**Tapirira guianensis** is Selectively Cytotoxic, Induces Apoptosis to the Glioblastoma and Decreases Tumor Growth and Angiogenesis *in vivo*

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**Key words**
Glioblastoma, cerrado, **Tapirira guianensis**, Anacardiaceae, apoptosis, angiogenesis

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**ABSTRACT**
Glioblastoma is the most frequent primary malignant brain tumor without effective treatment, which makes this work extremely relevant. The study of the bioactive compounds from medicinal plants plays an important role in the discovery of new drugs. This research investigated the constituents of **Tapirira guianensis** and its antitumor potential (*in vitro* and *in vivo*) in glioblastoma. The *T. guianensis* extracts were characterized by mass spectrometry. The ethyl acetate partition (01ID) and its fractions 01ID-F2 and 01ID-F4 from *T. guianensis* showed potential antitumor treatment evidenced by selective cytotoxicity for GAMG with IC50 14.1 µg/mL, 83.07 µg/mL, 59.27 µg/mL and U251 with IC50 25.92 µg/mL, 37.3 µg/mL and 18.84 µg/mL. Fractions 01ID-F2 and 01ID-F4 were 10 times more selective when compared to TMZ and 01ID for the two evaluated cell lines. *T. guianensis* also reduced matrix metalloproteinases 2–01ID-F2 (21.84%), 01ID-F4 (29.6%) and 9–01ID-F4 (73.42%), ID-F4 (53.84%) activities, and induced apoptosis mainly through the extrinsic pathway. Furthermore, all treatments significantly reduced tumor size (01ID p < 0.01, 01ID-F2 p < 0.01 and 01ID-F4 p < 0.0001) and caused blood vessels to shrink *in vivo*. The present findings highlight that *T. guianensis* exhibits considerable antitumor potential in preclinical studies of glioblastoma. This ability may be related to the phenolic compounds and sesquiterpene derivatives identified in the extracts. This study deserves further *in vivo* research, followed by clinical investigation.

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Introduction

Glioblastoma (GBM) is one of the most hostile malignant neoplasms that affect the central nervous system, classified as grade IV, and originates from astrocytic glial cells or their precursors [1]. The incidence of glioblastoma ranges from 3.19 to 4.17 cases per 100,000 person-years [2]. This tumor manipulates the microenvironment around it, supporting and stimulating angiogenesis, in addition to suppressing the immune system [3].

The current treatment for glioblastoma is surgical resection, followed by radiotherapy and chemotherapy with temozolomide, but these treatments have a low impact on median survival, which remains only 15 months [1–4]. Therefore, the attempt to achieve a cure for glioblastoma is still fruitless. Factors such as the blood vascularized network, the nonselective effect of TMZ, in addition to the blood–brain barrier, can lead to chemotherapy resistance, myelosuppression and difficulty in delivering drugs to tumor cells [5]. The inhibitory effects of natural products on angiogenesis and metastasis are encouraging. These effects also include the induction and acceleration of apoptosis and the production of reactive oxygen species, which have anti-metastasis effects. In addition, these effects may be combined with the reductions in toxicity and side effects [6].

Between 1981 and 2014, the Food and Drug Administration approved more than 1500 drugs, of which 38% were obtained or based on natural products [7]. Regarding antitumor drugs, 35% of these originate directly or indirectly from natural products, of which approximately 25% originate from plants [8]. We previously reported the cytotoxic activity of the ethyl acetate partition of *Tapirira guianensis* and its ability to inhibit tumor migration and invasion in oral cancer cell lines [9]. This plant also inhibited MMP-2 and its ability to inhibit tumor migration and invasion in oral cancer cell lines [9]. Here, we also demonstrate that the fractions 01ID and 01ID-F4, derived from this extract, have a purer constitution and, in addition to their cytotoxicity, were 10 times more selective.

Another characteristic of glioblastoma is its high ability to proliferate and invade adjacent tissues fast [17,18]. Only the ethyl acetate partition inhibited the glioblastoma cells’ migration. These can indicate that the separation process in the fractionation may have lost the compounds with this capacity or that compounds that act synergistically present in the partition have been separated and, thus, lost the activity [19,20]. The 01ID significantly reduced cell migration of GAMG and U251 glioblastoma cells at 24, 48 and 72 hours; p < 0.05, p < 0.01 (Fig. 1a). However, no significant capability to reduce cell migration was observed (Fig. 1b).

