary for the standardisation of the constituents of woad (*Isatis tinctoria*). In this case, when possible, HPLC-UV was preferred to HPLC-MS because of the lower RSD obtained [28].

Recently, other simple HPLC-UV methods for the standardisation of new herbal supplements coming on the market such as *Pueraria lobata*, also known as Kudzu (Japan), have been developed. In this case, a fully validated method following the ICH guidelines for the quantification of the isoflavone content of this plant has been reported [29]. Similarly a method for profiling coumarins in different *Angelica* species has also been validated [30].

HPLC-UV methods also can be used to compare the fingerprint profiles of closely related species or the same species from different locations. This was the case for Herba Oldenlandiae, a plant of the Chinese pharmacopoeia, which can be adulterated by different *Oldenlandia* species. A comparative analysis of chemical components was obtained by software calculation of correlation coefficients of the entire UV chromatograms for appraising similarity [31]. Fingerprint-based similarity also can be employed for chemotaxonomic study by applying hierarchical clustering analysis (HCA) and principal component analysis (PCA) to a set of HPLC-UV profiles, as was performed for different *Taxus* species [32].

In Fig. 1, the profiling of the constituents of the crude extract of the model plant *Arabidopsis thaliana* obtained with UV detection at two standard wavelengths (254 and 350 nm) is illustrated. A selective detection of flavonoids (F) and sinapoyl derivatives (S) was performed at 350 nm. Detection at 254 nm was less selective as more UV active constituents were recorded; this included glucosinolates (G), which have UV maxima at 227 nm. However, fewer peaks were recorded with UV than with MS or ELSD detection methods (see below). In this example, the high chromatographic resolution was provided by ultra-high-pressure liquid chromatography (UHPLC) [5], which yielded very high peak capacities (see inset c in the MS trace) [33], [34].
HPLC-FD (fluorescence detection)

Compared with HPLC-UV, fluorescence detection (FD) affords much greater sensitivity and selectivity (Table 1). In fluorescence, the molecular absorption of a photon triggers the emission of another photon with a longer wavelength. This difference in wavelengths (absorption vs. emission) provides more selectivity, and the fluorescent light is measured against a very low-light background, thus improving the S/N ratio. Unfortunately, relatively few NPs fluoresce in a practical range of wavelengths. However, many substances can be made to fluoresce by forming appropriate derivatives [8].

Compared with UV, FD has been scarcely used for the detection of NPs. Most of the applications are related to the very sensitive detection of aflatoxins in food because this class of NPs contains natural fluorescence [35]. An FD method for the quantification of aflatoxins in herbs also has been recently reported [36]. It combines sample cleanup with immunoaffinity columns followed by HPLC-FD, and it enabled the detection of aflatoxin content ranging from 7 to 20 µg/kg in the plant material. HPLC-FD also has been used for the screening of the major benzophenanthridine alkaloids produced by cell cultures of *Eschscholtzia californica* [37]. Thanks to the selectivity of the method (excitation 330 nm, emission 570 nm), the sample preparation was notably simplified compared with previous methods. The detection of NPs that do not naturally fluoresce has been successfully achieved after the addition of fluorescent tags. This was the case for the specific and sensitive detection of the plant hormone jasmonic acid (LOD 3 ng/mL), which was tagged with the ADAM reagent [38].

HPLC-CL (chemiluminescence)

Similar to fluorescence, chemiluminescence can be defined as the emission of light from a molecule or atom in an electronically excited state produced by a chemical reaction at an ordinary temperature without any associated generation of heat. Chemiluminescent (CL) nitrogen detection is a relatively new technology for the detection of nitrogen-containing molecules, including a broad range of pharmaceuticals with very high sensitivity up to the femtogram range [39]. Because many compounds are not chemiluminescent, they can only be detected chemiluminescently after being derivatised. The development of chemiluminogenic reagents has helped to extend the application of CL...
HPLC-ECD (electrochemical detection)

Many NPs have electroactive groups and are readily measurable and detectable by LC with electrochemical detection (ECD). This detection method is inexpensive, sensitive, selective, and widely accepted (Table 1). It is less selective than FD; however, NPs with electrochemical activity are common, and the technique can be applied to a large number of analytes in either the oxidation or reduction modes. Numerous functional groups are sensitive to oxidation, including phenols, aromatics, amines, thiols, and quinolines, and many are compatible with reduction, such as olefins, esters, ketones, aldehydes, ethers, and quinones. ECD differs from other methods of detection because it alters the sample. Cells for HPLC-ECD usually consist of three electrodes, the working, counter, and reference electrodes, which can be aligned in several different geometries, although two main designs exist. In coulometric systems, the eluent is directed through the electrode, while in amperometric systems, the eluent passes over it. ECD detection is usually performed by maintaining the potential of the working electrode at a fixed value relative to the potential of the electrolyte, which is measured by the reference electrode. The fixed potential difference applied between the working and the reference electrodes drives the electrochemical reaction, and the resulting current is measured as a function of the elution time. This allows for detection limits at the pmol level. The selectivity of ECD depends on the accessible potential range, the number of compounds that are active in this range, and the half-widths of the individual signals [10]. Pulsed techniques, which use multi-step potential-time waveforms to realise amperometric/coulometric detection while maintaining uniform and reproducible electrode activity, are also available for detection [42].

Modern interest in HPLC-ECD is stimulated by the recognition that electrochemistry is ideal for the study of the metabolism of aromatic compounds. However, proper use of LC-ECD requires knowledge of redox reactions and their dependence on mobile-phase composition. In this respect, the study of electrode reaction mechanisms can sometimes aid the elucidation of the mechanism of their interaction with living cells, such as for antioxidant NPs [43]. The interest in the method resides also in the fact that the ECD can be easily miniaturised and used in fully integrated chip-based systems.

