**Introduction**

The pituitary gland is the “maestro” of the body’s endocrine axes by regulating the secretion of most hormones from the other endocrine organs. The mature pituitary gland consists of the adenohypophysis (anterior and intermediate lobes) and the neurohypophysis or posterior lobe. The anterior pituitary consists of 5 distinct cell types producing 6 different hormones: somatotrophs, producing growth hormone (GH), thyrotrophs, producing thyroid stimulating hormone (TSH), lactotrophs, producing prolactin (PRL), gonadotrophs, producing follicle-stimulating hormone (FSH) and luteinizing hormone (LH) and corticotrophs, producing adrenocorticotropic hormone (ACTH). The intermediate lobe secretes pro-opiomelanocortin (POMC) which is the precursor of the melanocyte-stimulating hormone (MSH) and endorphins. The posterior lobe is responsible for storing and secreting antidiuretic hormone (ADH) and oxytocin, which are produced in the paraventricular and supraoptic nuclei of the hypothalamus.

The ontogenesis of the pituitary gland is a highly complex process that results from the interaction of transcription factors and signaling molecules. Congenital hypopituitarism is characterized by the deficiency in one or more pituitary hormones. The phenotype can be highly variable, consisting of isolated hypopituitarism or more complex disorders. The same phenotype can be attributed to different gene mutations; while a given gene mutation can induce different phenotypes. This review highlights the genetic variations that lead to congenital hypopituitarism and their associated defects. The overall incidence of mutations in known transcription factors in patients with hypopituitarism is low; therefore many gene mutations or even gene-epigenetic interactions have to be unraveled in the future to explain the vast majority of still unclear cases of congenital hypopituitarism.
Embryology of the Pituitary Gland (Based on Murine Models)

Human pituitary development is similar to the murine pituitary development and most of our knowledge is based on the murine animal model. The anterior and intermediate lobes are derived from the oral ectoderm, while the posterior pituitary is derived from the neural ectoderm of the ventral diencephalon. Its development occurs in 4 stages: (i) the pituitary placode, (ii) the rudimentary Rathke’s pouch, (iii) the definitive Rathke’s pouch and (iv) the mature pituitary gland. The placode appears ventrally in the midline of the anterior neural ridge at embryonic (E) day 7.5 giving rise to the roof of the oral cavity. The pituitary organogenesis starts with a thickening in this roof at E 8.5, corresponding to 4–6 weeks’ gestation in humans. At E9.0 it invaginates dorsally to form the rudimentary Rathke’s pouch. Rudimentary Rathke’s pouch deepens and folds in itself until it closes, forming the definite Rathke’s pouch at E11.5. The anterior and intermediate lobes are derived from Rathke’s pouch while the evagination of the posterior part of the presumptive diencephalon will give rise to the posterior pituitary. The pouch epithelium continues to proliferate between E10.5 and E12 and separates from the underlying oral ectoderm at E12.5. The progenitors of the hormone-secreting cell types proliferate ventrally from the pouch between E12.5 and 17.5 to populate the future anterior lobe [1] (▶ Fig. 1).

Syndromic Hypopituitarism

In this section cases of congenital hypopituitarism in the context of specific syndromes will be presented, as also summarized in ▶ Table 1.

Hypopituitarism with skeletal abnormalities (LHX3)

LHX3, a member of the LIM class of homeodomain proteins is expressed early during anterior pituitary development, within RP at E9.5 as well as in the ventral hindbrain and spinal cord. Its expression, which persists in the mature pituitary gland, is essential for the formation of gonadotrophs, thyrotrophs, somatotrophs, and lactotrophs. In humans, 14 homozygous [2–10], or compound heterozygous for 2 LHX3 mutations [11] and a heterozygous variant [12] have been reported so far. People with LHX3 mutations present with GH, TSH and FSH/LH deficiencies while ACTH deficiency is reported in 50% of the mutations. MRI images reveal different anomalies, as aplasia or hypoplasia of pituitary, a hypointensity resembling microadenoma [4] and enlargement with hyperintense signal [7]. In 10% of the cases MRI may be normal. Mutations that affect the carboxyl terminus of LHX3 proteins present with phenotypes of pituitary dysfunction only, while mutations affecting the entire gene or protein, the LIM domains or the homeodomain present with syndromes involving besides the endocrine also the nervous and skeletal systems. Extra-pituitary phenotype can include short neck with abnormal head and neck rotation (70% of cases), vertebral abnormalities (50% of cases), as rigid cervical spine, flattened lumbar vertebrae, thoracic kyphosis, progressive scoliosis and mild to severe hearing defect (50% of cases), due to the expression of LHX3 in defined regions of the sensory epithelium of the developing inner ear, which is known to be required for inner ear development. In 38% of the cases mental/learning deficiencies are reported. Two of the reported patients had also respiratory distress [11, 13]. Heterozygous family members are unaffected.

