Antinociceptive and Anti-Inflammatory Effects of Bixin, a Carotenoid Extracted from the Seeds of *Bixa orellana*

**Authors**
Samanta Daliana Golin Pacheco 1*, Alexia Thamara Gasparin 2*, Carlos Henrique Alves Jesus 2, Bruna Bittencourt Sotomaia 2, Ana Clara Sans Salomão Brunow Ventura 2, Daiany Darly Belo Redivo 2, Daniela de Almeida Cabrini 2, Josiane de Fátima Gaspari Dias 1, Marilis Dallarmi Miguel 1, Obdulio Gomes Miguel 1, Joice Maria da Cunha 2

**Affiliations**
1 Postgraduate Program in Pharmaceutical Sciences, Federal University of Paraná, Curitiba, Parana, Brazil
2 Laboratory of Pharmacology of Pain, Department of Pharmacology, Federal University of Paraná, Curitiba, Parana, Brazil

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**Correspondence**
Prof. Dr. Joice Maria da Cunha
Department of Pharmacology, Biological Sciences Building, Federal University of Paraná
R. Coronel Francisco H dos Santos S/N, P.O. Box 19031, 81531-980 Curitiba, Paraná, Brazil
Phone: + 55 41 33 61 17 20, Fax: + 55 41 32 66 20 42
joice.cunha@ufpr.br

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**Introduction**

Bixin (▶ Fig. 1) is a major liposoluble diapocarotenoid extracted from the seeds of *Bixa orellana* Linné (Bixaceae). Among the natural carotenoids, bixin stands out chemically for presenting the cis conformation, unusual among the carotenoids, and for presenting a carboxylic group and a methyl ester in its chemical structure, which confers fat solubility to the molecule [1, 2]. Bixin is widely used as an FDA-approved food colorant and additive, as well as a cosmetic and textile colorant. It is considered the main pigment of the seeds of annatto, the common name of the species *B. orellana*, being the greater responsible for conferring the reddish-orange tonality, characteristic of seeds [3, 4]. Among the several plants that have these compounds and high colorant potential, annatto is one of the most economically important plants [5].

* Samanta Daliana Golin Pacheco and Alexia Thamara Gasparin made equal contributions to this study.
In Mexico and South America, B. orellana has been traditionally used to treat infectious and inflammatory diseases of the skin, prostate, gastrointestinal tract, and chest pain [6]. In fact, several in vitro or in vivo studies have already demonstrated varied biological properties of B. orellana extracts or fractions, such as antibacterial, antifungal, antioxidant, and antimalarial [1]. Importantly for this study, Benoit et al., (1976) [7] described a significant reduction of carrageenan-induced paw edema in rats after treatment with ethanolic extract of B. orellana. Additionally, treatment with crude aqueous extract of B. orellana leaves exhibited significant inhibitory activity against bradykinin-induced inflammation [8]. Considering the nociception, preliminary data obtained by Shilpi et al. (2006) [9] have already demonstrated that methanol extract of B. orellana leaves significantly and dose-dependently reduced the acetic acid-induced writhing response in mice.

Although bixin is the main carotenoid of B. orellana and that carotenoids are known compounds with several pharmacological properties [10, 11], few studies evaluate the effects of isolated bixin. It has been already shown its protective effect on cells and tissues with antioxidant, antitymoma, antigenotoxicity, and anticlastogenicity actions, being considered a biological neutralizer of reactive oxygen species [12]. In addition, studies have demonstrated the anti-inflammatory activity of bixin through the antioxidant transcription factor Nrf2 activation [4, 13], and its efficiency in accelerating wound healing as well as reducing the scar tissue area [14]. Although pain is one of the cardinal signs of inflammation, to our knowledge there are no studies evaluating the antinociceptive potential of bixin. Thus, besides validating its anti-inflammatory activity, this study aimed to evaluate antinociceptive activities of bixin in murine models of inflammatory pain.