HPLC-ECD is commonly used in many pharmaceutical, clinical, and biochemical laboratories [10], [44]. Because antioxidants are electrochemically active, HPLC-ECD is well suited for their accurate quantification. In this respect, the method has permitted the detection and quantification of very low levels of antioxidants, such as carnosic acid in food ingredients [45]. In plant extract profiling, coulometric electrode array detection has allowed for the fingerprinting of 25 constituents (polyphenols and tanshinones) in Salvia miltiorrhiza after a careful selection of the optimal mobile-phase pH and working potentials [46]. The successful detection of various small phenolics, such as camaric or ferulic acids, with a gold polycrystalline electrode has been reported recently [47]. This method was also used for the quantification of the vanilloid content in Capsicum annuum extracts [48]. ECD was found to be more sensitive than UV for the normal-phase HPLC profiling of diarylheptanoids in Alpinia officinarum (lesser galangal) [49]. Similarly, the simultaneous determination of hyperforin and total hypericin in Hypericum perforatum was achieved after the photoconversion of the protoforms of hypericins in extracts and tablets [50].

The antioxidant potential of polyphenols can be estimated by HPLC with coulometric array detection because they are reducing agents, and their electrochemical responses result from the donation of electrons. Some degree of correlation between the electrochemical profiles of 17 flavonoids and 3 cinnamic acid derivatives and 3 different antioxidant assays was established [52]. HPLC-ECD also has been used in many applications related to the analysis of drugs in body fluids [44], [10]. Artemisinin, for example, was selectively detected and quantified in plasma using a glassy carbon electrode by reductive ECD [52]. Similarly, acteolside was analysed by amperometric detection in rat blood and brain microdialysate in a pharmacokinetic study [53]. Recently, different amino acid derivatives characteristic for Korean ginseng were analysed by pulsed amperometric detection in both ginseng and plasma samples [54].

HPLC-RID (refractive index detection)

The descriptors described previously are selective for analytes having specific spectroscopic or physicochemical properties. In order to obtain a more universal response, other detectors have been developed. The first, most simple, and least expensive of these detectors is the refractive index detector (RID), which was reported by Tiselius and Claesson [55] in 1942. It was indeed the first effective inline LC detector. However, RID lacks sensitivity, is very susceptible to changes in ambient temperature, pressure, and flow rate, and cannot be used for gradient elution. Despite these disadvantages, this detector has been useful for detecting compounds such as carbohydrates and polymers.

Recent applications in the NP field are very scarce, although this method was used for the quantification of sugars in strawberry fruits [56]. HPLC-RID was also used to determine the sugar concentration in different dry extracts of Hypericum perforatum after extensive SPE cleanup [57]. More recently, the terpene trilactones of Ginkgo biloba were successfully quantified by HPLC-RID after a very specific sample preparation based on the ionisation of these terpenes (RSD < 3%, LOD 0.1 μg, linear range 0.1 – 12 μg) [58].

Because of its inherent limitations, HPLC-RID has been favourably replaced by other universal detection methods such as those described below.

HPLC-ELSD (evaporative light-scattering detection)

Evaporative light-scattering detection (ELSD) was introduced in 1966 by Ford and Kennard [59]. ELSD is a quasi-universal detector for liquid chromatography, as it can detect any analyte less volatile than the mobile phase, regardless of the optical, electrochemical, or other analyte properties [60]. In this detector, the eluent is nebulised using a flow of nitrogen, and the resulting aerosol is transported through a heated drift tube where the vol-
atitle components and solvents are evaporated. The remaining solid fraction is subsequently introduced into a detection cell where a light beam is directed onto the particles that will cause scattering of the incident light detected by a photodiode or a photomultiplier. The most important parameters affecting the ELSD signal response are the nebuliser gas flow rate and the drift tube temperature [61]. The gas flow rate influences the droplet size of the column effluent before evaporation occurs. Higher flow rates result in the formation of smaller aerosol droplets and less scattering of light, with subsequent lower sensitivity, but they allow for a more stable baseline. The drift tube temperature facilitates the evaporation of the nebulised aerosol so that the light-scattering response of the non-volatile solute can exclusively be determined. Higher temperatures are needed for the mobile phase of high-aqueous-content or non-volatile solvents versus semi-volatile ones. However, the new generation of ELSD is able to vapourise the eluent at low temperature, facilitating the detection of semi-volatile analytes. It is pertinent to optimise these parameters to ensure that the optimal signal to noise ratio (S/N) is achieved [62]. ELSD is a mass-dependent detector, in contrast to UV, which is a concentration-dependent detector, and the generated response does not depend on the spectral or physicochemical properties of the analyte. In theory, this means that ELSD generates a similar response for equal amounts of mass present and thus a universal response factor. However, the ELSD response also depends on the volatility of the compound and the mobile-phase composition (in the case of gradient elution), which makes such quantification procedures experimentally inaccessible. Thus, this interplay of several factors leads to a non-linear response, which is one disadvantage of this type of detector and ultimately renders direct linear regression for making calibration curves inaccurate [63]. In order to avoid the variation generated by the gradient elution, the post-column addition of mobile-phase flow having an opposite composition is an interesting alternative [64].

The increased interest in ELSD as a universal detector is that it is more compatible with gradient elution than RID, and it is much less costly and easier to maintain than an HPLC-MS platform (see below). However, it is used mostly as an additional detector rather than a substitute. Recent papers have discussed advantages and potentialities as well as several limitations of the method for both NP and pharmaceutical applications [60], [63], [65].