▶ Table 1

<table>
<thead>
<tr>
<th>Oral ectoderm</th>
<th>Rudimentary pouch</th>
<th>Definitive pouch</th>
<th>Pituitary gland</th>
</tr>
</thead>
<tbody>
<tr>
<td>Rat</td>
<td>8.5</td>
<td>11</td>
<td>14.5</td>
</tr>
<tr>
<td>Mouse</td>
<td>8 – 8.5</td>
<td>9.5</td>
<td>12</td>
</tr>
<tr>
<td>Embryonic days</td>
<td></td>
<td>19.5</td>
<td>17</td>
</tr>
</tbody>
</table>

▶ Fig. 1 Stages of rodent pituitary development. a Oral ectoderm, b Rudimentary pouch; c Definitive pouch; d Adult pituitary gland. I: Infundibulum; NP: Neural plate; N: Notochord; PP: Pituitary placode; OM: Oral membrane; H: Heart; F: Forebrain; MB: Midbrain; HB: Hindbrain; RP: Rathke’s pouch; AN: Anterior neural pore; O: Oral cavity; PL: Posterior lobe; OC: Optic chiasm; P: Pontine flexure; PO: Pons; IL: Intermediate lobe; AL: Anterior lobe; DI: Diencephalon; SC: Sphenoid cartilage. Source: Sheng and Westphal (Trends Genet 1999; 15: 236–240) with permission (licence number: 4230341362009). [ref].
Hypopituitarism with cerebellar abnormalities and/or lung defects (LHX4)

Closely related to LHX3 is LHX4, which can be detected in Rathke’s pouch at E9.5 and in the anterior part of the pituitary at E12.5. LHX4 mutations result in the formation of a hypoplastic pituitary, containing all differentiated cell types but in reduced numbers. A lack of proliferation in Lhx4 mutants causes failure to respond to inductive signals and subsequent dysregulation of other transcription factor genes, for example, Lhx3, inevitably leading to increased cell death [14]. In humans, 15 sporadic or familiar LHX4 mutations have been reported with variable hypopituitarism (ranging from isolated GH deficiency to complete panhypopituitarism) [15] and brain abnormalities (pituitary hypoplasia, ectopic posterior pituitary and poorly developed sella turcica as well as corpus callosum hypoplasia or Chiari syndrome) [12, 16–19]. Of note, 4 patients also presented respiratory distress [20–22] and one presented heart defect [21]. A recent reported homozygous mutation has been associated with a lethal phenotype (severe panhypopituitarism associated with pituitary aplasia and posterior ectopia, mild facial hypoplasia, undescended testes and respiratory distress) [22]. In a very large cohort of 417 unrelated patients with hypopituitarism the prevalence of LHX4 mutations was 1.4% [23].