**Results**

The ▶ Fig. 2 (panel A) shows the effect of oral treatment with bixin (at doses of 15 or 30 mg/kg) or vehicle (corn oil) on carrageenan (Cg) induced edema in rats. Two-way analysis of variance (ANOVA) showed a significant effect on experimental groups [F(4, 35) = 16.74; p < 0.05] and time [F(4, 140) = 146.0; p < 0.05], besides an interaction between these factors [F(16, 231) = 28.28; p < 0.05]. The post hoc test of Bonferroni showed a significant difference between all groups that received Cg injection compared to the VEH/SAL-treated group (p < 0.05). Therefore, the injection of Cg causes an increase in MPO activity compared to the SAL group. Besides, the Bonferroni’s test showed a significant difference between bixin/Cg, at a dose of 30 mg/kg, and dexamethasone/Cg-treated groups, in comparison to the vehicle/Cg-treated group (p < 0.05). Thus, the treatment with bixin, at the dose of 30 mg/kg or with dexamethasone was able to significantly reduce the MPO activity in comparison to the VEH/Cg group (p < 0.05). No significant difference was observed between the bixin (30 mg/kg) and dexamethasone/Cg-treated groups (p > 0.05).

The ▶ Fig. 4 (panel A) demonstrates the cumulative number of paw flinches induced by formalin injection in rats treated with bixin (at doses of 15 or 30 mg/kg) or vehicle (corn oil). Two-way ANOVA with repeated measures showed a significant effect on experimental groups [F(2, 21) = 99.37; p < 0.05] and time [F(11, 231) = 314.4; p < 0.05], besides an interaction between these factors [F(16, 231) = 8.669; p < 0.05]. The post hoc test of Bonferroni showed a significant difference between all groups that received Cg when compared with the vehicle (VEH)/saline (SAL)-treated group (p < 0.05), demonstrating the Cg-induced increase in paw edema when compared to the SAL group peaked 2 and 3 h after Cg injection. Besides, the Bonferroni’s test showed that the treatment with bixin (30 mg/kg) significantly attenuated the paw edema at first and second hour after Cg injection (p < 0.05). The paw edema in dexamethasone/Cg-treated group was significantly different in comparison with vehicle/Cg-treated group, 2, 3, and 4 h after Cg injection (p < 0.05).

Analyzing the area under the curve (AUC) of total edema during the 4 h after Cg treatment (▶ Fig. 2, panel B), 1-way ANOVA showed a significant effect on experimental groups [F(4, 35) = 15.87; p < 0.05]. The post hoc test of Bonferroni showed that all experimental groups treated with Cg were significantly different when compared to the saline-treated group (p < 0.05). Additionally, this test demonstrated that the experimental groups treated with bixin (dose of 30 mg/kg) or dexamethasone were statistically different from the vehicle/Cg-treated group (p < 0.05), but not different from each other (p > 0.05).

▶ Fig. 3 shows the effect of oral treatment with bixin (at doses of 15 or 30 mg/kg) or vehicle (corn oil) on the myeloperoxidase (MPO) activity. One-way ANOVA showed a significant effect on experimental groups [F(4, 30) = 14.92; p < 0.05]. The post hoc test of Bonferroni showed a significant difference between all groups that received Cg injection in comparison to the VEH/SAL-treated group (p < 0.05). The post hoc test of Bonferroni showed a significant difference between bixin (at doses of 15 or 30 mg/kg) and dexamethasone/Cg-treated groups, in comparison to the vehicle/Cg-treated group (p < 0.05). Thus, the treatment with bixin, at the dose of 30 mg/kg or with dexamethasone was able to significantly reduce the MPO activity in comparison to the VEH/Cg group (p < 0.05). No significant difference was observed between the bixin (30 mg/kg) and dexamethasone/Cg-treated groups (p > 0.05).