In applications related to NPs, ELSD has been used mainly for the detection of compounds with weak chromophores such as terpenes, in both aglycone [62] and glycosidic forms [66], saponins [67], and some alkaloids [68]. The method has also been applied in complement to HPLC-MS for surveying NP libraries [69]. In this case, a wide range of products, including UV-active phenolics, were screened, and an ELSD signal was obtained for all compounds (>100 ng injected). ELSD has been used for the quantitative determination of the relevant active principles of herbal drugs. A recent method for the detection of 14 ginsenosides in Korean red ginseng, using digoxin as an internal standard, has been validated and applied to several ginseng products [67]. All calibration curves showed good linearity using a log-log transformation ($R^2 > 0.9833$), and LODs of 0.075–0.30 µg and LOQs of 0.125–0.80 µg were achieved. Thus, ELSD represents an interesting simple alternative to the UV detection method (203 nm) of the pharmacopeia. However, both UV and ELSD detections are often necessary for profiling extracts, as was shown for the simultaneous determination of triterpenoids (ELSD) and phytoecdysones (UV) in roots of *Achyranthes bidentata* (Amaranthaceae) [70].

The weakly UV-active ginkgolide and bilobalide of *Ginkgo* were also efficiently quantified by ELSD [71], [62] Camponovo et al. [71] noticed a specific response of ELSD for terpene triactones versus flavonoids when UV detection (220 nm) and ELSD detection were compared. Dubber et al. [62] were able to obtain a baseline separation of these compounds within 14 min, and they have analysed several *Ginkgo biloba* products with LOQs of ca. 60 ng. Inter-day percentage standard deviations were all better than 6%. ELSD thus represents an interesting alternative to the GC dosage usually performed for the standardisation of the terpene content of *Ginkgo* [72].

In the case of the crude extract profiling of *Arabidopsis thaliana*, shown in Fig. 1, the ELSD response provided many more peaks than those detected in UV (254 and 350 nm), especially for the detection of non-polar compounds (mainly lipids). Compounds detected by ELSD in this leaf extract included the notably different galactolipids such as (1), which were not visible in the UV traces.

**HPLC-CAD (charged aerosol detection)**

Charged aerosol detection (CAD) is a new technology for universal HPLC detection, which derived from ELSD and was introduced by Dixon and Peterson in 2002 [73]. The first steps of the operating principle of CAD are roughly comparable to that of ELSD. Contrary to ELSD, however, with CAD, the dried particle stream is charged with a corona discharge needle, and the resulting electrical charge is measured by an electrometer [63]. The use of a corona discharge indicates some commonality with APCI-MS (see below); however, CAD operates by detecting charge particles with a selected mobility range rather than measuring individual gas-phase ions as in MS. Generally, CAD is more sensitive than ELSD (Table 1), but this difference is strongly influenced by the LC modifier, as ELSD seems less sensitive to increases in salt concentration [63]. This detection method provides a uniform response to non-volatile analytes independently of their nature and a dynamic range up to 4 orders of magnitude [64].

For the detection of NPs with strong chromophores, UV usually performs better than CAD or ELSD. However, these latter detection methods are more dedicated to the analysis of non-UV-absorbing compounds, and of course their performances are in this case much better [63].

Because it is a new detection method, CAD has not yet been used for NP detection in crude extracts to our knowledge, although it has been used for a wide range of pharmaceutical products. Both CAD and ELSD were evaluated for a library of more than 700 compounds in the 150 to 700 Da range and failed only in the detection of two compounds having small MWs [74]. It is noteworthy that successful quantitative analysis of triacylglycerols (TGs) in plant oils has been achieved [75]. The method is advantageous in that no normalisation of the peak area is needed, as is the case in HPLC-MS, where large differences in responses can occur.

**HPLC-FID (flame ionisation detection)**

While considering a universal detection method for HPLC, it is worth mentioning that FID, which is a general detector for GC because it detects all carbon-containing molecules, also has been applied to HPLC. However, in both normal- and reversed-phase modes, the LC mobile phases contain organic molecules that will blind the FID [76]. Thus, HPLC-FID coupling implies working with 100% water mobile phases. A major problem is that water is a very weak mobile phase in RPLC at room temperature. However, for selected compounds and with appropriate
stationary phases, separations were realised with pure water at ambient temperature [77]. Another alternative is to work with superheated water in the HTLC mode because, when heated under pressure to temperatures between 80°C and 250°C, the polarity of liquid water decreases markedly to the extent that it can replace the organic modifier and act as the sole eluent [78]. While successful applications of HTLC-FID have been described [79] mainly for the separations alcohols, carbohydrates, and amino acids, the method has been used very rarely.

HPLC-MS (mass spectrometry)

The application of HPLC-MS in the analysis and characterisation of natural products has been recognised as a major breakthrough [80]. Indeed, while it is expensive, the use of a mass spectrometer as an HPLC detector provides excellent sensitivity and selectivity for the analysis of NPs in complex biological matrices. Furthermore, MS detection provides important structural information online such as MW, molecular formula, and diagnostic fragments, which are crucial for dereplication and rapid online characterisation of NPs [see below]. Historically, a main challenge in HPLC-MS was that high liquid flows from HPLC were not compatible with the high vacuum required for the mass spectrometer. Since the early 1980s, a number of different interfaces have been developed to address this issue and overcome these difficulties [81], [82]. One obvious way to prevent the harmful effects of the LC mobile phase for the mass spectrometer was to remove the solvent prior to ionisation. This has led to the development of moving belt interfaces for sample introduction into the low-pressure plasma sources. The advantage of such devices was the possibility to generate electron impact (EI) spectra, for example, for flavonoids [83], but they suffered from many other disadvantages and were abandoned. Another more modern “transport-type” interface in LC-MS is the particle beam [84], but it has scarcely been applied to natural products analysis. A new generation of direct-EI interfaces coupled to nano-HPLC seems promising for generating such information-rich types of spectra [85].