Condensation and fragmentation of nuclear material and apoptotic bodies were observed in the cells GAMG and U251 after exposure to the partition and fractions (Fig 2). Exposure to the *T. guianensis* partition and fractions decreased the activity of metalloproteinases 2 and 9. Exposure to 01ID caused an additional decrease in MMP-9 activity by 76.00% ± 11.71 (p = 0.0142) (Fig. 3a,b) regarding the effect on MMP-2 (33.31% ± 6.05 (p = 0.0161)) (Fig. 3e,f). Fractions 01ID-F2 and 01ID-F4 were also more effective in decreasing MMP-9 activity 73.43% ± 0.97 (p < 0.0001) and 53.84% ± 1.55 (p = 0.0004) (Fig. 3c,d), respectively. For MMP-2, the action of fractions 01ID-F2 and 01ID-F4 caused a decrease of 21.84% ± 2.14 (p < 0.0001) and 29.60% ± 2.46 (p < 0.0034), respectively (Fig. 3g,h).

| Table 1 The IC50 values of exposure to 01ID partition, fractions 01ID-F2 and 01ID-F4 of *T. guianensis* and TMZ in GAMG, U251 and NHA cells at 72 hours. |
|-----------------|-----------------|-----------------|-----------------|-----------------|
| **Cell lines**  | 01ID            | 01ID-F2         | 01ID-F4         | **TMZ**         |
| GAMG            | 14.1 ± 1.14     | 13.18 ± 1.12    | 22.34 ± 1.34    | 75.58 ± 1.87   |
| U251            | 25.92 ± 1.41    | 4.83 ± 0.68     | 6.72 ± 0.82     | 167.3 ± 2.22   |
| NHA             | 24.88 ± 1.39    | 130.3 ± 2.11    | 405 ± 2.6       | 48.49 ± 1.69   |

| Table 2 Tumor selectivity index after treatment with partition 01ID and fractions 01ID-F2 and 01ID-F4 and chemotherapy TMZ for 72 hours. |
|-----------------|-----------------|-----------------|-----------------|-----------------|
| **Cell line**   | 01ID            | 01ID-F2         | 01ID-F4         | **TMZ**         |
| GAMG            | 1.76            | 9.89            | 18.13           | 0.64            |
| U251            | 0.96            | 26.95           | 60.28           | 0.29            |

Results and Discussion

Temozolomide (TMZ) is the only effective chemotherapy in glioblastoma treatment [11]. However, the response is limited, with an average survival time of only 16 months, which changed little over the past decade [12]. These reactions can occur because of the low selectivity of TMZ for tumor cells, as demonstrated in another study by our group [13]. So, a desirable resource in the search for new therapies is the selectivity of treatment.

Among the evaluated *Tapirira guianensis* partitions, 01ID exhibited IC50 of 14.1, 25.92 and 24.88 µg/mL for GAMG, U251 and NHA cell lines, respectively. These values were lower than those found for temozolomide (Table 1). Regarding the selectivity index (SI), it was observed that the ethyl acetate partition is as selective as its fractions (Table 2). Fractions 01ID-F2 and 01ID-F4 were 10 times more selective when compared to both TMZ and 01ID for the two cell lines evaluated. The ideal drug should have a relatively high toxic concentration but with a very low active concentration [14]. Regarding cellular characteristics, we observed that cancer cells could migrate and form a tumor mass, which is not observed in normal cell lines, which are differentiated. In addition, the GAMG and U251 lineages have the TP53 and TERT records, and the U251 still has the CDKN2A/B and PTEN records [15,16,42,43)]. Meanwhile, the NHA does not present cells, being of normal cerebral tissue origin [15, 16].

Extracts derived from *T. guianensis* leaves showed high selectivity for tumor cells, making them relevant as possible chemotherapeutic agents for this neoplasm treatment. These results agree with our previous study, in which the ethyl acetate extract of *T. guianensis* decreased the cell viability of head and neck tumor cell lines [9]. Here, we also demonstrate that the fractions 01ID-F2 and 01ID-F4, derived from this extract, have a purer constitution and, in addition to their cytotoxicity, were 10 times more selective.
Fig. 1 Cell migration of GAMG and U251 cell lines after treatment with the 01ID and its fractions of T. guianensis. a Representative images at times 0, 24, 48 and 72 hours with treatment 01ID. b Representative images at times 0, 24, 48 and 72 hours with treatment 01ID-F2 and 01ID-F4. c Graphs of the wound area in the GAMG strain at times 0, 24, 48 and 72 hours with the treatment 01ID. d Graphs of wound area in the U251 strain at times 0, 24, 48 and 72 hours with the treatment 01ID. e Graphs of the wound area in the GAMG strain at times 0, 24, 48 and 72 hours with the treatment 01ID-F2 and 01ID-F4. f Graphs of wound area in the U251 strain at times 0, 24, 48 and 72 hours with the treatment 01ID-F2 and 01ID-F4. Data are represented as mean and standard deviation. Statistical analysis by Tukey's multiple comparisons test. * p < 0.05, ** p < 0.01, *** p < 0.001, n.s.: no significance.
**Fig. 2** Fluorescence morphological assay using AO/PI double staining in GAMG and U251 cell lines after exposure with the 01ID and its fractions. 