The ▶ Fig. 5 (panel A) and ▶ Fig. 5 (panel B) demonstrates the number of writhings induced by acid acetic in mice treated with bixin (at doses of 27 or 53 mg/kg) or vehicle (VEH; corn oil). Analyzing the time course of acetic acid-induced writhings (▶ Fig. 5, panel A),
2-way ANOVA showed a significant effect on experimental groups \( F(2, 28) = 31.87; p < 0.05 \) and time \( F(5, 140) = 375.9; p < 0.05 \), besides an interaction between these factors \( F(10, 140) = 17; p < 0.05 \). The Bonferroni post hoc analysis showed a significant difference between bixin (at doses of 27 and 53 mg/kg) and vehicle-treated group at 10, 15, 20, 25, and 30 min after acid acetic injection (\( p < 0.05 \)).

When analyzed the total writhing response (panel B), 1-way ANOVA showed the effect of the experimental groups \( F(2, 28) = 31.31; p < 0.05 \) factor. The post hoc test of Bonferroni showed that bixin treatment at both tested doses significantly reduced the number of acetic acid-induced writhings when compared to the VEH-treated group (\( p < 0.05 \)). Furthermore, the bixin doses also differ statistically from each other (\( p < 0.05 \)).

The \( \text{Fig. 6} \) (panel A) demonstrates the effect of bixin (at doses of 15 or 30 mg/kg) or vehicle (VEH; corn oil) on the latency time in the hot plate apparatus. One-way ANOVA showed a significant effect of the experimental groups \( F(2, 27) = 4.061; p < 0.05 \) factor. The post hoc test of Bonferroni showed that bixin treatment at both tested doses significantly reduced the number of acetic acid-induced writhings when compared to the VEH-treated group (\( p < 0.05 \)). Furthermore, the bixin doses also differ statistically from each other (\( p < 0.05 \)).

\( \text{Fig. 6} \) (panel B) shows the effect of bixin treatment (at doses of 15 or 30 mg/kg) or vehicle (VEH; corn oil) on the number of crossings at the open field apparatus. One-way ANOVA show no significant effect on experimental groups \( F(2, 16) = 0.3298; p > 0.05 \).

Discussion

In the current study, the anti-inflammatory potential of bixin was validated using preclinical models of acute inflammation. First, we demonstrated that oral treatment with bixin was able to prevent the development of carrageenan-induced paw edema in rats, which seems to be related to the inhibition of neutrophil migration to the inflammation site. Besides, we observed, to our knowledge, for the first time in the literature, the antinociceptive effect of acute treatment with bixin using preclinical models of thermal and chemical nociception in rats and mice. This effect does not seem to be associated with sedative effects since the bixin treatment did not change the locomotor performance in the open field test. Additionally, the ability of bixin to attenuate nociceptive re-
Responses induced by multiple stimuli (chemical and thermal) suggests possible participation of the molecule in peripheral and central anti-nociceptive mechanisms. This broad distribution of bixin is corroborated by previous findings in the literature which indicate that as a nonpolar substance, bixin reaches the systemic circulation (and therefore is detectable in plasma) 1 h later after oral administration in an oily vehicle [15, 16].

