Insulin Resistance and Free Androgen as Predictors for Ovarian Hyperstimulation Syndrome in Non-PCOS Women

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infertility, polycystic ovary syndrome, ovarian hormone, assisted reproductive techniques

Introduction
Ovarian hyperstimulation syndrome (OHSS) is a serious and completely iatrogenic complication of controlled ovarian hyperstimulation [1]. It is an important issue in the field of IVF (in vitro fertilization). However, the exact pathogenesis of the OHSS remains less clear, vascular permeability mediated by human chorionic gonadotropin (hCG) appears to be the most probable mediator. OHSS is categorized into four classes, including mild, moderate, severe, and critical forms of the syndrome on the basis of the severity of symptoms, signs, and laboratory parameters. Mild OHSS is characterized by the enlargement of bilateral ovaries (up to 8 cm) with abdominal bloating and mild abdominal pain. The moderate form of OHSS is described by the enlargement of the ovaries (up to 12 cm), as well as ultrasound finding of ascites. The severe form of OHSS is defined by the observation of large ovarian cysts (> 12 × 12 cm), clinical manifestation of ascites with or without hydrothorax, with abnormality findings like sodium, potassium, and osmolarity serum leading to decreased urine output and hypovolemic shock. Critical OHSS is characterized when there is severe ascites on ultrasound examination or hydrothorax, hematocrit of over 55 %, WBC over 25 000/ml, oliguria or anuria, creatinine ≥ 1.6 mg/dl, creatinine clearance less than 50 ml/min, thromboembolism, or acute respira-
OECD distress syndrome [1, 2]. The reported incidence of the severe and mild form of OHSS ranges from 0.5–33 %, respectively [2–4]. Polycystic ovary syndrome (PCOS) appeared to be the major risk factor for OHSS in many studies [2, 5–7]. PCOS increases the risk of OHSS because of the hyper-response to ovarian stimulation, since too many antral follicles are already present at the beginning of the cycle. Hyperinsulinemia and hyperandrogenism are identifying features of PCOS and both promote early folliculogenesis and frequently a multi-follicular response following the ovulation induction [2, 8, 9]. Hyperinsulinemia and hyperandrogenism are causes rather than consequences of polycystic ovarian morphology (PCOM), therefore high insulin and/or testosterone levels may increase the risk of OHSS by inducing PCOM. Insulin-like growth factor 1 (IGF-1) and insulin can stimulate vascular endothelial growth factor (VEGF) production by luteinized granulosa cells in vitro [10], but this is insufficient evidence to suggest that insulin resistance is directly responsible for OHSS. OHSS is prevalent among PCOS infertile women, however, it is also observed among infertile non-PCOS women. Therefore, we assessed the probability of the presence of hyperinsulinemia and hyperandrogenemia in the incidence of OHSS in the non-PCOS women undergoing ART cycles and to evaluate if a threshold level is present.

Subjects and Methods

This prospective follow-up study was conducted on 144 infertile and non-PCOS women with regular menstrual cycle undergoing IVF or intracytoplasmic sperm injection (ICSI) cycles from December 2012 to December 2014. Exclusion criteria were patients with PCOS diagnosis based on Rotterdam criteria [11], previous ovary surgery, ovarian insufficiency, endometriosis, some medical conditions like (diabetes mellitus, thyroid dysfunction, hyperprolactinemia, Cush-ing’s syndrome [12]), being smoker and use of the drugs like spironolactone, aspirin, corticosteroid, and metformin. The selection of the participants was made from the two referral infertility clinics at Ahvaz, Iran. The study protocol complies with Helsinki Declaration and was approved by the Research Ethics Committee of Ahvaz Jundishapur University of Medical Sciences. Informed consent was obtained from all participants. The blood samples of the participants were collected on 1–3 days of their cycles after a fasting period of at least 8 h. The samples were centrifuged at 3000 g for 10 min, and the serums were stored at −20 °C until assaying. Fast ing blood sugar (FBS) level was measured by Autoanalyzer (Selec tra, Flexer Model, Netherlands) using glucose oxidase method. Testosterone was assayed using a direct, competitive chemilumi nescent immunoassay (CLIA) performed on the manufacturer’s ana lyzer (Liaison Analyzer, Diasorin Inc., USA). The method for quanti tative determination of insulin was a sandwich chemiluminescent immunoassay performed on the manufacturer’s analyzer (Liaison Analyzer, Diasorin Inc., USA). The level of SHBG was detected by an enzyme-linked immunosorbent (ELIZA) kits (IBL International, Hamburg, Germany). The free index (FAI) was determined as the total testosterone × 100/SHBG. The IR was calculated using the homeostasis model assessment (HOMA) [HOMA-IR = (insulin × glu cose)/22.5] [13]. Anti-Müllerian hormone (AMH) was assayed using Elecsys Cobas e 411, Roche Kiet, Germany. Patients underwent vag inal ultrasonography scan for polycystic ovary morphology (PCOM) and antral follicle count (AFC) in 1–3 days of their cycles and then based on the indication of the treatment underwent long protocol with the use of GnRH agonist (bucerelin) and gonadotropins. The number of preovulatory follicles was recorded. Triggering of the ovulation was performed by hCG and oocytes retrieved were re corded, then the patients were followed-up for the signs and symp toms of OHSS (mild, moderate, and severe), then they were divid ed into two groups with and without OHSS (mild, moderate, and severe) for the analysis. We used 10 000 IU hCG for trigger of ovula tion, but for the patients with probability OHSS based on the number of preovulatory follicles and the level of estradiol or rapid increased level of estradiol, we choose the coaching method without further gonadotropin stimulation and delaying the use of hCG for 1–3 days until estradiol levels flatten or decline, and used lower dose of hCG (5000 IU). The total number of ART cycles was 1225 and finally of those, 330 patients developed OHSS (27 %, including 20, 5, and 2 % for mild, moderate and severe OHSS), respectively, then 66 non-PCOS patients with mild, moderate or severe OHSS were used for final analysis.