<table>
<thead>
<tr>
<th>Transcription factor</th>
<th>Transmission</th>
<th>Pituitary hormone deficiencies</th>
<th>Phenotype</th>
</tr>
</thead>
<tbody>
<tr>
<td>LHX3</td>
<td>AR</td>
<td>GH, TSH and FSH/LH, ACTH (53%)</td>
<td>Abnormal head and neck rotation (70%), vertebral abnormalities (50%), mild to severe hearing defect, pituitary hypo- or hyperplasia</td>
</tr>
<tr>
<td>LHX4</td>
<td>AD</td>
<td>IGHD or CPHD</td>
<td>Pituitary hypoplasia, ectopic posterior pituitary and small sella turcica, corpus callosum hypoplasia or Chiari syndrome. Respiratory distress</td>
</tr>
<tr>
<td>HESX1</td>
<td>AD/AR</td>
<td>IGHD or CPHD</td>
<td>SOD, hypoplastic or aplastic anterior pituitary, corpus callosum hypoplasia</td>
</tr>
<tr>
<td>SOX2</td>
<td>AR</td>
<td>LH, FSH, variable GHD</td>
<td>Microphthalmia, mental retardation, sensorineural hearing loss, esophageal atresia</td>
</tr>
<tr>
<td>SOX3</td>
<td>X-linked</td>
<td>IGHD or CPHD</td>
<td>Anterior pituitary hypoplasia, absent pituitary stalk, ectopic neurohypophysis, intellectual disability</td>
</tr>
<tr>
<td>OTX2</td>
<td>AD</td>
<td>IGHD or CPHD</td>
<td>Eye malformations, normal or hypoplastic anterior pituitary, normal or ectopic posterior pituitary, Chiari syndrome</td>
</tr>
<tr>
<td>PAX6</td>
<td>AR</td>
<td>GH, ACTH (?) , FSH, LH</td>
<td>Eye malformations, pituitary hypoplasia</td>
</tr>
<tr>
<td>BMP4</td>
<td>AR</td>
<td>CPHD</td>
<td>Ocular malformations, sensorineural deafness, developmental delay, spondyloepiphyseal dysplasia tarda, cerebellar, pituitary abnormalities, partial callosal agenesis</td>
</tr>
<tr>
<td>ARNT2</td>
<td>AR</td>
<td>CI, CPHD</td>
<td>Eye anomalies, hypoplastic anterior pituitary and thin pituitary stalk, microcephaly, renal anomalies</td>
</tr>
<tr>
<td>FGFR1</td>
<td>AD</td>
<td>CPHD</td>
<td>Absent corpus callosum, septo-optic dysplasia and midline defects</td>
</tr>
<tr>
<td>SHH</td>
<td>AR</td>
<td>CPHD , DI</td>
<td>Midline cerebral defects, holoprosencephaly (HPE)</td>
</tr>
<tr>
<td>Gli2</td>
<td>haploinsufficiency</td>
<td>CPHD</td>
<td>HPE, polydactyly, midline defects, ectopic pituitary lobe</td>
</tr>
<tr>
<td>PROKR2</td>
<td>AR/AD</td>
<td>FSH, LH</td>
<td>PSIS, corpus callosum hypoplasia, optic nerve anomalies, CI</td>
</tr>
<tr>
<td>GPR161</td>
<td>AR</td>
<td>GH</td>
<td>PSIS, anterior pituitary hypoplasia, ectopic posterior pituitary, empty sella, congenital alopecia</td>
</tr>
<tr>
<td>IGFB1</td>
<td>X-linked</td>
<td>TSH, GH, PRL</td>
<td>Macro-orchidism, delayed adrenarche</td>
</tr>
<tr>
<td>Pix2</td>
<td>AD/AR</td>
<td>ACTH, GH, TSH</td>
<td>Axenfeld-Rieger syndrome</td>
</tr>
<tr>
<td>NFkB2</td>
<td>AD</td>
<td>ACTH, GH, TSH</td>
<td>Immune deficiency</td>
</tr>
<tr>
<td>CHD7</td>
<td>AD</td>
<td>GH, TSH, FSH, LH</td>
<td>CHARGE syndrome, ectopic posterior pituitary</td>
</tr>
<tr>
<td>Piti1/POU1F1</td>
<td>AD/AR</td>
<td>GH, TSH, PRL</td>
<td>Pituitary hypoplasia</td>
</tr>
<tr>
<td>Prop1</td>
<td>AR</td>
<td>GH, TSH, PRL, LH, FSH, evolving ACTH</td>
<td>Pituitary hypoplasia or transient hyperplasia</td>
</tr>
</tbody>
</table>

AR: Autosomal recessive; AD: Autosomal dominant.
Septo-optic dysplasia and other eye abnormalities

Septo-optic dysplasia (SOD) is defined by the presence of at least 2 of the following: (i) optic nerve hypoplasia (ONH), (ii) midline forebrain defects and (iii) pituitary hypoplasia. Rarely, the eye abnormalities may be more severe including anophthalmia or severe microphthalmia. Neurological manifestations are common and range from focal deficits to global developmental delay. Mutations in HESX1, SOX2, SOX3, OTX2 have all been implicated in the etiology of SOD.