Interestingly, previous studies have already demonstrated that *B. orellana* extracts or fractions significantly attenuate the inflammatory response induced by diverse stimuli [8, 9]. When considering studies with isolated bixin, few *in vivo* and *in vitro* studies have also demonstrated the ability of bixin to reduce inflammation [13, 17, 18]. In our study, oral treatment with bixin promotes a significant reduction of paw edema in the first and second hour after administration of carrageenan. As previously described, paw edema induced by carrageenan produces a biphasic response [19] where-in the first phase (ranging from 0 to 60 min after administration of carrageenan) is characterized by the release of substances such as histamine, serotonin and bradykinin while the second phase (from 1, 2, and 3 h) is characterized by an increased production and release of prostaglandins (PGs), as well as reactive species of oxygen from migratory neutrophils [19, 20]. Considering this fact, our next experiment aimed to investigate the activity of MPO, an enzyme released essentially by activated neutrophils, in the model of...
carrageenan-induced inflammation. It was observed that bixin treatment significantly decreased the MPO activity in carrageenan-inflamed skin samples, suggestive of a lower infiltration of leukocytes to the injured tissue. Although not investigated in our study, previous work has already shown that bixin significantly decreases the levels of inflammatory markers such as interleukin 6 and tumor necrosis factor alpha [21] and reduces the in vitro activity of both cyclooxygenase isoforms activity, reducing the levels of PGs [17]. This anti-inflammatory effect of bixin has been also associated to its agonist action on peroxisome proliferator-activated receptor (PPAR) alpha and PPAR gamma, which, among other actions, promote the reduction of inflammatory cytokines and inhibition of macrophage activation [18]. Finally, it has been described some cytoprotective effects of bixin due to its activation of the transcription factor NRF2 (nuclear factor-E2-related factor 2) [22], which regulates the expression of numerous antioxidants, anti-inflammatory, and pro-survival genes [15, 17, 22].

Since pain is one of the cardinal signs of inflammation, our next experiments were designed to characterize an antinociceptive potential of bixin, an effect still unheard of in the literature. We first start testing the effect of bixin on the nociceptive responses induced by formalin. The formalin test is characterized by 2 distinct phases of nociceptive behavior [23]. The first phase, called neurogenic, begins shortly after the formalin injection, remaining until 3/5 min, and occurs due to chemical stimulation of the nociceptors by formaldehyde [24]. After occurs a quiescent period (6–15 min), and then the second phase (15–60 min) begins. This second phase, called inflammatory, is characterized by the return of nociceptive behaviors and the involvement of peripheral inflammatory mediators, which sensitize primary and spinal sensory neurons, triggering the activation of nociceptors [24].

The different properties of both phases allow this test to be widely used as a tool to indicate possible mechanisms of action for drugs being tested in regards a peripheral and/or a central mechanism of action [25]. In our study, bixin treatment (at both tested doses) induces a significant decrease of formalin-induced flinches during phases I and II of the formalin test, but not during the quiescent interphase, corroborating its anti-inflammatory activity and suggesting that bixin exerts its antinociceptive effect acting peripherally and centrally.

To confirm this, we also test the effect of bixin on the acetic acid-induced writhing in mice. It has been well described that the i. p. administration of acetic acid induces hyperalgesia by promoting the release of noxious endogenous substances, such as cytokines, PGs, substance P, and bradykinin, which are responsible for sensitization of nociceptors’ nerve endings [26, 27]. After injury stimulation, large amounts of various PGs are produced by polymorphonuclear cells, especially neutrophils, enhancing biosynthesis and release thereof into the peritoneal cavity [28, 29]. As mentioned, to our knowledge, there are no studies involving nociception using isolated bixin. However, Shilpi et al. [9] have also observed a decrease in the number of acetic acid-induced abdominal writhes after the treatment with the foliar methanolic extract of B. orellana. Drugs with anti-inflammatory and antioxidant properties have been described as being effective in decreasing nociceptive behaviors and PGs levels in the peritoneal cavity in this nociception test. Although not thoroughly investigated in the present study, the antinociceptive effect of bixin observed in this test is possibly due to its anti-inflammatory and antioxidant properties [28], its capacity to negatively modulate PG production by cyclooxygenase (COX) inhibition [17, 30], and its ability to reduce neutrophil migration (the reduction of MPO activity). In contrast to previous findings, in this test, the effect of bixin was not dose-related (i.e., the lower dose of bixin significantly reduced the total number of writhes performed during the 30 min when compared to the higher dose). This fact may be attributed to physiological and interspecific differences between the rats and mice since the acetic acid-induced abdominal writhing test was the only one conducted in Swiss albino mice in this study. Corroborating this hypothesis, the previous study from Pinzon-Garcia et al. [31] has also not observed a dose-response effect of bixin treatment on wound healing using Swiss mice as an animal model.