The data were analyzed using Statistical Package for the Social Sciences (SPSS, version 11.5, Chicago, IL, USA). To compare quant itative variables, independent sample t-tests were applied. Chi square test was used to test dichotomous variables between groups. The forward stepwise multiple logistic regression model was constructed for OHSS. We used the absence or presence of OHSS as the dependent variable. The univariate analyses were con ducted on the variables including age, BMI, HOMA-IR, FAI, AMH, AFC, preovulatory follicles, and oocytes retrieved, and dichotomous variable of PCOM. Then, the variables assessed significant in univariate regression analysis were entered as independent variables in multivariable logistic regression analysis. The receiver operating characteristics (ROC) curve was used to identify the best cut-points value for those variables that were independent predictors of OHSS development in non-PCOS patients.

Results

A total of 144 patients were followed-up in this study. The incidence of mild, moderate, and severe OHSS in two centers were 20, 5, and 2 %, respectively. There was no significant difference between patients who developed OHSS and those who did not, regarding age and oocytes retrieved using univariate logistic regression analyses (Table 1). Only two patients were admitted with sever OHSS (0.14 %). Patients with OHSS had higher chance to have ovaries with polycystic morphology (74 %), about three times more than patients who did not develop OHSS (29 %) (p < 0.001) (Table 1). In addition, univariate logistic regression analysis showed that mean scores of BMI, FAI, HOMA-IR, AMH, AFC, PCOM and preovulatory follicles were significantly higher in OHSS compared with non-OHSS patients (Table 1). Of the 9 variables: BMI, HOMA-IR, FAI, AFC, AMH, PCOM, and preovulatory follicles were risk factors, while the age and retrieved oocytes were not (Table 1). The 7 variables that provided significant in the univariate analyses were chosen as independent variables included in the multivariable logistic regression analysis, as a result, a total of 5 risk factors, BMI, HOMA-IR, FAI, PCOM, and pre-
ovulatory follicles entered the equation. The maximum contribution was HOMA-IR followed by PCOM, FAI, preovulatory follicles, and BMI (▶Table 2). To assess the optimal cut-points for HOMA-IR, FAI, AFC, and AMH to predict the development of OHSS in non-PCOS patients who undergo ART, ROC curves were obtained. The best cut-points for HOMA-IR, FAI, AMH, AFC, and preovulatory follicles were 2.36 (AUC = 0.78, sensitivity = 75%, specificity = 70%), 3.9 (AUC = 0.67, sensitivity = 54%, specificity = 84%), and 3.3 ng/ml (AUC = 0.79, sensitivity = 80%, specificity = 71%), respectively (▶Figs. 1 and ▶2).