- **a** Representative images of control and fractions in the GAMG. 
- **b** Representative images of control and treatments in the U251. 
- **c and d** Graphs of viable and dying cell numbers in the GAMG and U251 tumor lines, respectively. Data are represented as mean and standard deviation. Statistical analysis by t-test, **** p < 0.0001.

**Fig. 3** Zymography after treatment with the 01ID partition of *T. guianensis*. Zymographic images of the action of *T. guianensis* partition on MMP-9 (a) and MMP-2 (e) and graphs representing the percentage of this inhibition of activity in MMP-9 (b) and MMP-2 (f). Zymography images representing the action of fractions (01ID-F2 and 01ID-F4) of *T. guianensis* on MMP-9 (c) and MMP-2 (g) and the graphs representing the percentage of this inhibition in MMP-9 (d) and MMP-2 (h). Data are represented as mean and standard deviation. Statistical analysis by t-test, * p < 0.05, **p < 0.0034, ***p = 0.0004, ****p < 0.0001 compared to untreated control.
The 01ID reduced by 32.81% ± 5.79 (p < 0.0001) the cell adhesion (Fig. 4). There was no reduction in adherence after treatment with fraction 01ID-F2. However, fraction 01ID-F4 showed a significant decrease in cell adhesion of 20.38 ± 6.98 cell adhesion (p < 0.0013) (Fig. 4).

There was a significant increase in proteins related to the apoptotic death pathway when cells were treated with the compounds. An increase in TNF-R1 was observed in 52.5% (p < 0.001), 88.19% (p < 0.01) and 91.86% (p < 0.001) when cells were treated with fractions 01ID, 01ID-F2 and 01ID-F4, respectively. FAAD expression increased significantly by 37.93% (p < 0.001), 57.96% (p < 0.05) and 51.20% (p < 0.01) for treatments 01ID, 01ID-F2 and 01ID-F4, respectively. We also observed a significant increase in caspases 8, 7 and 3. Caspase 8 increased 72.41% (p < 0.001), 71.92% (p < 0.001) and 87.74% (p < 0.001) for treatments 01ID, 01ID-F2 and 01ID-F4, respectively. Caspase 7 also showed significant increases of 45.14% (p < 0.01), 71.92% (p < 0.01) and 77.74% (p < 0.001) for treatments 01ID, 01ID-F2 and 01ID-F4, respectively. Finally, there was an increase in caspase 3 of 72.91% (p < 0.01), 92.31% (p < 0.01) and 102.29% (p < 0.0001) (Fig. 5).

The increased expression of TNF-R1, FADD, and caspases 8, 7 and 3 found in our study is related to cell death by extrinsic apoptosis [21]. This action can be attributed to compounds from the class of flavonoids found in T. guianensis samples, such as luteolin 7-O-glycoside and quercetin, previously associated with the induction of cell death by this cell death pathway [22]. Furthermore, studies in prostate and breast tumors showing that catechins, also present in T. guianensis, can induce cell death through this pathway [23, 24] support our results.

Cell proliferation and invasion are related to the activity of matrix metalloproteinases (MMPs) as they degrade extracellular matrix components, which helps in tumor progression and angiogenesis [15, 16]. The expression of metalloproteinases plays a vital role in the invasion of cancer cells in normal tissue, indirectly reducing the effectiveness of chemotherapy [25]. Therefore, inhibiting the expression of MMPs may be a remarkable strategy to control these processes [25]. In this study, we observed a reduc-
tion in the activity of MMPs 2 and 9, which may be related to the presence of quercetin in fractions 01ID and 01ID-F4. Studies have shown the potential of this flavonoid to inhibit MMP-2 in GL-15 glioblastoma cells and to inhibit MMP-2 and MMP-9 expressions in U251 [26, 27]. However, this inhibitory effect may also be related to the compound genistein, which only inhibits MMP-2 [28]. Another compound present in the extracts is catechin, which is related to the potential for inhibiting metalloproteinases (MMP-2 and 9) in several types of tumors, such as digestive, prostate and lung tumors [29].

