Ethnoveterinary Uses of Certain Yemeni Plants: A Review of the Scientific Evidence

ABSTRACT
Livestock is an important and integral component of agriculture production in Yemen and contributes 28% of the total agricultural production income. Research in the field of Yemeni ethnoveterinary medicine is limited to a few studies. Therefore, our work aims to substantiate scientifically the ethnoveterinary use of some documented plant species based on a literature review of their bioactivities and toxicological properties. Searching the scientific literature has revealed various pharmacological activities that may support the claimed healing activities of 11 out of 14 plant species for some of their ethnoveterinary utilization. This comprises the use of Aloe spp. latex for constipation, worms, boils, and wounds; Boswellia sacra underbark for wounds and its oleo-gum resin for mastitis; Soqotraen Boswellia species as an insect repellent; Cissus rotundifolia for stomach pain; Cyphostemma digitatum as an appetite stimulant; Psiaida punctulata for bone fracture; Pulicaria undulata as an insect repellent; combinations of Aristolochia bracteolate with Sorghum bicolor grains for bloating; Rumex nervosus and salt for eye pimples; and Trigonella foenum-graecum seeds with Hordeum vulgare grains for constipation. Some plants were found to demonstrate various toxic effects in in vivo and in vitro experimental studies. The local administration of Calotropis procera latex was also reported to induce an intense inflammatory response. It can be concluded that our work has provided valuable scientific information on the biological and toxic activities of some Yemeni ethnoveterinary remedies that could be utilized for the benefit of farmers to ration the use of these remedies and avoiding their toxicity.

Introduction
Ethnoveterinary medicine is a complex system of beliefs, skills, knowledge, and practices concerning animal health care. In addition to the use of plants to treat animal diseases, the practice of ethnoveterinary medicine includes the use of diagnostic procedures, animal husbandry practices, and surgical methods [1]. Livestock is an important and integral component of agriculture production in Yemen. Animal production contributes 28% of the total agricultural production income. The most important livestock types in Yemen are cows, sheep, goats, donkeys, camels, horses, and chickens. Caring for livestock is a matter of constant concern for the rural population, whereby 95% of the labor force in rural areas are women who are primarily responsible for rearing livestock and possess the knowledge specific to treating sick animals [2]. Some plants used to control animal diseases in some areas of Yemen have been reported in some studies [2–4]. However, ethnoveterinary medicine is still poorly studied, and to the best of our knowledge, no research has yet been done to assess the biological and toxicological activities of the plants used for ethnoveterinary purposes in Yemen. Therefore, our work aimed to confirm the ethnoveterinary uses of 14 documented plant species in different areas of Sana’a, Al Mahwit, Taiz, Ibb, and Dhale governorates [2–3], as well as in the drier, more remote areas of...
Soqotra and South Arabian mainland of Yemen [4], based on their uses and the bioactivities associated with their uses as reported in the literature. Reported evaluation on the safety of these plants was also crucial to raise the farmers’ awareness of toxic plants that may harm their livestock and lead to harmful economic consequences.

Materials and Methods

Scientific information was retrieved from the electronic databases of Leiden University Library, including ScienceDirect, Google Scholar, PubMed, Scopus, and published e-books. Keywords used for searching of plants used for ethnoveterinary medicine in Yemen were “ethnoveterinary medicine”, “ethnoveterinary plants”, “animal health problems”, “livestock treatment”, and “animal diseases” in combination with “Yemen”.

Search terms used for the pharmacological activities associated with the ethnoveterinary uses as well as for the toxicological properties of the Yemeni ethnoveterinary plants found in the scientific literature were the scientific name of the plant, plant extract, or plant constituents in conjunction with pharmacological and toxicological activities such as “laxative”, “anthelmintic”, “antimicrobial”, “anti-inflammatory”, “antioxidant”, “wound healing”, “analgesic”, “antiulcer”, “insecticidal”, “side effects”, “adverse effects”, and “toxicity”. The collected information from the scientific papers were the Latin names of the plants used, names of plant families, plant parts used, uses including methods of administration, pharmacological activities of plant extracts and chemical constituents related to their uses, and the side effects and/or toxicological properties.

Results

Database searches revealed that 14 plant species are used in different areas of Sana’a, Al Mahwit, Taiz, Ibb, and Dhale governorates, and in the drier, more remote areas of Soqotra and South Arabian mainland of Yemen for the treatment of a variety of animal diseases [2–4] (Supplementary material, Table 1). Several pharmacological activities that may lend some scientific support for the ethnoveterinary uses were found for 11 out of 14 plants utilized in Yemeni ethnoveterinary medicine. Three plants (Aloe species latex, Aristolochia bracteolate whole plant, Trigonella foenum-graecum seeds) were reported to possess serious toxicological activities (Table 1).

No data were found that could help explain and support the claimed healing effects of some plants for treating some diseases, such as Aloe spp. latex for colic, as insecticidal, and for shivering attacks and distended belly; Aloe spp. leaves as an animal repellent; Boswellia sacra under bark for generalized edema; Soqotraen Boswellia species as an animal repellent; Calotropis procera latex for swellings; Euphorbia arbuscula latex as insecticidal; and Comophocarpus fruticosus for heatstroke. In addition, these plants or related species were found to exhibit adverse effects. Table 2 illustrates the ethnoveterinary uses and adverse effects of these plants (or related species) used for animal health problems.

Discussion

Traditional use of herbal remedies to treat animals by livestock owners is still prevalent in Yemen due to poverty, the availability of medicinal and aromatic plants, and better accessibility and low cost of herbal drugs, especially when compared to both inaccessible and expensive conventional drugs. However, few studies have reported on the ethnoveterinary use of some plants in Yemen [2–4]. Our work aimed to go a step further by searching for scientific confirmation of the ethnoveterinary use of the 14 Yemeni plants documented in these studies by reviewing their bioactivities and toxicological properties. The results of researching scientific literature have revealed various in vivo and in vitro experimental studies demonstrating different pharmacological activities that may support some of the claimed healing activities of 11 herbal remedies (Table 1).

The use of Aloe latex as a laxative by Yemeni livestock owners and elsewhere [5] is strongly supported by scientific evidence on its laxative properties, based on the well-established cathartic properties of anthraquinone glycosides such as aloin and aloe-emodin found in aloe latex [88]. The laxative activity of the Aloe vera latex is due to a common metabolite, aloe-emodin-9-anthrone, as well as aloin and other metabolites in the colon, where they produce their cathartic effects by multiple mechanisms leading to an increase in intestinal peristalsis, water, electrolyte content, and mucous secretion [6]. The laxative activity of fenugreek seeds and barley grains, on the other hand, is based on the well-known fecal bulking effect of dietary fibers, especially insoluble fibers contained in fenugreek seeds [78] and barley grains [79]. The use of Aloe latex to treat worms could be justified by the in vitro anthelmintic activity of its constituents (aloin and aloe-emodin, Table 1).

