

Mechanistic Pathways of ATP Sensitive Potassium Channels Referring to Cardio-Protective Effects and Cellular Functions

Authors

Vishal Kumar Vishwakarma¹, Prabhat Kumar Upadhyay², Hridaya Shanker Chaurasiya¹, Ritesh Kumar Srivasatav³, Tarique Mahmood Ansari⁴, Vivek Srivastava¹

Affiliations

- 1 Department of Pharmacology, R.R.S College of Pharmacy, Amethi, Uttar Pradesh, India
- 2 Institute of Pharmaceutical Research, GLA University, Mathura, Uttar Pradesh, India
- 3 Faculty of Pharmacy, Kamla Nehru Institute of Management and Technology, Sultanpur, India
- 4 Department of Pharmacy, Integral University, Lucknow, Uttar Pradesh, India

Key words

potassium channel, ischemic preconditioning, cardio-protection, skeletal muscle

received 04.10.2018

accepted 23.11.2018

Bibliography

DOI <https://doi.org/10.1055/a-0806-7207>

Published online: 4.1.2019

Drug Res 2019; 69: 365–373

© Georg Thieme Verlag KG Stuttgart · New York

ISSN 2194-9379

Correspondence

Vishal Kumar Vishwakarma
Department of Pharmacology
R.R.S College of Pharmacy
Amethi
227405 Uttar Pradesh
India
Tel.: +91/988/9654 167, Fax: +91/5662/241 687
vishal049uip@gmail.com

ABSTRACT

A study of potassium channels correlates the fundamentals of mechanistic pathways and various physiological functions. The knowledge of these pathways provides the background, how to determine unit cell functions and to affect cardio protection. ATP sensitive potassium channels adjust excitability of membrane and functions as per metabolic status of cell. A lot of energy consumption primarily occurred in skeletal muscles which also express high number of potassium channels. The increase in calcium release and high heat production is occurred due to loss of potassium channels. Such type of mediations determines metabolic changes and energy required in the dissipation. IPC reduces infarct size in anesthetized mice. In ischemic-reperfusion, pressure in left ventricle was watched while contractile power recovery did not happen. It was seen that elements of intact potassium channel are fundamental for Ischemic preconditioning (IPC). If more prominent is enactment of potassium channels and their cardiologic effects create high heart rate. All the more as of late, it has been suggested that mitochondrial ATP sensitive potassium channels are critical as closing stage effectors which trigger IPC as opposed to sarcolemmal potassium channels. Nevertheless, the importance of the potassium channels reconsidered in cardio-protection in present findings. These discoveries recommend that potassium channels in the adjusting ischemic-reperfusion damage in mice. The heart rate of the mouse occurred during ischemia; enhance watchful extrapolation applied to larger warm blooded animals.

Introduction

ATP-sensitive potassium (K_{ATP}) channels have the bend nature to alter membrane properties and control the cell metabolic status [1]. K_{ATP} channels are generally spread in various tissues including cerebrum, pancreas, heart and skeletal muscle.

Different types of these channels add to particular properties of cells and tissues. Skeletal muscle K_{ATP} channels are on a very basic level bound through physical relationship in 4 potassium channel

[2]. A potassium channel like Kir 6.1, a weak significant modifier, with 4 working sulfonylurea receptor subunits, SUR2A, and incredibly less articulation of SUR1 and subunits of Kir6.1 [3]. The channel mediated metabolic status occurs via path of the K^+ pore ATP sensitivity by subunit of SUR, which play a role in activation of channel by Mg-ADP; potassium channel openers and affected by sulfonylurea drugs. Skeletal muscles and heart are highly significant sites for physical activity where K_{ATP} are found in high frequency [4].

In the heart, direct opening of K_{ATP} channel happens in light of various burdens related with either decreased vitality accessibility, for example, hypoxia or ischemia [5], or expanded vitality utilization including expanding heart rate inside the typical, physiological range. The resultant cell potassium efflux advances activity potential shortening, hence restricting the drive for calcium deluge and calcium instigated calcium discharge. This spares cell vitality that would some way or another be used for calcium homeostasis and choking [6].

An abbreviated activity potential range (AP) furthermore achieves a more broadened diastolic interval, essential for vitality hold renewal. Along these lines, a fundamental bit of ventricular K_{ATP} channels is alteration of the AP to streamline cardiovascular limit over a far reaching extent of workloads while avoid fatigue of cell metabolic assets that provoke damage or brokenness [7].