Regarding the tumor perimeter reduction, 01ID, 01ID-F2 and 01ID-F4 were statistically significant (p < 0.01, p < 0.01 and p < 0.0001, respectively) (Fig. 6). Moreover, CAMs were histologically analyzed to quantify the number of blood vessels in which significant reductions (p < 0.0017) were observed after treatment with the ethyl acetate partition extract and with the 01IDF-F2 and 01ID-F4 fractions (Fig. 7). Furthermore, after exposure to T. guianensis, there was no effect on blood vessel lysis and bleeding (Fig. 8).

One of the hallmarks of cancer is angiogenesis, which plays an essential role in tumor growth, invasion and metastasis. The chicken chorioallantoic membrane (CAM) model offers excellent advantages for both the macroscopic and microscopic study of tumor development to angiogenesis [30]. Because it is partially immunodeficient, the CAM can receive xenotransplants from other species without interfering with an immune reaction. The natural vascular network is another advantage, as it provides an interface that allows human tumors survival, growth and vascularization [31].
The reduction in the perimeter of the tumor and angiogenesis found in our study may be related to the compounds present in the extracts, such as gallic acid, which has already been reported as an angiogenesis inhibitor in cervical tumors [32]. Catechins can also be related, as they were responsible for inhibiting tumorigenesis in several types of tumors, including glioblastoma, by inhibiting the apoptosis pathway and inhibiting proliferation, cell growth and angiogenesis [21-24, 28-34]. These substances, including epicatechin present in the 01ID-F4 fraction, may explain the best inhibition of tumor growth and corroborate the findings of Wang et al. (2019), who indicated that (−)-Epigallocatechin 3-O-gallate is a potent inhibitor of angiogenesis in an esophageal tumor [24].

The ethyl acetate extract (01ID) and the fractions 01ID-F2 and 01ID-F4 were analyzed by UPLC-QTOF/MS. Fifteen compounds were tentatively identified in 01ID: mainly phenolic compounds (Table 3). In the 01ID-F2 fraction, 10 compounds were tentatively identified: mostly sesquiterpenes or fatty acids derivatives (Table 4). In the 01ID-F4 fraction, 12 compounds were tentatively identified: mainly phenolic compounds (Table 5).

Among the various routes of chemotherapy administration, the intravenous way is one of the most used in its administration. However, leakage of the venous network may occur, causing subcutaneous toxicity and causing damage to adjacent tissues, ulcerations and necrosis [35]. The HET-CAM, a variation of the CAM that was initially used to assess the risk of irritation of the skin and mucous membranes caused by exposure to new substances, also allows the measurement of the risk of irritancy of the infusion site, reducing the use of tests in mammals [36]. Thus, we also verified that the extracts of *Tapirira guianensis*, 01ID, 01ID-F2 and 01ID-F4, did not cause irritability, hemorrhage or blood clotting, suggesting that these samples can be administered parenterally [37].

The ethyl acetate partition from *T. guianensis* and its fractions are cytotoxic to glioblastoma tumor cells, the latter being more selective than partition. Also, they can induce cell death by extrinsic apoptotic pathways. Furthermore, partition and fractions effectively inhibited tumor growth and angiogenesis in vivo. This capacity may be due to compounds of the extracts such as the flavonoid class, such as the compounds luteolin 7-O-glucoside and quercetin or catechins. Ultimately, these findings may lead to new therapeutic strategies discovery against glioblastoma.

**Materials and Methods**

**Obtaining extracts partition and fractions**

*Tapirira guianensis* leaves were collected at latitude S18°58′08″ and longitude W 49°27′54″ by Allisson Rodrigues de Rezende, and the exsiccate was deposited in the Depositary Herbarium of the Federal University of Minas Gerais (143407 BHCB), SISGEN Nº R688CA8. The leaves were dehydrated, pulverized and kept in a

![Fig. 8](image-url)
cool, dry, dark place. The powder was macerated in an alcoholic solution (70%) at a ratio of 1:5 and lyophilized to produce the crude extract (CE). The CE was suspended in an alcohol solution (70%) and fractionated by liquid–liquid extraction using hexane, chloroform and ethyl acetate in increasing order of their polarity. The ethyl acetate 01ID sample extract was collected, frozen and lyophilized. The fractions were obtained from the ethyl acetate partition, using columns (4 cm in diameter) for the fractionation.