Worldwide, disease caused by gastrointestinal strongyles (GIS) is considered one of the most critical health constraints affecting productivity in small ruminants. GIS may limit sheep production by causing retarded growth, weight loss, reduced food consumption, lower milk production, impaired fertility, and even mortality in heavy parasite burden [8]. Aloin and aloe-emodin effectively targeted the larval developmental stages of gastrointestinal strongyle larvae. When used at the same concentration, aloe-emodin especially showed the same efficacy as the reference drug thiabendazole (0.1%, Table 1).

A recent publication [112] confirmed via its in vitro and in vivo results the anthelmintic action of A. vera extract in sheep. The antimicrobial, anti-inflammatory, and antioxidant activities play an essential role in wound healing by reducing the microbial load, decreasing prolonged and chronic inflammation, and protecting vital tissue of the wound bed from free radicals’ destructive effect, respectively [89]. Consequently, these activities produced by Aloe species latex and its anthraquinone constituents (aloemodin and aloin) and by B. sacra under bark and its content of oleo-gum resin [90] (with its constituents, α-pinen [42,91] and boswellic acids [90,92]) alongside the wound healing activity of aloe-emodin and analgesic effect of B. sacra oleo-gum resin and boswellic acids, could substantiate their use for wound healing activity (Table 1).
**Table 1** Uses, pharmacological activities of extracts, chemical constituents, and toxicological properties of some plants utilized in ethnoveterinary medicine in Yemen.