Involvement of K_{ATP} Channel in Ischemic Preconditioning

IPC is a process by which compact irregular conditions of ischemia impenetrably protect the myocardium against different insults of ischemia, leading to decline of infarct size. At first it was recommended that initiation of sarc K_{ATP} channels expect a basic part in IPC. In fact, a number of examinations have shown that IPC was replicated by potassium channel openers [8] and lessening through sulfonyleurea K_{ATP} channel blocker glibenclamide [9].

Grover et al. (1995) [10] proposed that it was not connected between APD shortening & cromakalim associated cardioprotection in dogs. The class III threatening to arrhythmic medications Dofetilide and Terikalant which was not targets, each ineffectively to segregate the cardio-protective purpose behind IPC [11]. A later, elective proposition guesses that mitochondria port another kind of K_{ATP} channel, and union of mito- K_{ATP} channels comparing with sarc K_{ATP} channels, show cardio-protection comprising IPC. Likewise, it has been revealed that diazoxide and 5-HD are by and large particular activators and blockers of mito K_{ATP} occupy in cardiomyocytes, exclusively, at littlest under non ischemic situation [12]. It was at first hypothesized by Noma et al., that incitation of sarc K_{ATP} channels act as an endogenous guard of heart structure. Movement potential shortening because of sarc K_{ATP} channel establishment is required to decrease perfect open door for Ca^{2+} joining by methods involving L-type channels of Ca^{2+} and to extend perfect open door for Ca^{2+} removal.

Without any doubt, untreated KO hearts and WT hearts with HMR1098-treatment extended for a period for the discontinuance of of pressure in the midst of ischemia contrasting and untreated WT hearts, those are by and large credited to nonappearance of dampened cardiovascular contractions as a result of movement potential shortening. In this condition, it is noteworthy that quality movement of Kir-6.2 and SUR - 2 A to COS-7 cells can endure cost of the cytoprotection against Ca^{2+} over-load promoted by hypoxia/reperfusion [13].

The outcomes of movement were not delivered in the refined cells, hyperpolarization of cell layer coming to completion in the starting of sarc- K_{ATP} channel may accept a basic part in the cytoprotection. It was proposed by various researchers that mito K_{ATP} channels may depolarize the mitochondrial film, as needs be hindering

mitochondrial Ca^{2+} [14]. If narrowing of mitochondrial Ca^{2+} over-load is one of fundamental instruments of mito- K_{ATP} channel- interceded cardio affirmation, intracellular Ca^{2+} in the ischemia in KO heart cells was found too much for mitochondria damage.

Activation of sarc- K_{ATP} channel has moreover been gave off an impression of being fundamental for cell security in various tissues and furthermore the cerebrum [15] and skeletal muscle [16]. Different examinations have exhibited that inception of mito- K_{ATP} channel is for appropriate on time and put off preconditioning in animal [17] and furthermore human myocardium [18]. Right when non beating myocardial plan are used for preconditioning tests, mito- K_{ATP} channels may accept an all the more overpowering part in the cardioprotection. The limitation of development by sulfonyleureas and so forth, for example, Dofetilide and Terikalant should not be done under crazy ischemic conditions.

Certainly both sarc K_{ATP} and mito K_{ATP} channels are responsible for the cardio-protection, forgetting about way that several differentiations in their qualities among animal species and exploratory conditions. It has been questionable that sarc- K_{ATP} or mito K_{ATP} match some regular highlights in IPC [19]. The present examination has demonstrated that sarc K_{ATP} channel work is crucial for IPC, which is a high-heart-rate creature assortment in any way in mouse. It was proposed that digoxin, limits sarc K_{ATP} channel and conveys intracellular Ca^{2+} over-load by Na^+K^+ ATPase block, repealed IPC in vivo hearts of rabbit [20]. In all terms, it has been represented to that both of mito- K_{ATP} and sarc K_{ATP} channels in canine heart, expect an essential limit of infarct estimated after IPC mechanism [21] and in erratic pain relieving [22]. The ischemic daamage is higher at standard in KO hearts than in WT hearts may destabilize IPC conventions in the 2 different ways. The Kir6.2-lacking mice push the criticalness of sarc K_{ATP} that involves in cardio-security.