Silica gel 60 (230–400 Mesh–Sorbine Adsorbents Tecnologia Ltda.) was solubilized in the hexane/ethyl acetate (5:1) eluent and inserted into the column to a height of 15 cm with a silica pump vacuum (Prismatec). The fractions were collected every 10 mL. Thin-layer chromatography (CCD) evaluated fractions, and those with similar profiles were pooled and left six fractions at the end. Finally, the solvent of the fractions was removed using a rotary evaporator and desiccator, and the masses were measured in analytical balance (Mars).

From 6 g of crude extract, 0.633 g of ethyl acetate partition yield were obtained. From the 3.004 g mass of the partition, 0.045 g of fraction 01ID-F2 and 0.2026 g of fraction 01ID-F4 were obtained.

**Cell culture**

The commercial glioblastoma cell line used was GAMG (ACC 242) obtained from the German Collection of Microorganisms and Cell Cultures (DSMZ) and U251 (European Collection of Authenticated Cell Cultures (ECACC) Catalogue No. 09063001). In addition to the normal commercial NHA, Normal Human Astrocyte – CC-

| Table 3 Qualitative analysis by UPLC-MS of 01ID fraction. Hi – Hibiscetin; Myr – Myricetin, Querc – Quercetin, (Epi)cat – (Epi)Catechin, Kaemp – Kaempferol, Hex – Hexosides, Pent-Pentosides |
|---|---|---|---|---|---|
| r (min) | Ions m/z | Formula | Error (ppm) | Tentative Identification | Detection Mode |
| 0.697 | 191.0564 | C_7H_12O_6 | 4.2 | Quinic Acid | – |
| 1.812 | 169.0143 | C_7H_6O_5 | 3.6 | Gallic Acid | – |
| 2.54 | 495.0781 | C_{21}H_{20}O_{14} | 2.4 | Hi-hex | – |
| 2.557 | 495.0783 | C_{21}H_{20}O_{14} | 2.8 | Hi-hex | – |
| 3.026 | 479.0836 | C_{21}H_{20}O_{13} | 2.1 | Myr-hex | – |
| 615.0995 | C_{29}H_{24}O_{16} | 1.5 | Querc-galloyl-hex | |
| 729.1459 | C_{32}H_{30}O_{16} | 1.2 | (Epi)cat-gallate dimer | |
| 3.12 | 441.0837 | C_{22}H_{18}O_{10} | 4.8 | (Epi)cat-gallate | – |
| 463.0894 | C_{21}H_{20}O_{12} | 3.7 | Querc-hex | |
| 599.1056 | C_{28}H_{24}O_{15} | 3.2 | Kaemp-galloyl-hex | |
| 3.27 | 433.0783 | C_{20}H_{18}O_{11} | 2.8 | Querc-pent | – |
| 441.0832 | C_{22}H_{18}O_{10} | 3.6 | (Epi)cat-gallate | – |
| 463.0892 | C_{21}H_{20}O_{12} | 3.2 | Querc-hex | |
| 599.1056 | C_{29}H_{24}O_{15} | 3.2 | Kaemp-galloyl-hex | |
| 3.30 | 433.0781 | C_{22}H_{18}O_{11} | 2.3 | Querc-pent | – |
| 447.0936 | C_{21}H_{20}O_{11} | 2.0 | Kaemp-hex | |
| 3.33 | 447.0938 | C_{21}H_{20}O_{11} | 2.5 | Kaemp-hex | – |
| 3.026 | 617.1131 | C_{29}H_{24}O_{16} | – 1.9 | Querc-galloyl-hex + | |
| 3.125 | 303.0498 | C_{15}H_{10}O_{7} | – 2.3 | Quercetin + | |
| 465.1025 | C_{21}H_{20}O_{12} | – 1.7 | Querc-hex | – |
| 601.118 | C_{29}H_{24}O_{15} | – 2.2 | Kaemp-galloyl-hex | |
| 3.241 | 303.0493 | C_{15}H_{10}O_{7} | – 4.0 | Quercetin + | |
| 435.0914 | C_{20}H_{18}O_{11} | – 3.0 | Querc-pent | – |
| 601.1174 | C_{29}H_{24}O_{15} | – 3.2 | Kaemp-galloyl-hex | |
| 3.302 | 303.0495 | C_{15}H_{10}O_{2} | – 3.3 | Quercetin + | |
| 4.538 | 318.2996 | C_{19}H_{20}NO_{3} | – 2.2 | 4-hydroxysphinganine + | |
| 4.968 | 302.3047 | C_{19}H_{20}NO_{2} | – 2.3 | Sphinganine + | |
| 346.3309 | C_{20}H_{24}NO_{3} | – 2.0 | Aminoeicosanetriol | |
2565, Lonza). All strains were cultured in DMEM (Dulbecco’s modified Eagle medium) with 1% penicillin/streptomycin (P/S) antibiotics and supplemented with 10% FBS (fetal bovine serum). They were kept in an incubator of CO2 at 37°C.