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1,8 dihydroxy-anthraquinone glycosides (Aloin)
- Laxation (1 of intestinal peristalsis, water, electrolyte content, and mucous secretion in the colon) [5, 6]
**Anthelmintic effect**
Aloin
- A dose-dependent paralysis effect, which ultimately progressed to the death of roundworms (Ascaridia galli) and earthworms (Porpetto phostoma). The effect is comparable to standard piperazine [7].
Aloin and Aloe-emodin
(The in vitro anthelmintic activity on sheep gastrointestinal strongyle eggs and larvae)
- 0.5% aloin gave highly significant (p < 0.01) inhibition of egg hatch (39.7%), compared to the untreated controls (13.4%), followed by 0.1% aloe-emodin (16.6%) but these values were significantly lower than that observed in the reference drug 0.1% thiabendazole (TBZ) (94.8%) (p < 0.01) controls.
- Both 0.1% aloe-emodin (97.7%) and 0.5% aloin (96.9%) almost completely prevented the larval development from L1 to L3, showing no significant differences (p < 0.01) when compared to 0.1% TBZ control (100%).
- The larval mortality observed by 0.5% aloin (65.4%) was significantly higher (p < 0.01) than that observed in both the untreated controls (4.2%) and in 0.1% TBZ treated controls (27.7%) [8].
| Boils, suppurating abscess, large fresh laceration, and infected bites of ticks, lice, or flies by local application of fresh latex [4]. | **Antimicrobial activity**
Antimicrobial activity was assessed with broth tube dilution, MBC assay, agar diffusion method, microtiter assay using a metabolic color indicator Alamar blue. Anti biofilm activity screened with crystal-violet screening assay for biofilm inhibitors
**Latex:** active against
- Staphylococcus aureus 209, Streptococcus pyogenes, Corynebacterium xerosis, and Salmonella paratyphi [9].
- Escherichia coli, Staphylococcus epidermidis, Salmonella typhimurium, S. aureus, Bacillus subtilis, Enterococcus faecalis, Proteus vulgaris, and Pseudomonas aeruginosa [10].
**Aloe-emodin:** active against
- B. subtilis, S. aureus, enveloped viruses and plant-pathogenic fungi [5].
- Four strains of methicillin-resistant S. aureus (MRSA) and a strain of methicillin-sensitive S. aureus (with MIC = 2 µg/ml) [12].
- Bacillus cereus, Bacillus licheniformis, and S. epidermidis [13].
- S. aureus biofilm development in a dose-dependent manner [14].
- Candida albicans (MIC = 50 µg/ml) and Trichophyton mentagrophytes (MIC = 50 µg/ml) comparing to ketoconazole (MIC = 0.1 and 2.5 µg/ml, respectively) [15].
- Herpes simplex virus type 1 and type 2, varicella-zoster virus, pseudorabies virus, and influenza virus [16].
**Aloin**
- Effective against B. cereus, B. licheniformis, and S. epidermidis [13].
- Dose-dependent noncompetitive inhibitory effects on Clostridium histolyticum metalloproteinases and collagenases [6].
- Antiviral but not virucidal activity [6].
- Effective against S. aureus, B. subtilis, E. coli, Klebsiella oxytoca, P. aeruginosa, and C. albicans (MIC 6, 5, 7, 8, 12, and 14 µg/ml, respectively) [7].
- Classified as the American Herbal Products Association (AHPA) classes 2b (not to be used during pregnancy), 2c (not to be used while nursing), and 2 d (other specific use restrictions such as not to be used in persons with any inflammatory condition of the intestines) and contraindicated in children under 12 years of age [22–23]. It is not known if similar side effects will occur in animals.
- Prolonged use is associated with watery diarrhea leading to electrolyte imbalance, and the increased loss of potassium can lead to hypokalemia [6].
**Genotoxicity and Mutagenicity**
- A high proportion of anthraquinones including danthron (1,8 dihydroxy-anthraquinone) (synonym: chrysazin) and aloe-emodin were found mutagenic in several strains of Salmonella typhimurium, for example, TA1537, TA1538, TA102, and TA98 [24].
- The genotoxicity and mutagenicity induced by danthron, emodin, and aloe-emodin were due to the inhibition of the catalytic activity of topoisomerase II (Topo II), with danthron being the most potent [24].
**Carcinogenicity**
- The 1,8 dihydroxy-anthraquinones (danthron, aloe-emodin, chrysophanol, and rhein) were found to cause a 2–3-fold increase of DNA synthesis of primary rat hepatocytes and danthron, rhein, and chrysophanol to enhance the transformation of C3H/M2 mouse fibroblasts initiated by N-methyl-N-nitro-N-nitrosoguanidine or 3-methylcholanthrene [25].
- Carcinogenic activity of 1% hydroxy-anthrquinones and emodin (0, 280, 830, or 2500 ppm/rats) and (0–1250 ppm/mice) has been reported in several feeding studies [24].
- Induction of primary cutaneous melanin-containing tumors in C3H/HeN mice treated with aloe-emodin (in ethanol vehicle) and ultraviolet radiation, whereas no skin tumors were induced using aloe-emodin alone [24].
- Cells pre-treated with aloe-emodin showed increased sensitivity to both UV-A and visible light. Significant photo-oxidative damage to both RNA and DNA was associated with the phototoxicity induced by aloe-emodin. The phototoxicity mechanism involves the generation of reactive oxygen species and stable photo products with cellular components [6].
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<td><strong>Antiviral activity</strong></td>
<td><strong>Aloe-emodin</strong></td>
<td>- Dose-dependent inhibition of inducible nitric oxide synthase (iNOS) mRNA expression and nitric oxide (NO) production at 5–40 µM [17]. - Suppression of cyclooxygenase-2 (COX-2) mRNA expression and prostaglandin E2 (PGE2) production by 40 µM [17]. - Inhibitory activity (39.9%) on carrageenan-induced rat-paw edema [18].</td>
<td><strong>Aloin</strong> - Suppression of NO production at 5–40 µM, but it did not suppress PGE2 production [17]. - Strong inhibition (70%) of histamine release induced by Ca²⁺- independent inducer, compound 48/80, from mast cells comparing with indomethacin (35%) [19]. - Inhibitory activity (28.8%) on carrageenan-induced rat-paw edema [18].</td>
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<td><strong>Antioxidant activity</strong></td>
<td><strong>Aloe-emodin</strong></td>
<td>- Inhibitory activity against linoleic acid peroxidation catalyzed by soybean 15-lipoxygenase (IC₅₀ 6.5 ± 3 µmol/l) [20].</td>
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<td><strong>Wound healing activity</strong></td>
<td><strong>Aloe-emodin</strong></td>
<td>- A significant increase in wound healing rate (p &lt; 0.05), a higher level of re-epithelialization and angiogenesis, and a significant reduction of burn wound area (p &lt; 0.05) of excision wounds in mice [21].</td>
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<td><strong>Aristolochia bracteolate Lam.</strong></td>
<td>For bloating given together with <em>Sorghum bicolor</em> (Poaceae) grains [3]</td>
<td><strong>Aristolochia bracteolate</strong></td>
<td><strong>Aristolochia species</strong> were reported to be poisonous to man and livestock due to aristolochic acids, which were nephrotoxic, genotoxic, and carcinogenic in humans [35].</td>
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<td><strong>Antimicrobial activity</strong></td>
<td><strong>Methanol extract of leaf and bark</strong>: active against - <em>B. subtilis</em> (MIC = 39.06 µg/ml), <em>Salmonella typhi</em> (MIC = 625 µg/ml), <em>P. aeruginosa</em> (MIC = 1250 µg/ml), and <em>S. aureus</em> (MIC = 2500 µg/ml). [26]. - <em>Rhizopus stolonifer</em> and <em>Penicillium notatum</em> [26].</td>
<td><strong>Chloroform extract of leaf and bark</strong>: active against - <em>E. coli</em> (MIC = 312.5 µg/ml), <em>B. subtilis</em> (MIC = 2500 µg/ml), <em>P. aeruginosa</em> (MIC = 2500 µg/ml), and <em>S. typhi</em> MIC = 2500 µg/ml [26]. - <em>Aspergillus terreus</em> [26].</td>
<td><strong>Water extract of leaf and bark</strong>: active against - <em>A. terreus</em>, <em>Aspergillus niger</em>, and <em>P. notatum</em> [26]. 70% methanol extract and its chloroform fraction of the whole plant - Moderately active against <em>S. aureus</em>, <em>B. subtilis</em>, and <em>P. aeruginosa</em> [27]. <strong>Methanolic extract of the whole plant</strong>: active against - <em>S. aureus, Micrococcus luteus, B. subtilis, S. epidermidis, E. faecalis, MRSA and Enterobacter aerogenes, Yersinia enterocolitica, and P. vulgaris</em>, with a zone of inhibition nearly equal to that of the standard streptomycin against <em>B. subtilis</em> and <em>P. vulgaris</em> and exceeding that of streptomycin against <em>Y. enterocolitica</em> [28]. - <em>A. niger, Botrytis cinerea, Trichophyton rubrum, Curvularia lunata, A. flavus, and T. mentagrophytes</em> [28].</td>
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<td>Methanolic and aqueous extracts of leaves and stems of Yemeni <em>Aristolochia bracteolate</em></td>
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<td>Methanolic extract was more active than aqueous extract against standard <em>S. aureus</em> (MIC = 250 µg/ml), <em>B. subtilis</em> (MIC = 500 µg/ml), and <em>Micrococcus flavus</em> (MIC = 250 µg/ml). The methanol and aqueous extracts were active against MRSA strains (<em>S. epidermidis</em> 847 and <em>S. aureus</em> north German epidemic strain) [29].</td>
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<td>Acetone, ethanol, and petroleum ether extracts of the leaves, stem, and root</td>
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<td>Active against <em>E. coli</em>, <em>P. aeruginosa</em>, and <em>K. pneumoniae</em> except for the ethanol extract of the leaves and the acetone extract of the stems, which were inactive against <em>K. pneumoniae</em> [30].</td>
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<td>Chloroform, ethyl acetate, and methanol extracts of the roots</td>
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<td>Active against <em>B. subtilis</em> and <em>E. coli</em>, with ethyl acetate extract as the most active extract, especially against <em>E. coli</em> [31].</td>
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<td>96% ethanol extract of the plant</td>
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<td>Active against <em>E. coli</em>, <em>P. aeruginosa</em>, <em>B. subtilis</em>, <em>S. aureus</em>, and <em>C. albicans</em> [32].</td>