Mitochondrial ATP-sensitive K^+ Channel and Rodent Cardiomyocytes

The presence of Kir6.1 protein in coronary supply routes of rodents and cardiomyocytes was predictable with the far reaching assignment of Kir6.1 mRNA of rat heart [23] and in the smooth muscle of veins [24]. The subtypes of Kir6.1 in mitochondria which were considered to results in detached part of mitochondria and myocytic ventricles observed by an examination and localized with MitoFluor red, a mitochondrial pointer [25]. The repression of Kir6.2 in ER as well as cell layer in the cardiomyocytes of rat was related to results got in electron microscopy having neurons and glial cells of cerebrum of rat [26]. The Kir6.2 proteins on ER address the site before the course of action in reasonable octamers associated with sulphonyleurea receptors on cell layer [27]. A quality trade procedure did not represent in which mitochondria possess Kir6.1 or Kir6.2 proteins [28]. Additionally, an immunoblot examination without Kir6.1 or Kir6.2 proteins in mitochondria [29].

At that point, it has been kept up both Kir6.1 and Kir6.2 channels were not the parts of mitochondria. The proteins, Kir6.1 and Kir6.2 were available in mitochondria seen by immunoblot examination of mitochondrial parts following electron microscopy [30] and in separated ventricular myocytes in rats utilizing immunocytochemistry. Kuniyasu et al. (2003) [29] utilized approaches to change their antibodies and show specificities by immunoblot examination

are relatively same as Singh et al. (2003) [25] utilized, who had isolated the chain of amino acids restricted with Kir 6.1 and Kir 6.2 proteins from mitochondria.

The Role of ATP-sensitive Potassium Channels in Cellular Defense and CVS protection

Two self-decision explore offices can put forth a defense possessing first portrayed the ATP-fragile potassium channels [31]. Some report viewed the nearness of an outside current of K^+ ions in heart muscle cells when treated with hypoxia or metabolic dangerous substances [32]. It was turned around by ATP imbued into the cell. For all intents and purposes indistinguishable recognitions were made by another social event [33]. Such coordinates were next delineated in pancreatic beta cells, skeletal muscle [34], smooth muscle [35] and neurons. It has clarified that the channel showed electrophysiological properties and pharmacological actions [36]. In back to front fixes in ~ 140 mM balanced K^+ obsessions, conductance of single-channel is represented ohm^{-1} among a conductance of 70–80 pS. The lower regards now and again observed in composition has lower and upside down K^+ obsessions. The subdued channel development are produced less by other adenine nucleotides without magnesium because they are ground-breaking. On the other hand, Mg^{2+} , ATP and ADP are considered to produce stimulatory action. Undoubtedly, it was established that channels were known to have a rich pharmacology [37].

Surprisingly, sulphonylureas were discovered when it was well-known that antimicrobial sulphonamides were also have ability to cause hypoglycaemia and observed in animal models. It was clearly ended up that insulin release induced through beta cells of pancreas due to constraint of KATP channels. As observed in case first age administrator (e. g., tolbutamide, chlorpropamide) and second-age administrators (e. g., glibenclamide, gliclazide, glipizide) which are used in the treatment of diabetes mellitus which opens K_{ATP} channels. Furthermore, these administrators particular for K_{ATP} channels show extent of substance structures like diazoxide is a benzothiadiazine, pinacidil a cyanoguanidine and nicorandil a pyridyl nitrate [38].

Characteristic Role of the Mitochondrial ATP-sensitive K^+ Channel in Cardiac Functioning

Mitochondria are recognized in both bioenergetics and protection in heart against ischemia-reperfusion injury. A broad hypothesis is that the functioning of cardioprotection uses the physiological components of mito K_{ATP} channel. This work shows that the opening of mito K_{ATP} channels has 2 special outcomes for heart which depend upon major bioenergetic process when opening of channel occurs. At the point, when DW is high, as in case of opening of mito K_{ATP} channels in resting conditions, extended mitochondrial ROS creation and ROS constitutes kinases inside a positive banner expansion circle provoking quality interpretation and cell improvement.

This causes the opening of mito K_{ATP} channels lead to cardioprotection against ischemia– reperfusion damage. When, the reduction of DW occurred in ischemia which cause opening of mito K_{ATP} channel leads to homeostasis. Other K^+ flow through mito K_{ATP}

channel lowers fundamental force which is basic for keeping a significant transport of electrons and a low conductance of film VDAC. These results assure improvement the midst of work state as well as reperfusion after ischemia [39].