The overwhelming popularity of HPLC-MS today is largely due to atmospheric pressure ionisation (API) interfaces including electrospray ionisation (ESI) and atmospheric pressure chemical ionisation (APCI). For both ESI and APCI, ionisation occurs at atmospheric pressure. Some combination of high voltage and heat is used to provide the ionisation that is needed to produce the ions that are assayed by the MS system. In ESI, the high-voltage field (3–5 kV) produces nebulisation of the column effluent resulting in charged droplets that are directed toward the mass analyser. These droplets get smaller as they approach the entrance to the mass analyser. As the droplets get smaller, individual ions emerge in a process referred to as “ion evaporation”; these ions are then separated by the MS system. In APCI, heat is used to vaporise the column eluent, and then a corona discharge is used to ionise solvent molecules, which then produce the analyte ions via chemical ionisation mechanisms. More recently, a third type of ionisation source, termed atmospheric pressure photoionisation (APPI), has become available. In APPI, heat is used to vaporise the column eluent (similar to APCI), but the ionisation is produced by way of an ultraviolet (UV) lamp that produces 10–eV photons. Depending upon the solvent system used, the 10–eV photons will ionise either the mobile-phase solvent or a dopant (e.g., toluene) added to the column effluent; these ions then produce the analyte ions through various ionisation mechanisms including charge or proton transfer [86].

These API methods generally provide a soft ionisation and mainly molecular ion species in the form of either protonated molecules [M + H]⁺ (positive-ion mode, PI) or deprotonated molecules [M – H]⁻ (negative-ion mode, NI). Different adducts (e.g., [M + Na]⁺ (PI) or [M + HCOO]⁻ (NI)) are also produced, depending on the solutes and the modifiers used. It is important to understand that analyte ionisation is largely compound dependant and is governed mainly by proton affinity. As a rule of thumb for a good approximation, acidic molecules (e.g., carboxylic acids or phenolics) will produce mainly [M – H]⁻ in NI, while bases (e.g., alkaloids, amines) will generate [M + H]⁺ in PI. Compounds such as glycosides will have a high affinity for salts and will tend to form sodium adducts in PI. Molecule fragmentation or partial declustering of the adducts can be induced in one of the high-pressure regions of the ion beam from the source to the mass analyser by collision-induced dissociation (CID) either at the API source (in-source CID) or in conjunction with tandem MS (MS–MS) experiments [82].

For HPLC–MS applications, many types of mass spectrometers can be used. Those that are low resolution (LR), such as the single quadrupole (Q) mass spectrometers, are the most commonly used and least expensive. Those giving high resolution (HR) and exact mass capabilities, such as time-of-flight (TOF) instruments, are becoming increasingly popular because they provide molecular formula information online and very precise ion trace extraction (+/− 0.01 DA) for a specific detection. For measuring structurally relevant fragments or for very specific detection, triple-quadrupole (QQQ) MS–MS systems are the most commonly used. They are particularly useful for bioanalytical assays and for specific quantification purposes in complex matrices through multiple reaction monitoring (MRM) [87]. Ion-trap mass spectrometers have the unique capability of producing multiple-stage MS–MS (MS³) data that may be essential for structural elucidation purposes. In addition to these four kinds of mass spectrometers, there is a growing number of additional types, including hybrid systems that combine LR and HR analysers for specific applications [86], [81].

Besides their widespread use for qualitative analysis [dereplication studies and crude extract profiling (see below)], HPLC–MS–MS and HPLC–MS–MS have been successfully used for the specific detection and quantification of a large range of natural products. While mainly simple quadrupole or triple-quadrupole analysers are considered for quantitative studies, TOF instruments have recently increasingly been used as quantification tools, even though historically they suffered from a narrow dynamic range [88].

A systematic HPLC-MS-ELSD study was conducted on 96 structurally diverse NPs, including phenolics, alkaloids, amides, terpenoids, steroids, and polyketides, with a short HPLC generic gradient (C18, 5–95% ACN:H₂O 0.1% formic acid in 10 min). A good overall ionisation of most compounds with ESI in both PI and NI mode was obtained [69]. The generic separation used was suitable for the elution of 86 of these 96 compounds (RT > 1 min < 10 min); 75 were well detected in PI and 57 in NI, while all compounds were detected by ELSD (4 μg on-column). A relatively universal detection can thus be obtained if both PI mode and NI mode are used or alternated during acquisition (see, for example, the profiling of A. thaliana constituents in Fig. 1). ESI (PI) is used mainly for alkaloids, APCI (PI) for pigments and carotenoids, ESI (PI/NI) for triterpene glycosides, and APCI or ESI (PI/NI) for phenolics and flavonoids. APCI is generally more suitable for non-polar constituents, while ESI provides a softer...
ionisation and gives a good response for polar metabolites. On the other hand, APCI is generally recognised to be less prone to matrix effects than ESI [89]. Only a few studies have referred to the use of APPI for NP detection [90]. An interesting work on the rapid analysis of stilbenes and derivatives from downy mildew-infected grapevine has shown that APPI Ni with acetone as doping reagent was more sensitive than ESI and provided cleaner spectra [91]. Another study on anthocyaninins in wines has shown that both APPI and ESI-MS allow the identification of a great number of these pigments. However, APPI ionisation was found to be more specific towards the analyte, and the background noise was lower [92]. In drug discovery, the performances of APPI, APCI, and ESI were evaluated on 106 standards and 241 proprietary drug candidates occupying a wide chemical space [93]. The results indicated that the detection rates in PI were 94% for APPI, 84% for APCI, and 84% for ESI. When PI and NI mode detection were combined, APPI detected 98% of the compounds, while APCI and ESI each detected 91%. In APPI, for analytes of low polarity and low proton affinity, M+ was the predominant ion species generated, while for high-polarity and high-proton affinity compounds, [M + H]+ ion species dominated the spectra. APPI is thus regarded by some authors [90] as a more universal detection method for HPLC-MS than APCI and ESI, but, at present, it is still not widely used in natural products analysis. Because ionisation is compound dependant in HPLC-MS, the choice of the interface, the ion mode polarity, and the eluent with their HPLC-MS detection in either plant extracts formulation or body fluids. Indeed, while HPLC-UV [94], [95] and HPLC-ELSD [96] methods have been used for the detection of these saponins (see above), HPLC-MS is the method of choice. The latter provides sensitive and selective detection in shorter analysis time, as only isomers need to be separated and MS provides useful structural information for a confirmation of the identity of these glycosides.