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<td><em>Aristolochic acid I</em></td>
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<td>The dose of 50 µg/ml showed antimicrobial activity approaching that of the positive control (ciprofloxacin 50 µg/ml) against <em>E. coli</em> and exceeding that of the ciprofloxacin against <em>B. subtilis</em>. [31].</td>
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<td>Anti-inflammatory activity</td>
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<td>Significant reduction (60%) of induced hind paw edema in rats compared to positive control ibuprofen (69%) [33].</td>
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<td>Leaves ethanol extract</td>
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<td>Anti-inflammatory activity using cotton pellet method in rats [34].</td>
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<td><em>Aristolochic acid</em></td>
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<td>Inhibition of the inflammation induced by immune complexes as well as immunological and nonimmunological agents. Its activity mainly results from its inhibitory effects on phospholipase A2 (PLA2) in regulating the synthesis of arachidonic acid, an intermediate in the production of prostaglandins [35].</td>
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<td>Antioxidant activity</td>
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<td>Promising antioxidant activity in reducing 1-diphenyl-2 picryl hydrazyl (DPPH) in a concentration-dependent manner up to 1 mg/ml. The ether fraction showed scavenging activity against NO-induced release of free radicals with 0.5 mg/ml and 1 mg/ml exhibiting maximum NO inhibition of 45%, thereby supporting the anti-inflammatory activity of the ethanolic extract [33].</td>
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<td>Leaves ethanol extract, its ethyl acetate, and ether fractions:</td>
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<td>Significantly increase in the antioxidant enzymes (catalase, ascorbate oxidase, glucose 6 phosphate dehydrogenase), as well as superoxide, DPPH, and hydroxyl radical scavenging activity [36].</td>
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<td>Methanol extract of <em>A. bracteata</em> (leaf, stem, and roots)</td>
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<td>DPPH radical scavenging activity at high concentrations (69.9% by 1000 µg/ml and 30.7% by 500 µg/ml) [29].</td>
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<td>Methanolic extract of Yemeni <em>A. bracteate</em> (leaves, stems)</td>
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<td>DPPH radical scavenging activity at high concentrations (69.9% by 1000 µg/ml and 30.7% by 500 µg/ml) [29].</td>
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<td>96% ethanol extract of the plant</td>
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<td>Moderate DPPH radical scavenging activity (48.8%) compared to the standard propyl gallate (89.7%) [32].</td>
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<td>Ethyl acetate and crude methanol extracts of the leaves:</td>
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<td>Potent scavenging activity of 2,2’-azino-bis (3-ethylbenzo-thiazoline-6-sulfonic acid (ABTS•+) radical cations with IC₅₀ values of 17.08 ± 0.44 and 28.12 ± 2.87 µg/ml, respectively, compared to the positive control ascorbic acid (IC₅₀ = 11.25 ± 0.49) [37].</td>
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<td><strong>Antiulcer activity</strong></td>
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<td><strong>Aqueous extract of leaves</strong></td>
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<td>▪ Significant (p &lt; 0.01) decrease of the ulcerated area and the</td>
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<td>volume and acidity of the gastric juice of ethanol-induced and</td>
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<td>pylorus ligation-induced gastric ulceration in pre-treated rats</td>
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<td>with the extract 800 mg/kg/body weight/day [38].</td>
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<td><strong>Ethanol whole plant extract</strong></td>
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<td>▪ Significant (p &lt; 0.05) decrease in the gastric volume and total</td>
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<td>acidity, number of ulcers, and ulcer score index of HCl-induced</td>
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<td>ulceration in the albino pre-treated rats with the extract.</td>
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<td>The percentage of ulceration inhibition was 59.09%, compared to</td>
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<td>75.87% by the standard ranitidine [39].</td>
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<td><strong>Sorghum bran diets</strong></td>
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<td><strong>Suppression of ulcerative colitis</strong></td>
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<td>▪ Polyphenol (3-deoxyanthocyanins and condensed tannins)-rich</td>
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<td>sorghum brans mitigate colonic injury and epithelial dysfunction</td>
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<td>induced during dextran sodium sulfate colitis by modulating ep</td>
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<td>ithelial cell proliferation and upregulating the proliferation</td>
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<td>and expression of Tff3, Tgfβ, and short-chain fatty acids (SCFA)</td>
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<td>[40].</td>
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<td>**Boswellia sacra Flueck./underbark/oleo-gum resin/ Burseraceae</td>
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<td>**Infected wounds by local appl. of hot aqueous extract of und</td>
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<td>er bark or paste made of ground underbark [4].</td>
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<td><strong>Antimicrobial activity</strong></td>
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<td><strong>Essential oil and methanolic extract of the branches</strong></td>
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<td>▪ Active against reference strains such as B. cereus, E. foecal</td>
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<td>is, E. coli, Listeria innocua, Salmonella enterica, Shigella dy</td>
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<td>senteria, S. aureus, Staphylococcus camorum, and Proteus mirabilis</td>
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<td>(with MIC of 1–32 µg/ml for essential oil and MIC of 100–200 µ</td>
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<td>g/ml for methanol extract) and clinical strains such as S. a</td>
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<td>ureus (with MIC of 2 µg/ml for essential oil and 100 µg/ml for</td>
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<td>methanol extract) and P. aeruginosa (with MIC of &gt; 32 µg/ml fo</td>
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<td>r essential oil and 200 µg/ml for methanol extract), as well as</td>
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<td>the fungal strains (standard C. albicans, clinical isolates of</td>
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<td>C. albicans, A. niger, and Aspergillus sp. (with MIC of 0.25–</td>
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<td>1 µg/ml for essential oil and MIC of 50 – &gt; 200 µg/ml for metha</td>
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<td>nol extract) [41].</td>
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<td><strong>Essential oil of oleo-gum resins:</strong> active against</td>
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<td></td>
<td>▪ Clinical isolates of B. subtilis, M. luteus, S. aureus, K. p</td>
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<td>neumoniae, E. aerogenes, P. aeruginosa, E. coli, P. vulgaris an</td>
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<td>d standard S. aureus, E. coli, and P. aeruginosa [42].</td>
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<td>▪ Standard S. aureus (MIC = 4.0 mg/ml), B. cereus (MIC = 2.0 mg</td>
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<td>/ml), E. coli (MIC = 4.0 mg/ml), and P. vulgaris (MIC = 3.0 mg/</td>
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<td>ml) and C. albicans (MIC = 8.0 mg/ml) [43].</td>
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<td>▪ Acetyl-11-keto-β-boswellic acid was more active (with MIC</td>
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<td>range of 2–8 µg/ml), and 11-keto-β-boswellic acid, as well as</td>
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<td>β-boswellic acid, were moderately active (MIC = 8–64 µg/ml)</td>
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<td>against standard S. aureus, MRSA, E. faecalis, E. faecium, S.</td>
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<td>epidermidis, and 112 isolates of various bacterial pathogens (MR</td>
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<td>SA 50, E. faecalis 22, E. faecium 18, S. epidermidis 12, and va</td>
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<td>ncocycin-resistant E. faecalis 10). All compounds were bacteriostatic and exhibited a MIC of &gt; 128 µg/ml [44].</td>
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<td>▪ Acetyl-11-keto-β-boswellic acid effectively inhibited the for</td>
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<td>mation of S. aureus and S. epidermidis biofilms, with a 50% biofilm inhibition concentration of 16–32 µg/ml. It also effectively eradicated the preformed biofilms. The 50% biofilm reduction concentration ranged from 32–64 µg/ml for both bacteria [44].</td>
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</table>