![Table 1](https://example.com/table1.png)

<table>
<thead>
<tr>
<th>Numerical Variable</th>
<th>OHSS (n = 66)</th>
<th>Non-OHSS (n = 78)</th>
<th>95% CI</th>
<th>p-Value</th>
</tr>
</thead>
<tbody>
<tr>
<td>Age (years)</td>
<td>31.07 ± 5.63</td>
<td>29.85 ± 5.68</td>
<td>0.980–1.102</td>
<td>0.827</td>
</tr>
<tr>
<td>BMI (kg/m²)</td>
<td>26.21 ± 2.20</td>
<td>23.21 ± 1.96</td>
<td>1.517–2.260</td>
<td>&lt;0.001</td>
</tr>
<tr>
<td>HOMA-IR</td>
<td>3.21 ± 1.6</td>
<td>1.58 ± 0.79</td>
<td>2.263–5.251</td>
<td>&lt;0.001</td>
</tr>
<tr>
<td>FAI</td>
<td>5.52 ± 4.68</td>
<td>2.43 ± 1.53</td>
<td>1.235–1.748</td>
<td>&lt;0.001</td>
</tr>
<tr>
<td>AMH (ng/ml)</td>
<td>6.7 ± 5.6</td>
<td>3.1 ± 2.9</td>
<td>1.013–1.529</td>
<td>0.012</td>
</tr>
<tr>
<td>AFC (number)</td>
<td>13.73 ± 6.11</td>
<td>6.7 ± 5.6</td>
<td>1.108–1.723</td>
<td>&lt;0.001</td>
</tr>
<tr>
<td>PCOM</td>
<td>49 (74)</td>
<td>23 (29)</td>
<td>3.303–14.384</td>
<td>&lt;0.001</td>
</tr>
<tr>
<td>Preovulatory follicles (number)</td>
<td>15.3 ± 5.7</td>
<td>8.3 ± 2.5</td>
<td>1.147–2.063</td>
<td>&lt;0.001</td>
</tr>
<tr>
<td>Oocytes retrieved (number)</td>
<td>14.3 ± 8.6</td>
<td>9.9 ± 5.5</td>
<td>0.991–1.208</td>
<td>0.06</td>
</tr>
</tbody>
</table>

Data are expressed as means ± standard deviation or number (percentage). OHSS: Ovarian hyperstimulation syndrome; BMI: Body mass index; HOMA-IR: Homeostasis model assessment-insulin resistance; FAI: Free androgen index; AMH: Anti-Müllerian hormone; AFC: Antral follicle count; PCOM: Polycystic ovary morphology.

![Table 2](https://example.com/table2.png)

<table>
<thead>
<tr>
<th>Women underwent ART</th>
<th>Odds ratio (OR)</th>
<th>95% CI</th>
<th>p-Value</th>
</tr>
</thead>
<tbody>
<tr>
<td>BMI</td>
<td>1.434</td>
<td>1.070–1.921</td>
<td>0.016</td>
</tr>
<tr>
<td>HOMA-IR</td>
<td>4.704</td>
<td>2.422–9.137</td>
<td>&lt;0.001</td>
</tr>
<tr>
<td>FAI</td>
<td>1.613</td>
<td>1.181–2.204</td>
<td>0.003</td>
</tr>
<tr>
<td>PCOM</td>
<td>3.161</td>
<td>1.041–9.599</td>
<td>0.042</td>
</tr>
<tr>
<td>Preovulatory follicles (number)</td>
<td>1.539</td>
<td>1.147–2.063</td>
<td>0.004</td>
</tr>
</tbody>
</table>