HESX1, a member of the paired-like class of homeobox genes, can be detected in the anterior forebrain from E7.5 to E8.5 and in the Rathke’s pouch from E8.5 to E13.5, after which period it is rapidly down regulated. It regulates the expression of other genes such as PROP1, suggesting that both function as opposing transcription factors. Initial activation of HEX1 is dependent upon LHX1 and LHX3. Mice homozygous for Hesx1 present with anophthalmia or microphthalmia and midline neurological defects (e.g., absent septum pellucidum and pituitary hypoplasia), reminiscent of SOD. The first homozygous missense mutation [24] was detected in 2 siblings born to consanguineous parents with SOD, agenesis of the corpus callosum, optic nerve hypoplasia, a hypoplastic anterior pituitary gland and complete panhypopituitarism. To date, several homozygous and heterozygous HESX1 mutations have been reported with variable phenotypes ranging from IGHD to CPHD associated in some cases with anomalies such as SOD and pituitary malformations [25–30]. MRI also reveals variable abnormalities, as hypoplastic or aplastic anterior pituitary and ectopic posterior pituitary. The patients carrying mutations at the heterozygous state generally show a milder phenotype than the homozygous patients. Most cases are sporadic.

SOX2 and SOX3 are members of the SOX (SRY-related high mobility group box) family of transcription factors.

SOX2 is expressed within the Rathke’s pouch at 4.5–9 weeks of pituitary development and maintained throughout the anterior pituitary development as well as in the diencephalon. It plays critical role in normal development of the brain and the pituitary gland in addition to that of the eyes and inner ear. In humans, SOX2 variants have been observed in 14 patients with hypogonadotropic hypogonadism, bilateral microphthalmia, small corpus callosum, hippocampal abnormalities and inconstant mental retardation [31–35]. Pituitary phenotype usually includes inconstant GH [36], TSH or ACTH deficiencies. On MRI of the hypothalamus-pituitary region, morphologic anomalies include hippocampal abnormalities, hypoplasia of the corpus callosum, hypothalamic hamartoma-ta and pituitary tumors [37]. All reported patients demonstrated learning difficulties, 4 of them had marked motor deficits and one had evidence of sensorineural deafness. Extra-pituitary anomalies included esophageal atresia.

SOX3, located on Xp27.1 is expressed throughout the central nervous system, particularly in the ventral diencephalon. SOX3 contains a DNA-binding domain conserved among SOX proteins, a short N-terminal domain, and a longer C-terminal domain containing 4 polyalanine (PA) tracts shown to be involved in transcriptional activation. SOX3 dosage is critical for normal hypothalamic-pituitary development, and both duplications and loss of function PA expansions can lead to variable hypopituitarism. Patients with duplication of SOX3 present with IGH without the involvement of additional adenohypophyseal hormones, more frequently with respect to patients with SOX3 sequence variants [38]. Anterior pituitary hypoplasia, an absent pituitary stalk, and ectopic neurohypophysis are other findings due to SOX3 sequence variants or whole gene deletions/duplications. Intellectual disability is frequently reported in these patients. The degree of mental retardation and the characteristics vary among patients [39, 40].

Orthodenticle homeobox 2 (OTX2) is a transcription factor gene that plays a critical role in brain and ocular development. OTX2 is expressed from E10.5 to E14.5 in the ventral diencephalon, where it likely interacts with HESX1 and from E10.5 to E12.5 in Rathke’s pouch. OTX2 regulates various transcription factors including RX1, PAX6, SIX3, LHX2, MITF, GBX2, and HESX1 in order to coordinate cell determination and differentiation. In humans, 21 heterozygous de novo OTX2 mutations and one microdeletion [41] have been reported [42–45] in patients with structural eye malformations (microphthalmia, anophthalmia, underdeveloped left optic nerve, ocular coloboma) and congenital hypopituitarism. Patients present with variable hypopituitarism [46–48] with GH- the most vulnerable pituitary hormone- deficiency and variable brain abnormalities (MRI findings range from normal or hypoplastic anterior pituitary, normal or ectopic posterior pituitary and Chiari malformation). As OTX2 plays a key role in retinal development some of the patients with OTX2 mutations and pituitary deficiencies present also with a variety of structural eye malformations (microphthalmia, anophthalmia, underdeveloped left optic nerve, ocular coloboma). All but 3 mutations induced a loss of function in the GNRH1, HESX1, POU1F1, and IRBP (intertial retinoid-binding protein) gene promoters, while the others are responsible for dominant negative effect [42, 49]. No genotype-phenotype correlations exist in the reported patients.