Boswellia gum is included in the list of safe substances, and the U.S. FDA permits its use as a food additive. The side effects are relatively very mild and not severe, and it can be considered quite safe when taken in the required and therapeutic dosages [51].

continued
### Table 1 Continued

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<thead>
<tr>
<th>Species/part used/ Family</th>
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<tr>
<td><strong>α-Pinene</strong></td>
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<td>• Active against some strains of <em>S. aureus</em> (MIC = 10–20 µL/mL), <em>S. epidermidis</em> (MIC = 5 µL/mL), and <em>S. pyogenes</em> (MIC = 5–10 µL/mL) [45].&lt;br&gt;• More active than streptomycin (positive control) against <em>P. aeruginosa</em>, <em>E. coli</em>, <em>B. subtilis</em>, <em>S. aureus</em>, yeast strains (<em>Hansenula anomala</em> and <em>Saccharomyces cerevisiae</em>), and fungi (<em>A. niger</em>, <em>Chae- tomium globosum</em>, <em>Mucor racemosus</em>, <em>Monascus anka</em>). The MIC values ranging from 62.5 to 125 µg/ml [46].&lt;br&gt;A fraction containing 99.5% α-pinene • Active against <em>E. coli</em>, <em>P. aeruginosa</em>, <em>Corynebacterium</em> sp., <em>S. aureus</em>, <em>B. cereus</em>, fungi (<em>Alternaria</em> sp., <em>Aspergillus nidulans</em>, and <em>A. niger</em>), yeast (<em>C. albicans</em>) with inhibition zones approaching as well as exceeding those of some commercial antibiotics [47].</td>
<td>Anti-inflammatory activity Essential oil of Omani <em>B. sacra</em> oleo-gum resin (Shabi grade): • Significant reduction of rats hind paw edema by 21.3, 18.8, 17.1, and 25.8% after 1, 2, 3, and 4 h, respectively, compared to aspirin (standard drug), which showed 12.9, 14.4, 14.7, and 22.0% edema inhibition after 1, 2, 3, and 4 h, respectively [48].&lt;br&gt;α-Pinene • Inhibitory effects on histamine release in rat peritoneal mast cells, the activation of inhibitor of Kappa B Kinase (IKK-β), nuclear factor-kappa B (NF-κB), and caspase-1 in human mast cells, and NF-κB and Jun N-terminal kinase (JNK) activation in chondrocytes [49].&lt;br&gt;• Decrease in the production of interleukin-6 (IL-6), tumor necrosis factor-α (TNF-α), COX-2 and iNOS expression, PG E2 and NO production; activations of the mitogen-activated protein kinases (MAPKs) and NF-κB in lipopolysaccharide (LPS)-stimulated mouse peritoneal macrophages. [49].&lt;br&gt;• Abolishing mechanical sensitization induced by complete Freund’s adjuvant at 70 mg/kg, p. o. similar to the positive control drug gabapentin (70 mg/kg, p. o.). This indicates its role in the management of inflammatory and neuropathic pain [50].&lt;br&gt;Boswellic acids • Acetyl-11-keto-β-boswellic acid is the most important inhibitor of 5-lipoxygenase [51].</td>
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Table 1 Continued