ESI (NI) was used for qualitative [97] and quantitative [98] studies mainly on various Panax spp. extracts. In conjunction with multiple-stage MS-MS (MSn) experiments, fragmentation of the [M–H]– provided the identification of 25 ginsenosides [97]. Another study demonstrated that different negative-ion source conditions were necessary for an optimum ionisation of malonylated and non-malonylated ginsenosides [99]. Quantification of the ginsenosides Re, Rb1, and Rc in different dosage forms of ginseng was provided by integration of their specific [M–H]– ion traces [98]. ESI (PI) was also used successfully for the detection and quantification of these triterpene glycosides. Wang et al. reported a quantification method in MS-MS in the single-reaction monitoring mode (SRM) for a very selective dosage of Rb1, Rc, and Re [100]. In this case, both [M + Na]+ and [M + H]+ ions were observed, but the CID MS-MS spectra of the [M + H]+ species were found to be more informative than those of the sodiated adducts. ESI (PI) was also used more recently in other quantification studies of Panax ginseng [101] and root and leaves of Panax quinquefolium [102], with an RSD of about 8% in the latter case. With the same ionisation mode, a newly developed UHPLC-QTOF-MS method has been efficiently used for the metabolite profiling of different Panax species that were efficiently differentiated after PCA of the data [103]. Recently, APCI (PI and NI mode) was also investigated, and 30 ginsenosides were determined. The Ni mode provided [M–H]– ions, while in the PI mode, characteristic thermal degradation ions were recorded [104]. Yu et al. also used APCI (NI) for the quantification of 6 ginsenosides as well as other products in a complex traditional Chinese medicine (TCM) preparation (Naodesheng injection) containing five plants, including Rhizoma Notoginseng [105]. In this case, the addition of 0.1% formic acid into the mobile phase ensured the formation of stable [M + HCOO]– molecular ions. The detection of these ions in the SIM mode produced data with acceptable linearity, repeatability, precision and accuracy (LOQs 0.02–0.2 μg, linearity coefficient > 0.9900, RSD < 3%). It is also noteworthy to mention that ginsenosides have been analysed and quantified in body fluids with similar HPLC-MS methods [106]. Thus, as shown here, depending on the conditions and the matrix studies, both APCI and ESI (PI and NI) enable the quantification and identification of ginsenoside provided that a correct optimisation is insured. Other types of saponins could also be well detected by LC-MS. This was the case, for example, for the evaluation of the steroidal saponin content of Rhizoma Paridis and Chinese multi-herb remedies that contain this plant [107].

Another well-studied plant is Ginkgo biloba, for which both the terpene (ginkgolides and bilobalide) and the flavonoid contents need to be quantified for standardisation purposes. Different HPLC-MS methods have been used for the quantification of the terpenes because their detection in UV is compromised by the strongly UV-absorbing flavonoids (see above). APCI (NI) enabled a direct analysis of several Ginkgo preparations without pre-purification and provided LODs in the picogram range. Because the detection relies on the specific extraction of the [M – H]–, a fast HPLC gradient was applied (14 min), which provided a high-throughput scan of many samples [23]. More recently, ESI (NI) in MRM mode enabled an even more sensitive and specific detection by the monitoring of specific MS-MS transitions of these terpenoids from their precursor [M – H]– ions [108]. Ginkgolides have also been successfully analysed by APCI (NI) in MRM mode in plasma samples for pharmacokinetic studies [109]. Another validated method for the specific detection of these compounds relies on GC-MS [72]; in this case, however, a tedious sample preparation is needed. Because of its high sensitivity and selectivity, HPLC-MS also represents a key technique for the detection of toxic compounds present at trace levels. For example, HPLC-MS-MS was evaluated for the quantification of urushiol congeners, which are responsible for poison oak dermatitis. ESI (NI) in the MRM mode yielded LODs in the low ppb range that are relevant for toxicological, clinical, and environmental studies of these allergens [110]. Similarly, pyrrolizidine alkaloids, a class of hepatotoxic and tumorigenic phytotoxins, have been analysed in honey samples and in extracts of Echium plantagineum. ESI (PI) provided good detection limits for these compounds; however, previous sample enrichment with strong cation-exchange SPE was mandatory [110]. The consumption of slimming regimens made of Chinese herbal preparations has caused intoxications associated with species of the Aristolochia genus, which contain highly nephrotoxic and carcinogenic metabolites called aristolochic acids. Their detection in commercial dietary supplements was successfully achieved in suspect preparations by using both UV and MS detection [112]. A recent LC-MS-MS method enabled the simultaneous detection of aristolochic acid I and the marker compounds fanghongnine and tetrandrine in many TCM preparations [113].
In Fig. 1, the type of detection provided by ESI in both NI and PI modes for the profiling of Arabidopsis thaliana constituents is compared with the detection obtained by the UV and ELSD methods. As shown, very complex MS base peak ion chromatograms (BPIs) were obtained in both PI and NI mode. The number of constituents detected was higher than that observed in UV and ELSD traces. It is noteworthy to observe that PI and NI modes do not generate similar chromatograms and that some of the compounds were detected in only one of the ionisation modes. This demonstrates that MS represents a rather universal detection method of crude extract profiling if both PI and NI mode are used complementarily. On the other hand, very specific detection can be obtained if the extracted ion chromatogram of a given ion is displayed. For example, the ion trace m/z = 741.22 has been selected in Fig. 1. This ion corresponds to the protonated molecule of a flavonoid triglycoside F having the molecular formula C_{33}H_{41}O_{19}. Because TOF was used for detection in this example, a molecule of a flavonoid triglycoside F having the molecular formula chloro-4-fluoroaniline in rat body fluids [118].