To confirm that the antinociceptive effect of bixin involves central nervous system (CNS)-mediated mechanisms, in our next ex-
periment, the effect of bixin was investigated on the latency to
noxious thermal stimulus in the hot plate test. It has been well
characterized that drugs that act exclusively by their peripheral
actions, such as COX inhibitors, do not significantly alter the be-
havioral responses in this test, making this a widely used model
to evaluate drugs with potential action on the CNS [32]. Bixin
treatment significantly increased the latency to the noxious ther-
mal stimulus in the hot plate, demonstrating that central mecha-
nisms may contribute to the antinociceptive effect exerted by
bixin (at least at the higher dose). Especially because of this pos-
sible effect on CNS and in studies involving nociceptive behaviors
that depend on the display of active motor behaviors, it is ex-
tremely important to rule out the possibility that compounds
have sedative effects. To elucidate whether the treatment with
bixin promotes any locomotor deficit, the open field test was per-
formed. As shown, the treatment with bixin (at both tested dose)
did not alter the number of crossings in the open field test, ex-
cluding the sedative effect as a contributor to the antinociceptive
responses observed in the formalin and hot plate tests. Curiously,
Shilpi et al. [9] reported that the methanolic extract of the leaves
of B. orellana promoted a decrease in locomotion in this behav-
ioral test, which was not observed using the isolated bixin.

In this study, we demonstrated the anti-inflammatory property
of bixin. It seems to be due to its capacity of bixin to reduce the
neutrophil migration to the inflammatory site. Furthermore, this
is the first report showing the antinociceptive property of bixin,
which does not appear to be related to the sedative effect but is
associated with both peripheral and central actions. Further stud-
ies are necessary to characterize the mechanisms involved in
these effects.

The \(^1\)H and \(^{13}\)C NMR spectra and the absorption at the maxi-

Material and Methods

Plant material

B. orellana (annatto) seeds were collected at the Instituto Ambien-
tal do Paraná (IAP) in Morretes, Paraná, Brazil, in June and July
2015 (coordinates: COD 02548038/25°30’S, 48°49’W; altitude:
59 m). The plant material was identified by Osmar dos Santos
Ribas from Municipal Botanical Museum of Curitiba, where a
voucher specimen (#379394) was deposited. Permission to eval-
uate the bioactivities of the extracts from Brazilian plants was
granted by the Instituto Brasileiro do Meio Ambiente e dos Recur-

Isolation of bixin

The seeds (1000 g ± 0.1) were dried at 50°C, pulverized, and
passed through a 35-mesh sieve. The powdered material was ex-
tracted in a modified Soxhlet apparatus using increasing polarity
solvents (1 : 10; w/v) (hexane, chloroform, ethyl acetate, and etha-
nol) for 6 h and subsequently filtered through Whatman No. 41 fil-
ter paper. The red-purple powder of bixin (yield: 5.1%) was ob-
tained by extracting chloroform with hexane from the previously
defatted seeds [14, 33]. The polarity gradient was performed to
certify that bixin would not be present in other solvents such as
ethyl acetate and ethanol. The material was oven-dried and stored
at −16°C. The measured melting point of bixin (195°C) was sim-
ilar to previously described in the literature [3, 14, 34].

Bixin identification

The bixin identification was supported by nuclear magnetic reso-
nance spectroscopy (NMR; \(^1\)H and \(^{13}\)C; Fig. 15 and Fig. 25, Sup-
porting Information) as well as by UV (Fig. 3S, Supporting Infor-
mation) and IR [3, 35]. The purity of bixin was estimated to be
greater than 99%. In agreement with the literature [3, 36, 37],
our results showed UVλ nm: 489, 462, 432.7. IR \(^\nu\) (KBr) cm-
11716.64, 1660, 1385, 1300, and 900. 1H NMR (600 M
Hz. DMSO-d6), δ 7.89 (1H, d, J = 15.5 Hz, H-7), δ 7.26 (1H, d,
J = 15.5 Hz, H-7’), 6.45–6.87 (10 H, m, 10 x: CH), 5.83 (1H, d,
J = 15.5 Hz, H-8), 5.94 (1H, d, J = 15.5 Hz, H-8’), 3.70 (3H, s,
OMe), 1.92–1.99 (12 H, m, 4 x: CMe).