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<tr>
<td>Acetyl-11-α-keto-β-boswellic acid (AKBA)</td>
<td>▪ In the acetic acid-induced writhing test, AKBA inhibited dose-dependently the induced writhing (40.98% by 100 mg/kg and 55.19% by 200 mg/kg), compared to the positive control nimesulide (47.54% by 2 mg/kg p.o.), while in tail-flick method, AKBA at 100 mg/kg and 200 mg/kg exhibited similar antinociceptive activity (58.37% and 58.74 maximum possible effect [MPE], respectively, compared to positive control with 62.59% MPE), as early as 30 min. The % MPE of the AKBA (100 mg/kg and 200 mg/kg) was increased up to 60 min (97.28% and 114.79% MPE, respectively compared to positive control with 136.42% MPE) [53].</td>
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<td>Antioxidant activity</td>
<td>Essential oils of different grades (Hougari regular, royal upper, and Royal lower grades), as well as the sub-fractions (n-hexane and 40% MeOH/CH₂Cl₂ extracts of the crude methanol extract) obtained from Omani <em>B. sacra</em> oleo-gum resin: ▪ Inhibition of superoxide anion (&gt; 50%), while the CH₂Cl₂ fraction and the sub-fraction of 40% CH₂Cl₂/n-hexane fraction, as well as essential oil of Shabi frankincense grade, showed moderate inhibitory activity of DPPH free radicals [54].</td>
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<tr>
<td>α-Pinene</td>
<td>▪ Moderate DPPH, superoxide anion radicals scavenging activity, and reducing power activity [46].</td>
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<td>Mastitis by local appl. of a paste made of fresh oleo-gum resin boiled with milk [4].</td>
<td>Antibacterial, anti-inflammatory, and antioxidant</td>
<td>▪ Acetyl-11-α-keto-β-boswellic acid-mediated silver nanoparticles showed superior antibacterial, anti-inflammatory, and antioxidant properties than cefepime in mastitis-induced mammary glands in mice [55].</td>
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<td>Soqotraen <em>Boswellia</em> species (such as <em>B. amero</em> Balf.f.; <em>B. builoto</em> Thulin; <em>B. dosiconidis</em> Thulin; <em>B. elongate</em> Balf.f.; <em>B. nan</em> Hepper; <em>B. popoviana</em> Hepper; <em>B. socotran</em> Balf.f.)/wood, oleo-gum resin/Burseraceae [4]</td>
<td>Insecticidal activity</td>
<td>α-Pinene</td>
<td>▪ As an insecticide, especially on mosquitoes like <em>Culex pipiens</em> causing avian paludisme and Nile fever vector, or on dengue’s vector <em>Aedes aegypti</em> [56].</td>
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<td>Cissus rotundifolia Vahl/unspecified part/Vitaceae [3]</td>
<td>Stomach pain by giving boiled plant to livestock [3].</td>
<td>Antibacterial activity</td>
<td>Methanolic extract of the leaves of Yemeni plant</td>
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<td>Anti-inflammatory activity</td>
<td>70% Methanolic extract of nonflowering aerial parts</td>
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<td><strong>Analgesic activity</strong></td>
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<td>70% methanolic extracts of nonflowering aerial parts</td>
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<td>Central analgesic effect</td>
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<td>- Significant delay of the mean reaction time on the hot plate (9.86 and 10.56 s at 100 and 200 mg/kg bw, respectively) compared to the reference drug indomethacin (10.30 s at 10 mg/kg bw) [58].</td>
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<td>Peripheral analgesic effect</td>
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<td>- Significant reduction of acetic acid-induced writhing in mice (19.22 ± 0.86 and 18.25 ± 0.58 writhes at 100 and 200 mg/kg bw, respectively) compared to the indomethacin (17.60 ± 1.21 writhes at 10 mg/kg bw) [58].</td>
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<td><strong>Antioxidant activity</strong></td>
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<td>Fresh Yemeni plant juice 0.59 ml/g fresh weight</td>
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<td>- Moderate antioxidant activity (41.26 ± 3.96%, 69.95 ± 0.13%, 85.92 ± 1.64%) inhibition of guinea pig liver homogenate oxidation at 50, 100, and 200 µl, respectively measured by the thiobarbituric acid reagent substances method [59].</td>
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<td>Methanolic extract of leaves and stems of Yemeni plant</td>
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<td>- 59.8% and 92.4% DPPH radical scavenging activity at 500 and 1000 µg/mL, respectively [29].</td>
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<td>Ethanolic extract and its fractions of the aerial part (leaves and stems) of Yemeni plant</td>
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<td>- n-Butanol fraction showed a high activity to scavenge DPPH radical with IC₅₀ 25.04 ± 0.76 µg/ml compared to ascorbic acid (IC₅₀ 38.55 ± 0.83 µg/ml). In streptozotocin-induced diabetic rats, the ethanolic extract (300 mg/kg, p. o.) and its fractions (methylene chloride and n-butanol) at 2 doses (150 and 300 mg/kg, p. o.) showed a significant decrease in the content of malonaldehyde (lipid peroxidation products). All the tested samples at dose (300 mg/kg p. o.) significantly normalized the liver non-enzymatic antioxidant glutathione content of diabetic rats [60].</td>
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<td>Astragalin (kaempferol-3-O-β-D-glucopyranoside) and β-Amyrin isolated from unflowering aerial parts exhibited significant DPPH radical scavenging activity (IC₅₀ = 5.77 and 5.79 µg/mL, respectively) relative to that of ascorbic acid (IC₅₀ = 33.3 µg/mL) [61].</td>
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<td><strong>Anti-ulcerative activity</strong></td>
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<td>70% methanolic extracts of nonflowering aerial parts</td>
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<td>- Pre-treatment with 100 and 200 mg/kg bw produced a significant protection against ulcer formation in indomethacin-induced ulcers in pyloric-ligated rats [58].</td>
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<td><strong>Cyphostemma digitatum</strong> (Forssk.) Desc./leaves/ Vitaceae [3]</td>
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<td>Appetite stimulation by a beverage made of boiled leaves with salt given with water [3].</td>
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<td><strong>Bone fracture</strong> by applying plant mixed with eggs as a cast [3,63]</td>
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<td><strong>Food flavoring</strong></td>
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<td>- Acidic [62] and salty taste.</td>
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<td><strong>Nutritional value</strong></td>
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<td>- Vitamin C and E (including β- and γ-tocotrienol forms that are very rare in nature), provitamin A, carotenoids, and phenolic compounds [62].</td>
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<td>The shrub is known to be avoided by goats even during severe drought even though its leaves remain green for a long time as it is quite drought resistant [65].</td>
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<td><strong>Psidia punctulate</strong> (DC.) Vatke/unspecified part/ Asteraceae [3,63]</td>
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<td>Analgesic activity</td>
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<td>Leaf exudate</td>
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<td>- Analgesic effect in a mouse tail-flick experiment [64]. A diterpene (6 p, 17, 19-ent-trachylobanetriol) isolated from leaf exudate and stem extracts</td>
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<td>- Appreciable analgesic effect at a dose of 50 mg/kg bw with a p-value of 0.02 when compared with aspirin [64].</td>
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<td>The fresh leaves are highly toxic and corrosive to the mouth and palate mucosa, and no herbivore was found feeding on this plant in more than 1,500 visited sites in central Yemen. The toxicity may be caused mechanistically by microscopic spiny crystals (presumably composed of calcium oxalate) on the leaves photographed under the polarized light microscope [62].</td>
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### Table 1

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</table>
| **Pulicaria undulata** (L.) C. A. Mey./ unspecified part/ Asteraceae [3] | Insect repellent in livestock shelters [3]. | **Insecticidal activity**  
Aerial part essential oil:  
▪ Active against *Tregoderma granarium* and larvae of *Tribolium castaneum* [66].  
**Methanolic extract**  
▪ Toxic effect on *Spodoptera littoralis* (the cotton leafworm) [67].  
**Carvotanacetone**  
▪ A high larvicidal activity on 6 mosquitoes with LC50 lower than 10 µg/ml [68]. | Adding the aerial part methanolic dried extract to chicken food was found as safe growth promotors [69]. |
| **Rumex nervosus** Vahl/unspecified part/Polygonaceae [2] | Pimples in goats’ eyes, treated with *Rumex nervosus* mixed with salt [2]. | **Antibacterial activity**  
▪ Buffered methanol, ethanol leaf extracts of Yemeni plant [70], and methanol extracts of aerial part [71] and leaves [72], as well as the ethanolic leaves extract [73], were active against *S. aureus*.  
▪ Sub-fraction (containing palmitoleic acid methyl ester [28.35%] and palmitic acid methyl ester [25.37%] as major components) obtained from the ethyl acetate fraction of the crude methanolic extract of the Yemeni plant leaves showed promising activity against *S. aureus* [74].  
▪ Salt was found to have antimicrobial activity against several microorganisms, including *S. aureus* [75]. |  |
| **Trigonella foenum-graecum** L. seeds/ Fabaceae [3] | For constipation by giving the fenugreek seeds together with *Hordeum vulgare* (Poaceae) grains [3]. | **Mild bulk-forming laxative effect**  
*Trigonella foenum-graecum* L. seeds  
▪ Mild bulk-forming laxative effect due to the content of dietary fibers (insoluble [20%] and soluble fraction [30%], which is mostly galactomannan) [78].  
*Hordeum vulgare* grains  
▪ Mild bulk-forming laxative effect due to high content of dietary fibers (soluble [mostly β-glucan] and insoluble fibers) [79]. |  
*Trigonella foenum-graecum* seeds  
▪ Included in the AHPA class 2b. (not to be used during pregnancy), as well as in the list of generally recognized as safe when used as a spice or flavoring [80].  
▪ Myopathy in ruminants [80].  
▪ Some serious toxicological side effects, such as teratogenic, reproductive (antifertility, abortifacient), effects, neurotoxicity, and other cellular and molecular alterations, not without controversy, have been reported [81–84]. |
The mechanisms of action of these pharmacological activities were reported for some constituents. The antimicrobial activity of anthraquinones was ascribed to several mechanisms such as the inhibition of the microorganisms enzymes (such as penicillinase) by rhein, emodin, and aloe-emodin [93]; the inhibition of the microorganisms enzymes (such as penicillinase) by anthraquinones was ascribed to several mechanisms such as the inhibition of nucleic acid synthesis in MDCK cells. A 50% inhibitory concentration value of aloe-emodin on virus yield was less than 0.05 µg/ml [95]. The antibacterial mechanism of action of acetyl-11-keto-β-boswellic acid (AKBA) was attributed to its ability to disrupt the permeability barrier of microbial membrane structures [44]. α-pinene was also found to induce toxic effects on the membrane structure (membrane expansion, increased membrane fluidity) and functions (inhibition of a membrane-embedded enzyme) of yeast and bacteria [56].