In vitro experiments using the cells were performed in triplicate.

MTT cytotoxicity and selective index

Cells were seeded in 96-well plates at a density of 5 × 103 cells per well and incubated overnight in a CO2 (5%) at 37°C for plate adhesion. Treatments were prepared in DMSO (1%) and DMEM supplemented with 0.5% FBS. After 24 hours, cells were treated with extracts at concentrations of 0, 5, 15, 30 and 40 µg/mL and fractions of 0, 5, 10, 15, 20, 30 and 35 µg/mL concentrations for 24 and 72 hours. The cell viability was quantified using MTT colorimetric reagent (3- [4,5-Dimethylthiazol-2-yl] - 2,5-diphenyltetrazolium bromide; Invitrogen – M6494)), and absorbance was measured using an Elisa reader, at a wavelength of 570 nm, after incubating the cells with MTT for 3 hours.

Table 4 Qualitative analysis by UPLC-MS of 01ID-F2 fraction.

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<thead>
<tr>
<th>rt (min)</th>
<th>Main Ions m/z</th>
<th>Formula</th>
<th>Error (ppm)</th>
<th>Tentative Identification</th>
<th>Detection Mode</th>
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<td>3.512</td>
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Table 5 Qualitative analysis by UPLC-MS of 01ID-F4 fraction.

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<th>rt (min)</th>
<th>Ions m/z</th>
<th>Formula</th>
<th>Error (ppm)</th>
<th>Tentative Identification</th>
<th>Detection Mode</th>
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<td>495.0781</td>
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<td>2.8</td>
<td>Syringic acid</td>
<td>–</td>
</tr>
<tr>
<td></td>
<td>417.0831</td>
<td>C₂₀H₁₈O₁₀</td>
<td>2.2</td>
<td>Kaemp-pent</td>
<td>–</td>
</tr>
<tr>
<td>3.818</td>
<td>301.0359</td>
<td>C₁₃H₁₀O₂</td>
<td>3.7</td>
<td>Quercetin</td>
<td>–</td>
</tr>
<tr>
<td>4.127</td>
<td>285.0410</td>
<td>C₁₃H₁₀O₆</td>
<td>3.9</td>
<td>Kaempferol</td>
<td>–</td>
</tr>
<tr>
<td>3.026</td>
<td>617.1129</td>
<td>C₂₃H₂₆O₁₆</td>
<td>−2.3</td>
<td>Querc-galloyl-hex</td>
<td>+</td>
</tr>
<tr>
<td>3.241</td>
<td>190.0596</td>
<td>C₉H₁₀O₅</td>
<td>−2.5</td>
<td>Syringic acid</td>
<td>+</td>
</tr>
<tr>
<td>3.302</td>
<td>303.0495</td>
<td>C₁₃H₁₀O₇</td>
<td>−3.3</td>
<td>Quercetin</td>
<td>+</td>
</tr>
<tr>
<td>4.129</td>
<td>287.0546</td>
<td>C₁₃H₁₀O₆</td>
<td>−3.5</td>
<td>Kaempferol</td>
<td>+</td>
</tr>
</tbody>
</table>

Hi – Hibiscetin; Myr – Myricetin, Querc – Quercetin, Kaemp – Kaempferol, Hex – Hexosides, Pent-Pentosides
From the value obtained from the IC50, the selectivity index (SI) was calculated. The calculation was performed from the formula $SI = \frac{IC50}{n}$ non-tumor cells/IC50 tumor cells [36].