In vivo experiments

Animals

Adult male Wistar rats (180–220 g) and Swiss albino mice (18–
35 g), supplied by the Federal University of Parana colony, were
used in this study. Animals were housed in plastic cages
(41 × 32 × 16.5 cm) and maintained in standard conditions of
room temperature (21 ± 2°C) and illumination cycle (12-h light/
dark) with food and water provided ad libitum. Bixin (15 or
30 mg/kg in rats; 27 or 53 mg/kg in mice, calculated according
to its basal metabolic rate, using the method proposed by Freitas
and Carregarro [38]) or vehicle (1 mL/kg; corn oil) was adminis-
tered orally by gavage after 12-h fasting. Animals were habituated
to the experimental room for at least 1 h before the experiments.
The study was conducted following the National Institutes of
Health Guide for the Care and Use of Laboratory Animals and ap-
proved by the Federal University of Parana Institutional Commit-
tee on the Ethical Use of Animals (CEUA/BIO-UFPR; authorization
#1087, approved on August 15, 2017). All efforts were made to
minimize the number of animals, following the reduction princi-
ples recommended by Russell and Burch [39]. For this reason,
pharmacological positive control groups were not conducted for
behavioral experiments related to the potential antinociceptive
effect of bixin since the antinociceptive effect of drugs such as
non steroidal anti-inflammatory drugs (NSAIDs) or opioids has
been extensively observed using the same animal models [40, 41].

Carrageenan-induced paw edema and measurement
of the MPO activity

To evaluate the potential anti-inflammatory effect of bixin, the
paw edema was induced by intra-plantar injection of carrageenan
(Cg; Sigma-Aldrich; purity: approximately 52%), at the dose of
200 µg/paw in 0.1 mL of saline, according to previously described
by Hirota et al. [42]. Briefly, 4 experimental groups were designed
(n = 8 rats/each): negative control group orally (p. o.) treated with
corn oil (bixin vehicle; equivalent volume); bixin-treated groups
(15 or 30 mg/kg; p. o.); and positive control treated with dexa-
methasone (1 mg/kg, subcutaneous injection). All treatments
were administered 1 h before the injection of carrageenan. Bixin
doses were selected based on previous studies [4, 13, 14]. As a
control, contralateral paws received saline (Cg vehicle; 0.1 mL). The paw thickness was evaluated before (basal measurement) and again 1, 2, 3, and 4 h after Cg or saline (0.1 mL/paw) injections, using a digital pachymeter and expressed in millimeters (mm).

In another set of experiments, all the above experimental groups and procedures were repeated. However, 3 h after Cg (peak of edema) or saline injections, rats were euthanized and segments of the sub-plantar region of both hind paws were collected, weighed and stored at −80°C. The MPO activity was determined according to the methodology described by De Young et al. [43] with modifications. In brief, tissue samples were homogenized in 1.5 mL of sodium phosphate buffer (80 mM, 0.5% hexadecyl trimethylammonium bromide [HTAB], pH 5.4) for 15 s at 0°C. The homogenate was then centrifuged at 11,200 g at 4°C for 20 min. Then triplicates of 30 µL supernatant were transferred to plates, which previously received 200 µL of peroxide solution (100 µL of 80 mM sodium phosphate buffer, 85 µL of 0.22 mM sodium phosphate buffer plus 15 µL of hydrogen peroxide 0.017%). The reaction was started with the addition of 20 µL of TMB solution (18.4 mM dissolved in 8% aqueous dimethylformamide). The plate was then transferred to the greenhouse for 3 min at 37°C and thereafter the reaction was stopped by the addition of 30 µL of sodium acetate (1.46 M) in each well. The enzymatic activity was evaluated by the colorimetric method using a plate reader (Bio-Tek Ultra Microplate reader EL808), with a wavelength of 620 nm. The results were expressed as optical density (OD)/total sample weight.