PAX6 is a well-known key regulator gene of eye development, and its heterozygous mutations in humans cause congenital eye anomalies. PAX6 is an early dorsal marker of early anterior pituitary gland, and its expression determines the boundaries of somatotrope, lactotrope, and thyrotrope cell types [50]. Recently, 3 papers have been published, reporting that PAX6 mutations may be associated with impaired pituitary function. Hergott et al. reported ed corticosterone deficiency in 5 PAX6 mutation carriers [51]. Shimo et al. reported one case of PAX6 mutation with subtle hypogonadotropic hypogonadism and borderline GH deficiency [52] and Takagi et al. [53] identified PAX6 anomalies in 2 patients with GH deficiency. They both had GH deficiency and pituitary hypoplasia while the first one had also bilateral cryptorchidism, reminiscent of hypogonadism.

Bone morphogenetic protein 4 (BMP4) plays a critical role in pituitary gland development. It is required for RP formation and maintenance and is expressed in the optic vehicle, in the diencephalic floor, consistent with its role in pituitary development and in the medial ganglionic eminence and in developing limbs [54]. Screening of 215 individuals with ocular malformations identified a chromosomal deletion of 14q22–q23 encompassing BMP4 in a familial case of anophthalmia, retinal dystrophy, brain malformation, and poly/syndactyly [54]. Deletions in BMP4 presented with bilateral anophthalmia/microphthalmia, associated with hypothyroidism,
cryptorchidism, partial agenesis of corpus callosum, sensorineural deafness, developmental delay, and cerebellar and pituitary abnormalities. Although OTX2 has been identified within this region, it cannot explain the reported complex phenotype. Another study of 19 patients with CPHD genetic analysis of BMP4 gene revealed missense mutation in a patient presenting with skeletal abnormalities (spina bifida with myelomeningocele) [55].

FGF receptor 1 (FGFR1) is a tyrosine kinase receptor that acts as a cell-surface receptor for FGF and is essential for regulation of embryonic development. It is the main receptor mediating FGF8 signaling. Genetic defects in FGFR1 have been detected in patients with CPHD [56], absent corpus callosum, septo-optic dysplasia and midline defects [57]. Recently, a Japanese female patient with CPHD and FGFR1 haplo insufficiency was described. She presented with a combined deficiency of GH, LH and FSH, central hypothyroidism and multiple neurological abnormalities. The phenotype of this patient suggests that FGFR1 abnormalities can be associated with various developmental defects of the anterior midline in the forebrain [58].

Aryl hydrocarbon receptor nuclear translocator (ARNT2) is a member of the basic HLH-PAS (Per-ARNT-Sim homology) subfamily of transcription factors. Its precise roles during pituitary and brain development are poorly known. In the mouse, Arnt2 is expressed in hypothalamus, neural retina kidney and urinary tract in a way similar to that observed in humans. In humans its involvement in pituitary hormone secretion was identified by exome sequencing in a large consanguineous Saudi Arabian family. The 6 children presented shortly after birth with central diabetes insipidus and corticotroph deficiency; 4 had TSH deficiency, 2 had GH deficiency and one had gonadotroph deficiency with undescended testes. Brain MRI showed pituitary and extra-pituitary anomalies: hypoplastic anterior pituitary, thin pituitary stalk, hypoplastic frontal and temporal lobes, thin corpus callosum, and a global delay in brain myelination; a lack of pupil response to light was reported with post-retinal eye anomalies. All children were dysmorphic with a microcephaly, prominent forehead, deep-set eyes, a well-grooved philtrum, and retrognathia. Hydronephrosis, vesico-ureteral reflux and neurogenic bladder were also present. All children carried a homozygous c.1373_1374dupTC ARNT2 variant [59].