The anti-inflammatory mechanisms of action of Aloe latex, B. sacra, and A. bracteolate were reported for some of their constituents (Table 1). AKBA, as the most important inhibitor of 5-lipoxygenase, binds to 5-lipoxygenase in a calcium-depen-
Mastitis is a disease with a severe economic impact, causing production losses in the dairy industry. *S. aureus* is one of the leading causes of mastitis, resulting in an annual economic loss of $2 billion globally. *S. aureus* is also responsible for the emergence of resistant organisms and antibiotic residues in dairy products [55]. Acetyl-11-α-keto-β-boswellic acid-mediated silver nanoparticles (BANS, 0.12 mg · kg$^{-1}$) were reported to exert superior antibacterial (36%), anti-inflammatory (95%), and antioxidant effects compared with the antibiotic cefepime (1 mg · kg$^{-1}$ ; intraperitoneal) in *S. aureus*-induced murine mastitis. BANS treatment significantly (p < 0.05) reduced bacterial load, *C*.-reactive protein, superoxide dismutase, catalase activities, and neutrophil infiltration in affected mammary glands [55]. The antibacterial, anti-inflammatory, and antioxidant effects of acetyl-11-α-keto-β-boswellic acid could lend some support for the local use of oleo-gum resin of *B. socra*, which may be able to target pathogens such as *S. aureus* present at the teat opening and thus help to prevent bacteria that cause mastitis from entering the teat canal and colonizing the mammary tissue, as well as reduce the inflammation of the udder surface.

*S. aureus* is among the most common causes of ocular infections, including blepharitis, dacryocystitis, conjunctivitis, keratitis, and endophthalmitis. Humans are not the only reservoir for this organism because the organism can be isolated from companion animals, livestock, and wild animals [100]. In the infection process, myeloperoxidase, NADPH oxidase, and NO synthase generate reactive species [101]. Therefore, the antibacterial activity of both *Rumex nervosus* and salt against *S. aureus*, together with the antioxidant activity of *R. nervosus* to reduce the oxidative stress triggered by this bacterial infection, could justify their combined use to treat pimplies in goats’ eyes.

Bloating is a disorder of ruminants caused by gas retention in the stomach. It results most commonly from excessive foaming of the contents of the rumen, which is the primary cause of bloating in animals feeding on legumes or lush young grass, and of feedlot bloating. Nonfoamy or free gas bloating is less common than foamy bloating and results from a variety of causes. It is predominantly a disorder of cattle but may also occur in sheep and other domestic ruminants. The total economic cost of bloating comes in many forms: losses by death and culling of bloating prone animals; losses of production from animals that were bloated and survived and the disruption of regular farm work and management programs; losses due to the use of less productive but safer pastures; and the cost of preventive measures and treatment [102]. Bloating can develop by the action of ruminal microflora, such as bacteria, fungi, and protozoa, that break down cellulose and protein by microbial fermentation. As such, antibiotics can control bloating based on the principle of reduced microbial activity. Penicillin was the first antibiotic used to control legume bloating, but its use was soon discontinued because of the drug’s rapid development of microbial resistance. Recently, the ionophore antibiotics monensin (rumensin) and lasalocid have been used for bloating protection [103]. Consequently, the anti-microbial activity, together with the anti-inflammatory, antioxidant and antiulcer activities of *A. bracteolate* (with its constituent aristolochic acid) and the antiulcer effect of *S. bicolor* grains (Table 1), could support their combined use to reduce livestock bloating by decreasing microbial fermentation, and inflammation (e.g., inflammation of the peritoneum, a common cause of vagal nerve damage, in the case of free gas bloating [104]) as well as the stress facing the animals during bloating.

In cattle, the source of abdominal pain can originate from visceral stretch receptors within the mesentery, organ capsules, or ligaments, muscular spasms or inflammation, and ischemia. Pain can also arise from parietal sources, including the parietal peritoneum, abdominal muscles or rib cage, or bacterial-associated mechanical or functional obstruction, inflammation, or other damage to the gastrointestinal tract wall. Additionally, extra-abdominal conditions can mimic intra-abdominal pain [105]. Hence, the pharmacological properties (antibacterial, anti-inflammatory, antioxidant, analgesic, and antiulcer activities, Table 1) of *C. roundifolia* and its constituents, astragalin and β-amyrin, could ex-
plain its effectiveness for combating livestock abdominal pain and associated inflammation.

The use of *P. punctulata*, mixed with eggs as a cast, for bone fracture (► Table 1) could be supported by the analgesic effect of a diterpene, isolated from its leaf exudate and stem extracts, that may lessen the pain associated with a bone fracture. The suitability of this method will depend on the type of fracture. In simple closed fractures, casting may be enough to stabilize the bones and allow healing, and the analgesic action of *P. punctulata* may support the animal’s wellbeing. However, this will not be enough to heal the bones in open and complicated fractures, and other measures are needed.