This was the case for studying the metabolic fate of 3-fluoroaniline. This was also the case for studying the metabolic fate of 3-fluoroaniline [117]. However, it is not used for this purpose, mainly because of its lack of sensitivity and the high costs associated with the use of this technique (Table 1). HPLC-^{19}F-NMR has been used for the specific detection of fluorinated derivatives. This was the case for studying the metabolic fate of 3-chloro-4-fluoroaniline in rat body fluids [118].

HPLC-NMR (nuclear magnetic resonance)
HPLC-NMR is used mainly for providing structural information online (see below), but, as ^1H-NMR can detect all analyte-bearing protons, HPLC-NMR also can be regarded as a universal detection method [117]. However, it is not used for this purpose, mainly because of its lack of sensitivity and the high costs associated with the use of this technique. HPLC-^{19}F-NMR has been used for the specific detection of fluorinated derivatives. This was the case for studying the metabolic fate of 3-chloro-4-fluoroaniline in rat body fluids [118].

Detection Methods for Online Structural Identification

Some of the spectroscopic methods used for the detection of NPs (DAD, MS, NMR, CD, IR) also can be used in hyphenation with HPLC to generate structurally relevant data for their online identification and dereplication purposes. When both chromatographic and spectroscopic dimensions are used, the methods are referred to as “LC-hyphenated techniques.” Dereplication of NPs with these methods is strategic for guiding an efficient lead-finding process [119]. However, this task is not easy to perform, because no exhaustive LC spectroscopic databases are available. Furthermore, even if libraries of NP spectra could be efficiently generated on a given LC-hyphenated system, their widespread use is complicated by the fact that, contrary to GC-MS, most of the data, especially HPLC-MS and HPLC-MS-MS, are instrument dependant [120]. Thus, for each new compound, both the order of the atoms and the stereochemical orientations have to be elucidated de novo. Consequently, a single LC-hyphenated technique does not exist that is capable of identifying online all secondary metabolites in a plant extract [121]. Ideally, hyphenated methods should provide the same quality of spectroscopic information as that acquired from isolated pure products. However, this is not the case, and each technique presents its limitations. The coupling of various hyphenated methods together (hyphenation) requires even more compromises to be made (compatibility of the eluent composition, sensitivity and dynamic range of the methods being hyphenated), but it has the advantage that all structural information is obtained within one single analysis [122].

For natural product analysis, the dereplication procedures rely mainly on HPLC-DAD-MS [82] and chemotaxonomic information provided by NP databases [123]. When unknown natural products cannot be dereplicated by these means, complementary information can be obtained by NMR either directly hyphenated with LC (HPLC-NMR) [124] or used at-line with preconcentration methods such as HPLC-SPE-NMR [125], [126] or after microfractionation with microflow HPLC-NMR methods such as CapNMR [119], or even with a combination of both techniques by CAP-SPE-NMR [127]. Other hyphenated methods, such as HPLC-IR [128] or HPLC-CD [129], can bring useful additional structural information.

The goal is to cover HPLC hyphenation exhaustively here, as a dedicated review on plant fingerprinting can be found in this issue. However, the methods will be discussed briefly to complete the aspects regarding HPLC detection.

HPLC-DAD (photodiode array detection)
Photodiode array detection (DAD) [130] provides UV spectra directly online and is particularly useful for the detection of natural products with characteristic chromophores [131]. For example, polyphenols can be efficiently localised by this method because they possess characteristic chromophores. With this type of compound, DAD-UV spectral libraries can be built and used for dereplication, but compounds have to be analysed under the same HPLC conditions, as the composition of the mobile phase might affect the UV bands slightly. Furthermore, the choice of modifiers affects UV detection and might blind the bands in the low-wavelength range. The possibility to acquire several UV spectra across a given LC peak allows the assessment of peak purity [131].

HPLC-DAD analysis has been applied alone for the partial characterisation of many NPs with characteristic chromophores, mainly phenolics [132], polyketides, alkaloids, and terpenoids [131]. The technique also has been used successfully in combination with UV shift reagents, which are classically used for the structural characterisation of flavonoids in their pure form [119], [133]. Comparison of the genuine and shifted online DAD spectra enables a precise localisation of the hydroxy groups on the polyphenols online. Another interesting point regarding DAD-UV detection is that all wavelengths are stored during analysis, and thus multiple wavelengths can be monitored at the same time for detection of different classes of compounds. This has been found to be very useful for simultaneous detection of the naphthodianthrones (590 nm) and chloroglycosides (270 nm) of Hypericum perforatum [18] or for the relatively selective determination of glucosinolates (227 nm), indolic derivatives (270 nm), flavonoids and cinnamic derivatives (325 nm), and anthocyanidins (520 nm) in different Brassicaceae [134]. As for simple HPLC-UV, applications of HPLC-DAD in the analysis of NPs are manifold [82], [131] and cannot be extensively dis-
discussed here. HPLC-DAD systems are commonly used in multi-hyphenated systems with MS [135] or ELSD [70] detection methods.