Holoprosencephaly

Holoprosencephaly (HPE) is a cephalic disorder in which the forebrain fails to develop in 2 hemispheres. Its etiology is multifactorial, caused by environmental or genetic factors. Specific gene mutations have been recognized in only 5–10% of cases. SHH is expressed in the ventral diencephalon and in the adjacent oral ectoderm but not in Rathke’s pouch; however, it is required for midline formation, forebrain development, brain lobe determination, eye formation and BMP2 expression induction. Disruption of SHH secretion and signaling during central nervous system development are one of the main causes of HPE. The clinical phenotype is highly variable ranging from closely spaced eyes or a single central maxillary incisor, to the extreme of a single cyclopic eye and superiorly placed proboscis. SHH mutations or variants are a common cause of congenital hypopituitarism in patients with usually complex midline cerebral defects [60]. SHH binds to its receptor, leading to the activation/repression of target genes via the GLI family of zinc finger transcription, such as GLI1, GLI2 and GLI3 with GLI1, GLI2 being most important.

GLI2 is expressed in the ventral diencephalon, where it induces BMP4 and FGF8 expression, and is also expressed in the oral ectoderm, where it induces pituitary progenitors. GLI2 mutations are associated with HPE or HPE-like features with pituitary abnormalities and polydactyly [61–63]. Patients with GLI2 sequence variants have been reported to have craniofacial anomalies ranging from severe midline defects to absence of clinical findings in mutational carriers [64, 65]. In some patients, these findings associated with hypopituitarism [66], although polydactyly is a frequently observed concomitant abnormality [61]. After an extensive review of more than 200 patients, Bear et al. [63] concluded that HPE resulting from pathogenic GLI2 mutations is rare and these individuals present with pituitary anomalies, polydactyly and facial defects (mid-face hypoplasia, cleft lip/palate and hypotelorism). Among 284 patients with hypopituitarism from 3 cohorts in whom the complete coding region of GLI2 was sequenced, 15 patients (5%) had either loss-of-function or non-synonymous GLI2 variants found in updated databases [60, 66]. Three GLI2 mutations were recently described in patients with hypopituitarism, ectopic pituitary lobe and an absence of HPE [65].

FGF8 is important in setting up and maintaining the midbrain/hindbrain. It is expressed in the ventral diencephalon. Mutations in this gene were only recently described in patients with phenotypes associated with congenital hypopituitarism. [67]. A female patient with homozygous mutation presented with semilobar HPE, diabeties insipidus, TSH and ACTH insufficiency and another with heterozygous mutation with an absent corpus callosum, hypoplastic optic nerves, and Moebius syndrome.

Pituitary stalk interruption syndrome (PSIS)

PSIS is a congenital defect of the pituitary gland characterized by the triad of a thin pituitary stalk, an ectopic posterior pituitary gland and hypoplasia or aplasia of the anterior pituitary gland identified by MRI. Patients with PSIS may present with either IGHD or CPHD. Over the last decade, mutations and/or single nucleotide variants in HESX1, LHX4, OTX2, SOX3, and PROKR2 have been associated with PSIS [68–70]. LHX4 mutations have been reported to cause CPHD with variable neuroradiological abnormalities: pituitary hypoplasia, ectopic posterior pituitary and small sella turcica, corpus callosum hypoplasia or Chiari syndrome.

OTX2 gene defects were also reported in 3 PSIS patients with no ocular abnormalities [41, 45]. SOX3 may be associated with X-linked hypopituitarism, PSIS, with or without mental retardation [39]. Mutations in prokineticin-2 (PROK2) or its receptor are also implicated in PSIS. PROK2 is a G protein-coupled receptor involved in sexual and olfactory bulb development in mice, and necessary for proper neuronal migration and angiogenesis.

Human mutations of PROK2 have been reported in isolated hypogonadotropic hypogonadism. Three heterozygous PROK2 variants have been reported in 72 patients with PSIS [69] and 9 others in patients with variable phenotype from hypogonadotropic hypo-
gonadism associated with SOD to panhypopituitarism; some of them had corpus callosum hypoplasia and optic nerve anomalies, and one of these patients presented with central diabetes insipidus [57, 71, 72].