Mechanism of Mito K_{ATP} Channel Opening through Signaling Pathways of IPC and CPC

Cardioprotective effects through IPC and CPC is distressed by mito K_{ATP} inhibitors for example, 5-HD and Glibenclamide. This shows not only that mito K_{ATP} is stressed in these techniques for protection, except for those mito K_{ATP} channel is opened by endogenous mechanisms [40].

Principally, the cell hyperpolarization occurred in vascular smooth muscle (VSM) is responsible for the opening of K_{ATP} channels [41]. A quick phosphorylation of channel subunits KIR6.1/SUR2B (like T633, S1387 and S1465 on SUR2B; S385 on KIR6.1) was observed directly by dynamic sub-nuclear examinations [42]. Additional control may occur through dephosphorylation of these developments for the Ca^{2+} subordinate phosphatase calcineurin [43]. Vasoconstrictors like angiotensin II and endothelin-1, activation of PKC which adjust K_{ATP} channel activity through such pathways [44]. The Ca^{2+} programmed through PKC ϵ mediation organizes phosphorylation of KIR6.1 channel which is responsible for gathering of serine developments in distal C-end as main part. There may be outcomes for channel camouflage and reusing possibly by methods for caveolae [45]. The vasoconstrictors interact with PKA results extra inhibitory assurance to K_{ATP} channels [46].

One technique that has been little researched is the piece of PIP2 utilization despite PKC initiation. In some cases, it was found that the control is abolished by PKC inhibitors. KIR6.1 channel seems to have a courteously high activity for PIP2 and channel action may be kept with huge utilization. It was proposed that PKC-assisted VSM K_{ATP} activity is subject to produce small movement of vasoconstrictors. The vascular K_{ATP} channel and cloned KIR6.1/SUR2B equivalents are focused on hormonal control through direct phosphorylation of subunits. The PKC change has been included as central role in cell protection as well as preconditioning in cardiac cells [47]. In early examinations, PKC was thought to start sarcolemmal cardiovascular K_{ATP} and there is a biphasic lead with order took after by an inhibitory action contrasting with channel camouflage [48].

In any case, with unrivaled pipette Ca^{2+} in whole cell annals in flawless cells and we saw biphasic control. Inhibitory effect was a direct result of channel mask and happened due to phosphorylation of S372 in KIR6.2 protein [49]. The course of sarcolemmal K_{ATP} channels through PKA has been inspected. PKA with KIR6.2/SUR1, phosphorylation prompts increased direct development through stores in SUR1 and KIR6.2 that are homologous to those in SUR2B and KIR6.1. One charming component is the subcellular control of K_{ATP} channels which radiate an impression of being gathered at the fore-end of T-tubule, putting forward commencement could affect excitation pressure coupling [50].

Mitochondria are a quantitatively significant wellspring of ROS, which supply to tissue harm amid ischemia, but on the other hand are middle people of IPC flagging [51]. Gathering affirmation recommends that redox flagging pathways assume an essential part in IPC [60, 61], and ready to advance m K_{ATP} actuation [52]. The es-

sential ROS created by means of mitochondria is superoxide ($O_2 \bullet^-$) [53], even as hydrogen peroxide (H_2O_2) or lipid peroxides can be shaped optionally. Both $O_2 \bullet^-$ and H_2O_2 are thought to activate mK_{ATP} [54], while clashing reports exist with respect to O_2 . The impact of extra peroxides on mK_{ATP} isn't known. In addition, it is evident that a few yet not a wide range of cancer prevention agents can lessen IPC and mK_{ATP} action, justifying more examination mK_{ATP} channel. Nitric oxide ($NO \bullet$) is additionally worried in IPC and inspires a major range of cardioprotective effects [55]. $NO \bullet$ have been recognized in the separated mitochondrial courses of action [56] and can alternatively provide various responsive nitrogen species [57] which can provide either harmful or supportive fading parts [58].

The mK_{ATP} may be a likely center for such RNS, and whereas while high doses (10 mM) of a S-nitroso-thiol (SNT) have been displayed to activate directly in the faultless mitochondria and affirmation for more physiologically pertinent effects of $NO \bullet$ has observed for the foremost portion depended on the circuitous measures of channel movement [59] or examination of the channel ousted from its mitochondrial condition [60].