**Formalin test**

The potential antinociceptive effect of bixin was firstly evaluated in the formalin test according to Hirota et al. [42] with modifications. Briefly, 50 min after oral treatment with vehicle (corn oil; 1 mL/kg) or bixin (15 or 30 mg/kg), rats (n = 7–9 each group) were acclimated in the formalin test apparatus (an inverted glass funnels 290 mm wide and 410 mm high) for 10 min. Then animals received the formalin injection (2.5%, 50 µL/rat) into the dorsal surface of one of the hind paws. Flinches were scored immediately after formalin injection for 60 min, divided into 5-min periods with the phases defined as the following time intervals: phase I (0–5 min), quiescent phase (6–15 min), and phase II (16–60 min). Results were expressed as the cumulative number of flinches during the 60 min of the test and the sum of total flinches at each phase of the test.

**Acetic acid-induced writhing test**

The writhing test was performed following the procedures previously described by Hirota et al. [42]. For this, 50 min after oral treatment with vehicle (corn oil; equivalent volume; p.o.) or bixin (27 or 53 mg/kg; p.o.), mice (n = 10–11 each group) were placed to acclimate in an inverted glass funnel (290 mm wide and 410 mm high) for 10 min. The doses of bixin used in mice were calculated according to the general method of calculation for the allometric scale of drugs, based on the basal metabolic rate of the animals [32]. One hour after the vehicle or bixin treatment, each mouse was injected with acetic acid 0.6% (10 mL/kg; i.p. injection) and individually housed in the glass cylinder. The cumulative number of writhes (characterized by abdominal constriction and stretching of at least 1 hind limb) was scored for 30 min.

**Hot plate test**

The potential antinociceptive effect of bixin over an acute thermal stimulus (50 ± 1°C) was evaluated using a hot plate apparatus (Ugo Basile SRL), as previously described [44]. For this, rats (n = 6–7 each group) were divided into 3 different groups treated with vehicle (corn oil; 1 mL/kg; p.o.) or bixin (15 or 30 mg/kg; p.o.). The latency (in seconds) for animals to display behaviors such as licking of the fore and hind paws or jumping was measured before and 1 h after corn oil or bixin treatments. The cutoff time used to prevent skin damage was 25 s.

**Open field test**

The open field test was conducted to evaluate the effect of treatments over the spontaneous locomotor activity, according to Meotti et al. [45]. Briefly, 1 h after treatment with vehicle (corn oil; 1 mL/kg, p.o.) or bixin (15 or 30 mg/kg, p.o.) rats (n = 6–7 each group) were placed in the center of the open field apparatus (a rectangular wooden arena; 40 cm wide × 50 cm long × 63 cm high; divided into 9 rectangular units). The locomotor activity was video recorded, and the number of units crossed with all 4 paws was counted for 5 min.

**Statistical analysis**

The data were presented by the mean plus standard error of the mean (SEM) for 6 to 14 animals per group. Data were compared using 2-way ANOVA with repeated measures (time-course behavioral data, where the independent factors used were treatment and time) or 1-way ANOVA (column graphs). When appropriate, the post hoc analysis of Bonferroni was applied. The level of significance was established at p < 0.05. All the tests were carried out using the GraphPad Prism program (version 6).

**Supporting Information**

The 1H and 13C NMR spectra and the absorption at the maximum lambda in the UV region of bixin are available as Supporting Information.

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**Conflict of Interest**

The authors declare that they have no conflict of interest.

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