Recently, 2 female siblings with short stature due to GH deficiency, anterior pituitary hypoplasia and ectopic posterior pituitary and empty sella were reported carrying a homozygous missense mutation L19Q in GPR161 [73]. Both sisters had also anatomical anomalies with a short 5th finger, congenital alopecia, and ptosis of the left eye. GPR161 is an orphan member of the G protein–coupled receptors’ family and was first described in a mutant mouse presenting with congenital cataract and neural tube defects. GPR161 is widely expressed in the early stages of embryo development including the neural folds, the pituitary and the hypothalamus in both mouse and human. GPR161 is a key negative regulator of Sonic hedgehog (Shh) pathway the pituitary target of which is GLI2. This would suggest that gain-of-function mutations of GPR161 could lead to abnormal pituitary development by repressing the Shh pathway.

Furthermore, Dakou-Voutetakis et al identified 2 novel mutations in the HPE-related genes (TGIF gene and SHH gene) in 2 patients with PSIS and a single central incisor [68].

Central hypothyroidism and macro-orchidism

IGSF1 gene located in Xq26 is expressed in Rathke’s pouch, in adult pituitary gland and the testis, whereas IGSF1 protein is detected in GH, prolactin and TSH-secreting cells. Loss of function mutations or deletions of the IGSF1 gene causes an X-linked syndrome, characterized in males by congenital central hypothyroidism, delayed testosterone rise in puberty despite normal timing of testicular enlargement, adult macro-orchidism, and in some cases (65 %) deficiencies of PRL and/or GH [74, 75]. A delayed increase in dehydroepiandrosterone sulfate and delayed pubarche was recently described in a male with a novel hemizygous mutation [76]. A small proportion of heterozygous females show central hypothyroidism, PRL deficiency, and/or delayed menarche.

Deficient anterior pituitary with variable immune deficiency (DAVID) syndrome

Deficit in anterior pituitary gland function and common variable immune deficiency are 2 very different clinical presentations. Mutations in several genes have been associated with these 2 conditions, but until recently no gene has been associated with deficiencies in both systems in the same patient. In 2012, such patients were identified in France, and defined a novel disorder, called DAVID syndrome [77]. Chen et al. [78] reported mutations in the NFKB2 gene in 2 families from Utah with features consistent with DAVID syndrome. Since then de novo mutations near the C-terminus of NFKB2, have been reported in patients with similar phenotypes [79, 80]. However, the precise mechanism how this factor may lead to endocrine deficits remains largely unclear.

Axenfeld–Rieger syndrome

Ptx2, a member of homeobox genes, is expressed in the stomodeum at E8, Rathke’s pouch at E10.5 and pituitary anterior and intermediate lobes at E12.5. In humans, Ptx2 mutations are associated with Axenfeld–Rieger syndrome, which is characterized by anomalies in the ocular anterior compartment together with craniofacial dysmorphism, dental, cardiac, and umbilical anomalies [81].

CHARGE syndrome and pituitary deficiencies

CHARGE is an autosomal-dominant syndrome with a combination of eye coloboma, heart malformations, choanal atresia, retardation of growth and development, and genital and ear abnormalities. Gregory et al. first reported 2 novel CHD7 variants in patients with CHARGE syndrome and ectopic posterior pituitary: the first patient presented with central hypothyroidism, GH deficiency, and anterior pituitary hypoplasia associated with ectopic posterior pituitary and the second with GH, TSH, ACTH, LH/FSH deficiencies and the same MRI abnormalities [82].

Non-syndromic Hypopituitarism

Many cases of IGHD or CPHD are found without any concomitant abnormalities and are therefore not a part of a specific syndrome. They are most commonly attributed to single gene mutations as listed in Table 1.

Prop1

Prop1 is a pituitary specific paired-like homeodomain transcription factor. It is located on chromosome 5q35, consists of 3 highly conserved exons and 2 introns and encodes a protein of 226 amino acids. It is first expressed within Rathke’s pouch at E10, peaks at E12 and then rapidly decreases until E15.5 when it disappears. Prop1 can act as both a transcriptional repressor (for Hesx1 expression) and a transcriptional activator (for POU1F1).

Recessive mutations within Prop1 are the commonest genetic cause of CPHD both in sporadic (6.7 %) and familiar cases (48.5 %) [83–89]. Among 30 PROP1 variants causing CPHD reported so far, 2 are the most common – the deletion of 2 nucleotides (NM_0006261.4: c.[301_302delAG]; [301_302delAG] [90], and the deletion of one nucleotide (NM_0006261.4: c.[150delA]) which represent more than 90 % of the mutated alleles in the Eastern European cohorts. Both lead to the production of an abnormally short protein that cannot function properly. Recessive PROP1 mutations are responsible for GH, TSH, PRL and gonadotrophin deficiencies, although the time of onset and the severity varies. Most patients present with early-onset GH deficiency and growth retardation; however, normal growth in early childhood has been reported in a patient who attained a normal final height without GH replacement therapy [91]. TSH deficiency is also highly variable and has been reported as the first presenting symptom in some cases [92], while others show delayed TSH deficiency [93].