Biophysical Characteristics of K_{ATP} Channels in Managing Vascular Diseases

Potassium channels vascular infections changed vascular potassium channel work beneath neurotic conditions may well be either a reason or an affect of contamination. Vasoconstriction and course of action that of a vein limit to broaden are after effects of damaged K_1 channels restrain in veins and may be direct result of an al-

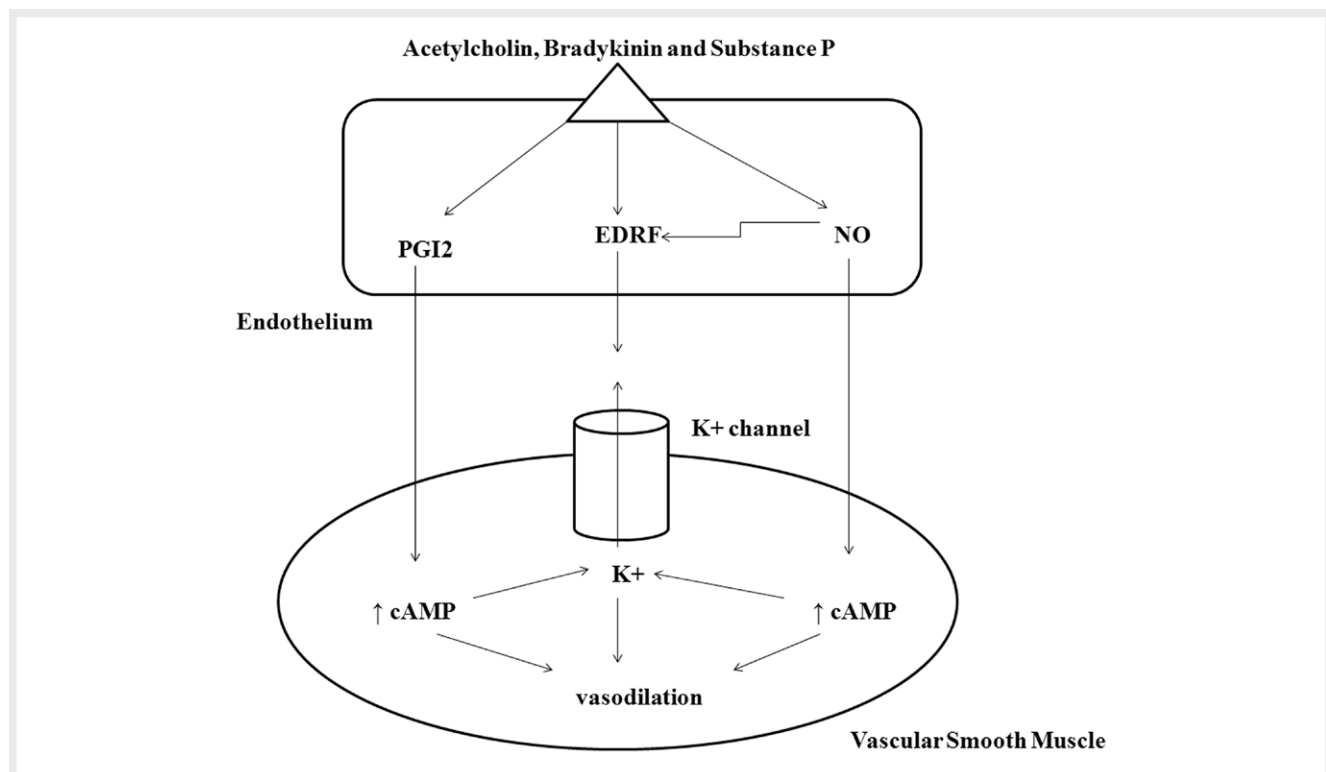
teration in number, basic conductance, and open validity of the channel(s).

All together at this level with respect to K_1 channel articulation can starting at now be given just by ponders atomic and also settle prop approaches and thusly, are limited to examinations of veins or cardio-myocytes are abolished from the physiological condition [61]. In this manner, where probability permits (i. e., the accessibility of specific pharmacological instruments), it will stay basic to check such revelations with data about vascular K_1 coordinate limit procured in the joined physiological condition in vivo (► Fig. 1).

Particularly few circulated examinations have been broken down impact of infection on biophysical features of K_1 channels or subunits. As shown above, the greater piece of our understanding into affect of illnesses on vascular K_1 channel articulation remains underhanded and subject to clarification of the test information attained by utilizing K_1 channel modulators that are accepted to be modestly particular pharmacological specialists [62].