In Fig. 1 (inset a), an example of a UV-DAD spectrum of a flavonoid (F) recorded online in the profiling of the constituents of Arabidopsis thaliana is shown. The presence of two maxima at 280 and 360 nm attributed, respectively, to the benzoic (band II) and cinnamoyl (band I) chromophore moieties was characteristic for this type of compound. The complete dereplication was achieved thanks to the complementary MS information obtained online (see below).

HPLC-MS, MS-MS, and MS^n
HPLC-MS represents a key technique for the online identification of natural products. Besides detection, a mass spectrometer gives the possibility of generating either nominal mass molecular ions or accurate mass measurements for the determination of empirical formulas [86]. Furthermore, the use of tandem or hybrid MS instruments [80] provides in-depth structural information through fragmentation of the molecular species by CID reactions. Several reviews describe aspects related to online NP identification for screening and dereplication [82], [114], [115], [119], [136], for quality control, for fingerprinting for authentication or standardisation [80], or for the identification of biomarkers [137].

For online dereplication purposes, the determination of molecular weight is of great importance. This, however, necessitates the comparison of MS data obtained under different detection conditions in order to differentiate protonated or deprotonated molecules from adducts or fragments [119], [138]. The use of high-resolution instruments, such as a TOF-MS [34], [103] or FT-MS [139], [140] system, enables direct determination of the molecular formula of crude mixtures. This strategic information allows for more precisely targeting the search in natural product libraries for dereplication purposes [119].

Complementary structural information can be generated by in-source collision-induced dissociation (CID) in HPLC-MS-MS or MS^n experiments [141]. The generated CID spectra are, however, not comparable to those recorded by EI, and this hampers a direct use of the standard EI-MS libraries for dereplication purposes. For performing automated dereplication procedures, specific MS-MS libraries have to be built based on standards available in a given laboratory, which consequently limits this approach. The development of hybrid instruments combined with high-resolution spectrometers, such as TOF, FT, or orbitrap analysers, allows the possibility to record MS-MS spectra with high mass accuracy, facilitating the interpretation of the obtained spectra [142].

In natural product chemistry, MS-MS spectra are mainly useful for the partial determination of sugar sequences of various glycosides [143]; the detection of characteristic losses, such as those of prenylated compounds [144]; the classic fragmentation of flavonoids or related compounds for the determination of substituent positions on A or B rings [145]; and the differentiation of isomers [120]. For other studies, the interpretation of the MS-MS spectra often requires the analysis of many related products [146] in order to extract structurally relevant information and establish rules that can be used for structure prediction. It is also worth mentioning that MS-MS is a well-established method for the analysis of peptides and provides unambiguous information on the sequence of amino acids based on precise fragmentation rules [147]. The method is also applicable to naturally occurring plant cyclic peptides after conversion into acyclic forms by partial hydrolysis [148]. For the analysis of fully unknown constituents, MS-MS usually cannot provide enough information to ascertain structure determination, and thus the combination with other online information is mandatory.

In Fig. 1 (inset b), an example of an MS spectrum obtained with ESI (PI) on a TOF instrument is presented for the accurate mass determination of a flavonoid triglyceride (F) (kaempferol 3-O-β-[β-D-rhamnopyranosyl(1→2)-o-glucopyranosyl]-7-O-α-L-rhamnopyanoside) of Arabidopsis thaliana. As shown, the [M+H]^+ ion was recorded with high precision (m/z = 741.2242) with the help of a reference compound analysed during the whole procedure on a separated sprayer (LockSpray) [149]. This exact mass measurement enables the molecular formula determination (C_{32}H_{41}O_{19}) with an accuracy of less than 5 ppm. A cross-library search with this molecular formula and chemotaxonomic information (genus Arabidopsis) in a library of natural products (DNP Chapman and Hall [123]) enabled the dereplication of this compound. Its UV-DAD spectrum (Fig. 1, inset a) confirmed that this compound was a flavonoid derivative (see above).

HPLC-NMR (nuclear magnetic resonance)
Because of the lack of efficient commercial MS-MS databases, the dereplication process often requires additional spectroscopic information to ascertain the identity of known natural products or to partially identify unknown metabolites. In this respect, HPLC-NMR provides a powerful tool that can yield important complementary information or, in some cases, a complete structural assignment of natural products online [119], [124], [150], [151]. HPLC-NMR should ideally enable the complete structural characterisation of any plant metabolite directly in an extract, provided that its corresponding LC peak is clearly resolved. In practice, however, this is not entirely true, as many factors hinder online structure determination [150]. These problems are linked mainly to the inherent low sensitivity of NMR for the detection of microgram or sub-microgram quantities of natural products separated by conventional HPLC and the need for solvent suppression.

Indeed, HPLC-NMR can be regarded as relatively insensitive (Table 1) due to the intrinsic properties of NMR detection. Consequently, the LODs are several orders of magnitude higher than those of HPLC-UV or HPLC-MS. These limits also depend on the type of magnet used (400 – 900 MHz) and the type of flow probe employed (standard HPLC-NMR probes 40 – 120 μL, microcoils 20 nL–1 μL or cryo-flow probes). The mode of operation also strongly affects sensitivity. Indeed, in the on-flow measurements the time of acquisition is limited, and only few transients are recorded when the LC peak crosses the flow cell, while in the stop-flow mode an important number of transients can be acquired, consequently improving the S/N ratio. With a standard HPLC-NMR flow probe (60 μL on a 500-MHz magnet), the detection limits are about 20 μg of NP injected on-column in the on-flow mode, but they can reach the nanogram range when HPLC-NMR is used at-line with sensitive microflow probes [119].