**Wound healing cell migration**

The wound healing assay evaluated the inhibitory potential of cell mobility of the extracts. The GAMG and U251 tumor cells were seeded in DMEM supplemented with 10% FBS in 24-well plates at the density of $5 \times 10^5$ cells/well. The cells were incubated overnight at CO2 and 37°C. Thus, two wounds were made in each well oven at 37 °C. Afterward, excess liquid was removed and 100 µL diluted with 10 µg/mL BSA (1 : 10 PBS1X) and incubated in a CO2. The wells of the 96-well plate were coated with 100 µL of Matrigel.

**Zymography**

The proteolytic enzyme activity of pro-MMP-2 was measured by gelatin zymography. The gelatinolytic activity of MMPs was investigated by zymographic. MMP-2 (10 ng/µL – REF: ab81550) and MMP-9 (10 ng/µL – REF, Ab82955) samples and treatments (IC50) were used [10]. The samples were subjected to electrophoresis through a 7% Zymogram Ready Gel (Bio-Rad Laboratories). After electrophoresis, the gel was incubated twice in 2.5% TritonX-100 for 60 min at room temperature and, then, incubated at 37°C for 18 h in an activation buffer (10 mM Tris-HCl buffer (pH 8.0) containing 5 mM (Tris-Cl)2). Gels were stained (0.25% Comassie blue G-250, 30% ethanol, 10% acetic acid) for 1 h and destained (30% ethanol, 10% acetic acid) for 2 h. Results of the mean ± standard deviation of inhibition were converted into percentages and normalized in the presence of controls using Excel 2010 software.

**Cell adhesion assay**

The wells of the 96-well plate were coated with 100 µL of Matrigel diluted with 10 µg/µL BSA (1 : 10 PBS1X) and incubated in a CO2 oven at 37°C. Afterward, excess liquid was removed and 100 µL BSA solution (0.1 %) was added to each well of the plate and again incubated for 2 hours under the same conditions. Finally, the solution was removed, the wells were washed with PBS, and GAMG and U251 (6 × 10^4 cells/well) were seeded in DMEM (FBS free) with the treatments IC50. The plate with the cells was incubated for 2 hours in CO2 at 37°C and, then, fixed with 10% trichloracetic acid (1 hour). Finally, they were stained with violet crystal, and 100 µL of Tris-5 mM was added to the wells for a spectrophotometer reading (540 nm). Absorbance values were converted to percentage mean ± standard deviation of cell viability and normalized in the presence of the DMSO vehicle using Excel 2010 software [39].

**Western blotting**

Cells were cultured (7 × 10^5 cells/well) in DMEM (10% FBS and 1% P/S) for 24 hours. After that, they were replaced by DMEM (0.5% FBS and 1% P/S) for 24 hours. Then, the cells were treated with the IC50 of the compound for 24 hours. The harvested cells were prepared using lysis buffer (NaF, Na3Vo3, PiroNa, Leu, APRO, DTT, EDTA and PMSF) at 4 ° C and were collected by centrifugation. Concentrations of proteins were determined by the Bradford assay (Bio-Rad, Hercules, CA) [40]. Identical amounts of proteins (20 µg) were separated by SDS-PAGE (sodium dodecyl sulfate-polyacrylamide gel electrophoresis) (concentration gels 10% and another 15%). Precision Plus Protein molecular weight (PM) standard (Bio-rad) was used as PM control. The proteins were transferred to nitrocellulose membranes (Amersham Biosciences) using the semi dry transfer equipment (Amersham) for 4 hours at 140 V. The membrane was blocked using 5% non-fat dry milk diluted in TBST (Tris-buffered saline and Tween 20) for 1 h at room temperature and then incubated at 4°C overnight with the specific primary antibodies (Table 1 S, Supplementary material). Subsequently, membranes were incubated with their respective peroxidase-conjugated secondary antibodies (anti-mouse or anti-rabbit) and were developed with enhanced chemiluminescence solution (0.1 M Tris (pH 8.5), 0.25 M luminol (Sigma-Aldrich) and 0.1 M coumaric acid (Sigma-Aldrich)). The protein expression was determined by densitometric analysis and normalized to the corresponding loading controls. Densitometry quantification was performed by ImageJ (NIH) software [41].