Discussion and Conclusions

PCOS is known as a major primary risk factor for OHSS in a large number of studies [2, 7, 14, 15]. On the other hand, vascular endothelial growth factor (VEGF) is proved to be the principal mediator in OHSS pathophysiology [16]. In addition, it has been reported that an increased expression of VEGF’s mRNA within the hyperthecal stroma of women with PCOS may be responsible for their higher risk of OHSS [17]. It was shown that insulin and insulin-like growth factor (IGF) promote VEGF production in luteinized granulosa cells [18]. It has been shown that the incidence of OHSS is related to the stimulation regimens used [3]. PCOS isolated characteristics (i.e., PCOM, hyperinsulinemia, and hyperandrogenism) were revealed to be possible predisposing factors for OHSS. In the
current study, BMI, HOMA-IR, FAI, preovulatory follicles and the presence of PCOM retained as risk factors for OHSS based on multivariate logistic regression analysis. In some moderate-quality evidences, the use of insulin-sensitizing agents (i.e., metformin) decreased the risk of OHSS in PCOS women who underwent ART [14, 18]. They reported that in the metformin group, there was a reduction in the testosterone concentration and in the free-androgen index [14]. Salamalekis et al., suggested that there is no association between insulin levels or IR levels and the development of OHSS in women, with or without PCOS, undergoing ART. The fasting glucose/insulin ratio (FGIR) was used for the evaluation of insulin resistance (IR) in their study [19]. One possible explanation for the observed discrepancy is the difference in IR measurement methods while in our study the HOMA-IR test was used. Considering simple surrogates for HOMA-IR are among the best and most extensively validated, they are probably more reliable than FGIR [20, 21]. The other explanation is the difference in sample size, while our sample size was 1.5 times that of the aforementioned study [19]. Moreover, the age and the number of retrieved oocytes were not retained as independent risk factors in the multivariable logistic regression analysis. Several studies evaluate the impact of BMI on the development of OHSS and have reported contradictory results [2, 22, 23]. In contrast to our finding, body weight/BMI did not appear to be a useful marker for predicting risk of OHSS according to the study of Fiedler et al., [22]. PCOM is a criterion for identifying PCOS but not necessarily predicting severity or presence of endocrine dysfunction [11, 24]. However, we found that PCOM is a risk factor for OHSS in multivariable analysis. In addition, in the current study, other risk factors including BMI, HOMA-IR, FAI, and preovulatory follicles were identified as a risk factor for OHSS.

In our study, ROC analysis revealed that HOMA-IR, FAI, AFC, AMH, and preovulatory follicles could be good markers to predict the OHSS’ risk in non-PCOS women undergoing controlled ovarian hyperstimulation. The best thresholds for HOMA-IR, FAI, AFC, AMH, and preovulatory follicles in patients were 2.36, 3.9, 8, 3.3 ng/ml, and 10, respectively. Similar to our study, Ocal et al. have shown that the cut-off value of AMH of 3.3 ng/ml had a sensitivity and specificity of 90% and 71%, respectively, for predicting OHSS [25]. Also, Nardo et al. showed a cut-off value for AMH 3.5 ng/ml with a sensitivity and specificity of 88% and 70%, respectively, in both PCOS and non-PCOS together [26]. In contrast to our study, Vembu et al. described the cut-off value for AMH 4.85 ng/ml in non-PCOS group provided a sensitivity of 85.7% and a specificity of 89.7% [27], which is higher than the value reported in our study. This could be attributed to a different assay. Dickerson et al., found a greater total number of follicles at the end of ovarian stimulation during IVF in patients with greatest IR, suggesting that the relatively higher level of insulin stimulates follicle recruitment or development. They suggested a positive correlation of HOMA-IR levels above a level of 2.5 and a continuous positive correlation of FAI to total ovarian follicle count following medically-assisted reproduction (MAR) in the non-PCOS women and they assessed the effect of IR and androgen status in non-PCOS women on the follicular outcome of MAR [8]. In consistent with our study, the number of follicles on the day of hCG administration appears to be a good prognostic indicator for the occurrence of OHSS in women undergoing ART [28]. Hence, it could be hypothesized that HOMA-IR and FAI status in non-PCOS women are good markers for predicting ovarian response, including an exaggerated one (i.e., OHSS). Surprisingly, we noticed that the threshold level of HOMA-IR above which the OHSS could be predicted with good sensitivity and specificity (i.e., IR > 2.36) was very close to the level above which a good ovarian response (i.e., a greater total number of follicles) could be expected (i.e., IR > 2.5) [8]. The cut-off point used for defining biochemical hyperandrogenemia in the diagnosis of PCOS is an FAI value of 8.5 [29]. Our result demonstrated that the effect of androgen on ovarian response starts from much lower values of FAI (i.e., 2.25–3.9) than those are used to delineate classical hyperandrogenemia.

The same pattern is described by Dickerson et al., as they showed a continuum of effect of androgen on ovarian response, even in those not classically demonstrated to have hyperandrogenemia [8]. As OHSS treatment is largely supportive, prevention is crucial. So it seems logical to assess IR and androgen status even in non-PCOS women and take preventive measures including administration of insulin sensitizing agents or use of GnRH antagonist cycles and triggering ovulation by GnRH agonist in ART cycles.

This study has some limitations. No GnRH antagonist cycles were in group ART and participants were Caucasian, so our results cannot be extrapolated to other ethnic groups.

In conclusion, patients with a higher value of BMI, FAI, HOMA-IR, and preovulatory follicles and the presence of PCOM are more likely to develop OHSS, which are not confined to PCOS patients.

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Conflict of Interest

The authors declare that they have no conflict of interest.

References