HPLC-NMR, in the on-flow mode, can provide 1H-NMR spectra of the main constituent of a crude extract rapidly without the need for complex automation. This type of information, in addition to other online spectroscopic data (e.g., DAD and MS), may ascertain the structural determination and facilitate the dereplication process. Examples of on-flow HPLC-NMR analyses for crude extract profiling are manifold (e.g., for flavonoids, alkaloids, ter-
protons of hypericins in the region of 14–15 ppm in CD3OH solutions of the same plant was also efficiently performed with a reversed-phase solvent and using microflow capillary HPLC-NMR probes (CapNMR) [154], [155], [156]. These at-line HPLC-NMR analyses provide a good HPLC peak preconcentration and high quality 1D and 2D NMR spectra in fully deuterated solvents without the need for solvent suppression. The work at-line has the disadvantage that more sample handling or automation is needed compared with the on-flow approach. However, these techniques are crucial for the complete de novo structure elucidation of given HPLC peaks, as they can provide $^1$H-$^1$C correlation experiments in the low microgram range [150].

Conclusions

The quest for universal and sensitive HPLC detection with uniform response factors for all analytes is ongoing. Thus far, no ideal solution has been found, and specific detectors need to be chosen according to the type of NP analysed. The lack of a sensitive and universal detection technique suitable for both qualitative and quantitative analysis under a wide range of conditions is one of the reasons why the development of generally applicable methods for HPLC is still challenging. As shown, many LC detectors have been developed over the years, with different advantages and limitations, and a summary of their main features is provided in Table 2. A literature search restricted to journals focused on NP studies revealed that more than 70% of HPLC applications are performed by HPLC-UV and HPLC-DAD, with HPLC-MS applications accounting for about 20% (one-third being related to MS-MS), HPLC-EC for 3%, HPLC-NMR for 2%, and ELSD for 1.5%. The other detection techniques are reported in less than 1% of all HPLC applications. The large majority of analyses are still performed by HPLC-UV, which is widespread in many laboratories. For the detection of non-UV-active constituents, aerosol-based detectors, such as ELSD or CAD, represent a cost-effective alternative to HPLC-MS and will probably increase in popularity. HPLC-MS is a very versatile and powerful tool. It is sensitive and can be considered both a specific and a universal detection method. The technique is still expensive, but the development of benchtop instruments that are easy to operate will continue to increase its use in many areas of NP research. However, quantitative analysis with HPLC-MS is still less robust than with other techniques, as the response depends on the ionisation process and the method suffers from ionisation suppression problems when complex matrices are analysed.

The introduction of sub 2-μm columns for HPLC and the development of UHPLC represent important steps forward for crude extract profiling. The speed of separation provided by these new separation methods challenges the LC detectors, which have to provide faster responses. The strong development of...
“-omic’s” approaches in NP research (metabolomics, genomics, etc.) over the last decade also challenge the analytical methods as they aim to detect and quantify all metabolites in a given organism. For these types of studies, both separate and non-separate methods must be complementary because none of them alone can fulfill all the ideal needs of sensitivity and throughput. For online structural determination, HPLC-DAD-MS represents a key strategic tool, and it is used mainly for dereplication purposes in laboratories involved in drug discovery programs. However, the lack of general HPLC-MS databases restricts the possibility of prediction and automation in the dereplication process. Thus, complementary methods such as HPLC-NMR and derived techniques used at-line (SPE-NMR or CapNMR) are necessary. A topic not discussed in this review is the use of biological assays and integrated lead-finding platforms.

With the increasing requirements for quality control, profiling and fingerprinting, dereplication, and metabolomics, the detection issue in HPLC is still challenging. New detectors still have to be developed to fulfill the needs of sensitivity, linearity, speed, robustness, and universal response requested by the investigations performed currently in natural products research.

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References

6 David F, Vanhoenacker G, Tienpoint B, Francois I, Sandra P. Coupling columns and multidimensional configurations to increase peak capacity in liquid chromatography. LC Gc Europe 2007; 20: 154–8


10 Ozkan SA. LC with electrochemical detection. recent application to pharmaceuticals and biological fluids. Chromatographia 2007; 66: 53–513


42 LeCrosire WR, Modi SJ. Microelectrode applications of pulsed electrochemical detection. Electroanalysis 2003; 15: 1141–52


74 Gamache PH, McCarthy RS, Freeto SM, Asa DJ, Woodcock MJ, Law K et al. HPLC analysis of non-volatile compounds using charged aerosol detection. LC GC Europe 2005; 13: 345 – 9


84 Brewer TM, Castro J, Marcus RK. Particle beam sample introduction into glow discharge plasmas for speciation analysis. Spectrochim Acta Part B At Spectrosc 2006; 61: 134 – 49


96 Wan JB, Yang FQ, Li SP, Wang YT, Cui XM. Chemical characteristics for different parts of Panax notoginseng using pressurized liquid extraction and HPLC-ELSD. J Pharm Biomed Anal 2006; 41: 1596 – 601
107 Xie JS, Ding CG, Ge QH, Zhou Z, Zhi XJ. Simultaneous determination of ginkgolides A, B, C and bilobalide in plasma by LC-MS/MS and its application to the pharmacokinetic study of Ginkgo biloba extract in rats. J Chromatogr B 2008; 864; 87–94
121 Hall C. Dictionary of natural products on CD-ROM: Chapman & Hall; 2008
128 Huber L, George SA. Diode array detection in HPLC. New York: Marcel Dekker; 1993
133 Cobbo-Neto L, Lopes NP. Online identification of chlorogenic acids, sesquiuterpenes lactones, and flavonoids in the Brazilian arnica Lycophora ericoides Mart. (Asteraeaceae) leaves by HPLC-DAD-MS and HPLC-DAD-MS/MS and a validated HPLC-DAD method for their simultaneous analysis. J Agric Food Chem 2008; 56: 1193 – 204

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147 Biemann K. Sequencing of peptides by tandem mass spectrometry and high-energy collision induced dissociation. Methods Enzymol 1990; 193: 455 – 79