The use of *C. digitatum* leaves (dried thick paste of boiled leaves) of plants. Thus, it is worth investigating the insecticidal activity of these spice and food flavoring in salads and some Yemeni dishes) for is utilized in Yemen for a variety of culinary purposes (► Table 1). Besides the toxicological activities of *Aloe* latex constituents, the use of the latex as a laxative in human medicine has not been prohibited, but it was restricted in dosage and period of treatment for cases of occasional constipation and when no laxative effect can be attained through diet change or the use of bulk-forming products [23]. Moreover, it has been reported that by considering the minimal use of Aloes in veterinary medicine (very infrequent, short-term treatment of individual animals only) and the limited absorption of the active principles after oral administration, the risk from veterinary use of Aloes to the consumer of foodstuffs of animal origin is considered negligible [5].

Fenugreek is included in the list of GRAS (generally regarded as safe) when used as spice or flavoring and is also classified as AHPA class 2b, not to be used during pregnancy (this classification is based on relatively high doses used for therapeutic purposes in contrast to lower amounts generally used in cooking and has not been associated with its use as a spice) [80]. Studies in animal models suggest a low acute toxic potential of fenugreek seeds by oral route at 2 and 5 g/kg bw in mice and rats, respectively, and intraperitoneal route at 0.65 and 3.5 g/kg bw in mice and rats, respectively [81]. As described in cattle [80], myopathy may be caused by feeding cattle solely on fenugreek straw, leading to intoxication or an induced deficiency state [110].

The literature showed controversial results regarding fenugreek seeds’ teratogenic, antifertility, and neurotoxic effects, as well as resulting histopathological changes in vital organs (e.g., liver), hematological parameters, and clinical biochemistry. Several human and animal studies revealed teratogenic effects from congenital malformations to death; testicular toxicity and antifertility effects in male animals associated with oxidative stress and DNA damage; and antifertility, anti-implantation, and abortifacient activity in female animals. These effects have been suggested to be the consequences of a saponin compound of fenugreek seeds. In addition, the studies suggest that antifertility effects, hormonal disturbances, and adverse effects during pregnancy can be due to long-term daily use of at least 100 mg/kg bw of animal [81].

On the other hand, the prenatal oral exposure of 250, 500, and 1000 mg/kg/day of furostanol saponin glycoside (75.23%) and low molecular weight galactomannans based standardized fenugreek seed extracts were found safe and devoid of maternal and embryo-fetal toxicity [82–83]. Additionally, no effect on the fertility, and fetus’s growth and development was observed in pregnant rats treated with fenugreek seed at 5 or 20% of the diet. Moreover, 75 mg/kg of trigomeline administered orally to pregnant and nonpregnant female rats showed no antifertility or abortifacient effects.

Although several studies have shown fenugreek to be neuroprotective, accumulating evidence suggests that fenugreek seeds may have neurodevelopmental, neurobehavioral, and neuropsychological side effects [81]. Several studies claim the lack of histopathological changes in vital organs (e.g., liver), hematological parameters, and clinical biochemistry caused by fenugreek [81, 84]. However, several cellular and molecular alterations have been noted in animal models studies, especially in some vital organs (liver, kidney), hematological parameters, and blood biochemistry (e.g., a significant increase of hematocrit and reduction of serum total protein levels) by a daily oral administration of alcoholic extract 200 mg/kg rat bw for 4 weeks; A necrotic effect on the liver and the kidney tissues was observed at doses of 100,150, or 200 mg/kg of aqueous extract orally administered twice a week.
for 4 weeks to rats; crude saponins treatment to Hisex-type chicks caused necrosis of hepatocytes with lymphocytic infiltration and epithelial degeneration of renal tubules [81]. Due to multiple beneficial health properties of fenugreek seeds (e.g., antidiabetic effect, hypcholesterolemic influence, antioxidant potency, digestive stimulant action, and hepatoprotective effect) and its usage as a spice for seasoning and a flavoring agent and in comparatively larger quantities in making soups and pancakes [78], the reported toxicological effects should be taken seriously. Further research is needed to clarify these serious side effects and demonstrate their mechanisms of action. However, different extraction methods can lead to a different spectrum of constituents. Since the seeds were fed without extraction and no adverse effects in target animals were described, the risk of ethnoveterinary use may be limited.

The local use of C. procera latex in Yemen to treat swelling was documented without mentioning any side effects [3]. However, the local administration of the latex was reported to induce an intense inflammatory response. In addition, the dried latex was found to produce significant inflammation and hyperalgesia in different models of inflammation in rats, and hence, the dried latex-induced inflammation was used to investigate the anti-inflammatory and analgesic activity of anti-inflammatory drugs [85, 111]. No side effects were reported for E. arbuscula latex, although related Euphorbiaceae species were found to contain diterpenes derivatives (e.g., phorbol derivatives with skin inflammatory and tumor-promoting activities) [86]. Therefore, further research of these remedies is needed to illustrate their effectiveness and safety. Moreover, we emphasize the need to raise the livestock owner’s awareness of the harmful side effects of herbal remedies and the appropriate way to use them either by limiting the dosage and period of treatment, avoiding their use during pregnancy and lactation, or by using safer alternatives.

Conclusion

In this paper, 14 ethnoveterinary plant species used in different areas of Yemen have been reviewed for their uses based on a scientific literature search on their biological activities and toxicological properties. The scientific data has indicated several in vivo and in vitro pharmacological studies that could support the claimed healing activities of 11 plant species, namely, A. bracteolate, S. bicolor, C. rotundifolia, C. digitatum, P. punctulate, P. undulata, R. nervosus, T. foenum-graecum, and H. vulgare, for their indications, and of Aloe species latex, B. sacra, and Socotraen Boswellia species for some of their indications. Three of these plants (Aloe species, A. bracteolate, T. foenum-graecum) exhibited some slight or even serious toxicological properties. Hence, our work offers valuable information on some Yemeni ethnoveterinary remedies that could be utilized for the benefit of livestock owners to justify using these remedies and increase awareness of their side effects and the rational way to use them. Moreover, we highlight the need for further scientific research on the effectiveness and safety of these plants.

Supporting information

The Supporting Information (Supplementary Table 1S) describes the plant part, livestock species, indication for use, preparation, ACTvetcode, and references for the 14 remedies for livestock.

Contributors’ Statement


Acknowledgements

The authors thank Leiden University for kindly giving access to their library.

Conflict of Interest

The authors declare that they have no conflict of interest.

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