**Chicken chorioallantoic membrane assay**

CAM assay was used to assess in vivo tumor proliferation and angiogenesis. Fertilized chicken eggs (Gallus gallus) were incubated for three days at 37°C and 70% humidity. CAM assay was used to assess in vivo tumor proliferation and angiogenesis. Fertilized chicken eggs (Gallus gallus) were incubated for three days at 37°C and 70% humidity. Then, an opening of 1 mm in diameter and then incubated at 4°C overnight with the specific primary antibodies (Table 1 S, Supplementary material). Subsequently, membranes were incubated with their respective peroxidase-conjugated secondary antibodies (anti-mouse or anti-rabbit) and were developed with enhanced chemiluminescence solution (0.1 M Tris (pH 8.5), 0.25 M luminol (Sigma-Aldrich) and 0.1 M coumaric acid (Sigma-Aldrich)). The protein expression was determined by densitometric analysis and normalized to the corresponding loading controls. Densitometry quantification was performed by ImageJ (NIH) software [41].

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removed and histologically analyzed by staining hematoxylin and
eosin. The images were obtained at 200× magnification and the
area of the tumor perimeter and quantified blood vessels with
ZEN software. The Ethics Committee of the Federal University
of São João Del Rei provided ethical approval under protocol number
039/2017 (December 11th, 2017). All applicable international,
national and institutional guidelines for the care and use of ani-
males were followed.

Hen’s egg test – chorioallantoic membrane (het-cam)
test method
The eggs were incubated for 10 days in a digital automatic
brooder at 37°C and 60% humidity. After this incubation period,
the shell and its membrane around the inner tube was removed,
exposing the chorioallantoic membrane. Then, treatments (300 µL)
were applied. The negative control and positive control
used were 0.9% NaCl and 1% SDS, respectively. Images were ob-
tained at times 0, 20, 120 and 300 seconds using the MOTIC ste-
reomicroscope (Moticam 580–5.0MP). The chorioallantoic mem-
brane was examined and the observed physiological reactions
were graded according to their time of onset [42].

Characterization by mass spectrometry (esi-ms)
and uhplc-esi-qtof
The ultra-performance liquid chromatography was performed using
the UHPLC System (Agilent Technologies). An ACQUITY BEH
C18 (130 Å, 1.7 µm, 2.1 × 0.4 id × 100 mm) column (Waters) at
30°C was employed to perform the separation. The ethyl acetate
fraction was diluted in 200 mL of ethanol 70% (v/v) and addition-
ally diluted 1:10 with ETOH 70%. The fractions 01ID-F2 and
01ID-F4 were diluted in 200 mL of ethanol 70% (v/v), and only
fraction 01ID-F4 was additionally diluted 1:10 with ETOH 70%.
The injection volume was 1 µL. The mobile phases consisted of
solvent A (0.1% formic acid in water) and solvent B (0.1% formic
acid in acetonitrile), and the gradient elution conditions were 0
min, 1% B; 0.75 min, 1% B; 6.0/min, 1–99% B; 8.0–9.0 min, 99% B;
9.0–10.0 min, 1% B, with a flow rate of 0.4 mL/min and gradient with
linear changes (min/% B): 0/1, 0.75/1, 6.0/99, 8/99, 9/1, 1. The mass spectrometer was oper-
ated using an electrospray ionization source in positive and nega-
tive ion modes (one mode for run). The MS parameters were set as
capillary voltage 3.00 kV, nozzle voltage 1.00 kV, drying gas N2
temperature 350°C, flow rate 10 L/min, nebulizer pressure
35 psi, sheath gas temperature 300°C, sheath gas flow 10 L/min and
cssan range 100–1700 at a scan rate of 3.00 spectra/s.

Statistical analysis
The results obtained were analyzed using GraphPad Prisms soft-
ware version 5.01 (La Jolla). The significance in all statistical anal-
yses was considered as p < 0.05. Data are represented as the
mean ± SD and values with different superscripts indicating sta-
tistically significant differences (**p < 0.01; ***p < 0.001; *p < 0.05).
Nonlinear regression was used to calculate the IC50.

Supporting Information
The effects used in the Western blotting assay and their concen-
trations are described in the supplemental material.

Contributors’ Statement
RIMAR: conceived and designed the analysis, reviewed and edited
the manuscript and acquired funding; AGSO: data collection, per-
formed the experiments; analysis and interpretation of the data,
drafting the manuscript; M. A. R., L. S. A., A. T. M. C., P. S.: data col-
lection, analysis and interpretation of the data. R. C. R.C; H. B.S.;
R. G. T.; P. S.; E. W.; B. K.; R. M. R.: critical revision of the manuscript
and edited the manuscript, also contributed to data analysis tools.

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made this study possible.

Conflict of Interest
The authors declare that they have no conflicts of interest.

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