Patients with PROP1 mutations present with normal ACTH/cortisol levels in early life but often demonstrate a cortisol deficiency later in their life, significantly correlated with increasing age [94, 95].
Although PROP1 is essential for the differentiation of gonadotrophs in fetal life, the spectrum of gonadotrophin deficiency ranges from hypogonadism presenting at birth with microopenis and undescended testes to complete lack of pubertal development or even spontaneous pubertal development with infertility [93, 95, 96]. The pituitary morphology is also highly variable; most patients have normal pituitary stalk and posterior lobe, with a small or normal anterior pituitary gland on MRI. However, in some cases, an enlarged anterior pituitary gland has been reported [90] with subsequent regression over time [97].

**Pit1/POUIF1**

Human POUIF1 (previously known as PIT-1) is a member of the POU family of transcription factors responsible for pituitary development. It has been localized to chromosome 3p11 and encodes a 291-amino acid protein with a molecular mass of 33kD. The protein has 3 functional domains: a N-terminal trans-activating domain involved in protein–protein interactions, a POU-specific domain, and a POU-homeodomain involved in DNA binding and in interactions with transcriptional cofactors. It is expressed relatively late during pituitary development (e13.5) and its expression persists throughout postnatal life and adulthood. POUIF1 is important for regulation of the genes encoding GH, PRL and TSHβ. POUIF1 requires PROP11 expression and is able to interact with other transcription factors such as CREB-binding protein which interacts with GH promoter [98], PITX1 which activates the PRL and GH promoters [99], LHX3 which activate the POUIF1, TSHβ and PRL promoters [11]. More than 30 POUIF1 mutations are reported to cause CPHD, which are associated usually with complete TSH, GH and PRL deficiencies. Gonadotroph and corticotroph axes remain usually functional. Brain MRI can be normal or show small anterior pituitary development. The commonest mutation is the dominant R271W mutation [100], which has been identified in several unrelated patients from different ethnic groups. No extra-pituitary anomaly has been reported up to date.

**Conclusions**

Congenital hypopituitarism may present either as isolated pituitary hormone or combined pituitary hormone deficiencies and can be a part of a syndrome comprising anomalies of the central nervous system or other organs. The same phenotype can be attributed to different gene mutations, while a given gene mutation can provoke different phenotypes. Moreover, a given gene mutation may give rise to different MRI findings of the pituitary region ranging from normal anterior pituitary to pituitary hypoplasia or even aplasia, while posterior pituitary may be either ectopic or eutopic. Therefore, a careful delineation of the clinical phenotype and the associated morphological findings is the crucial step in the investigation of the genetic background of cases of congenital hypopituitarism. So far, still many cases have not yet received their molecular identity and still many gene mutations or even gene–epigenetic interactions have to be unraveled in the future to explain as yet unclear cases of congenital hypopituitarism.

**Conflict of Interest**

The authors declare that they have no conflict of interest.

**References**


Xatzipsalti M et al. Congenital Hypopituitarism... Horm Metab Res 2019; 51: 81–90


[29] Fang Q, Benedetti AF, Ma Q et al. HESX1 mutations in patients with congenital hypopituitarism: Variable phenotypes with the same genotype. Clin Endocrinol (Oxf) 2016; 85: 408–414


[51] Xatzipsalti M et al. Congenital Hypopituitarism... Horm Metab Res 2019; 51: 81–90


[85] Abrao MG, Leite MV, Carvalho LR et al. Combined pituitary hormone deficiency (CPHD) due to a complete PROP1 deletion. Clin Endocrinol (Oxf) 2006; 65: 294–300


[91] Arroyo A, Pernasetti F, Vasilyev VV. A unique case of combined pituitary hormone deficiency caused by a PROP1 gene mutation (R120C) associated with normal height and absent puberty. Clin Endocrinol (Oxf) 2002; 57: 283–291