Contribution of K_{ATP} Channels in Cerebro-Vascular Hypertension

The persistent hypertension is the most frequent thought about cardiovascular infectivity state similarly as its effects on vascular K_1 channel exertion. At present, there is unpredictable working of each one of the 4 vital K_1 channels makes in midst of Effects out of Hypertension on electrical membrane (E_m). The resting E_m of vascular smooth muscle cells is represented to be more depolarized in veins from hypertensive versus normotensive animals. Extended



► Fig. 1 Some potential instruments including K_1 channel-mediated, endothelium-subordinate hyperpolarization and involvement of various mediators.

vascular depolarization of vessels is related with overhauled myogenic tone in halls from hypertensive animals [63].

Few examinations propose that the restrain of vascular K_{ATP} channels is prevented within the middle of hypertension. Existing K_{ATP} channel activators are less sensible dilators *in vivo* in both enormous [64] and small cerebral vessels of persistently hypertensive rats [65]. This alteration shows up to most likely consolidate a weakened layer hyperpolarization response to these administrators, as essentially indistinguishable disclosures from settle catch contemplates demonstrate that a glibenclamide-unstable K_1 current started by levromakalim is reduced in mesenteric entry in smooth muscle cells of tenaciously hypertensive animals [66]. Cromakalim provoked extricating up of isolated mesenteric course is unclear prevented in case of hypertension initiated by unending NO synthase prevention [67].

The vascular K_{ATP} channels are accepted to be inactive under most normal basal conditions and amplified vascular tone within the middle of perpetual hypertension is improbable to be related to disabled K_{ATP} channel effort. For improved K_{ATP} channel effort, in continual hypertension, an extended vasodilator effect of cromakalim and significant choking of channel by glibenclamide in carotid ways from stroke-slanted SHR against normotensive WKY rats has been considered [68].

K_{ATP} Channels in Diabetes and Resulting to Hypertension in Vascular Bed

Most of information right now open for vascular K_1 occupies exertion in diabetes involving K_{ATP} channels. While for never-ending hypertension, different reports of debilitated vascular relaxant properties produced to open K_{ATP} channels which deal diabetes.

These examinations have for the mostly exploited the streptozotocin induced rat model of diabetes and have explored vessels at 2.5–4 months coming approximately to treatment with streptozotocin. In this method, the plasma glucose level are increased by 3 to 4-times; debilitated extricating up of the segregated aorta [69] and mesenteric vascular bed [70] and decreased dilatation of far reaching [71] and little [72] cerebral passages *in vivo*. These developments are accepted to be the possible result leads decrease in number of K_{ATP} channels and moreover decreased affectability of said channels design openers. Redirection of streptozotocin induced cytotoxic effects of show up a fantastical purpose behind these movements in light of the way that, as various indications of vascular brokenness, flighty vasodilator actions through K_{ATP} channel opening are neutralized by prescriptions that keep away from hyperglycemia.

Streptozotocin aggravated diabetes can essentially alter utilitarian response of K_{ATP} facilitates in different tissues, with pancreatic beta-cells [73] and ventricular myocytes, [74] showing that hyperglycemia started debilitation of K_{ATP} channels is not compelled to vasculature. Since, diabetes is characterized with raised levels of cholesterol, LDL and triglycerides in plasma. It is conceivable that some vascular difficulties of diabetes are not directly linked to hyperglycemia which is a resultant of balanced plasma lipid profile [75].

Period of exploratory hyperglycemia has each one of the saves of being indispensable parameters observed impacts of diabetes on

vascular K_{ATP} channel work in light of the way that that by disparity, reactions to K_{ATP} channel institution are represented to be dominating in the early diabetic state (► Fig. 2). For illustration, cromakalim actuated dilatation of colossal coronary supply courses in pooch are extended multi week behind treatment with alloxan, and responses of the small coronary paths are unaffected [76]. Additionally, glibenclamide produce stamped narrowing of vessels. The extended articulation and basal order of K_{ATP} channels occur together in the renal stream in front of schedule in midst of diabetes [77].

This condition can include to the upgrades in glomerular filtration rate and renal plasma stream, which develop in in beginning times of diabetes in both clinical and test settings [78]. An expanded K_{ATP} channel activity right directly may along these lines reflect a elevated metabolic state (i. e., low ATP levels) of smooth muscle cells not long after the starting of hyperglycemia. Extended K_{ATP} arrange activity in veins through metabolic weight, for occasion, within the middle of ischemia, may important for keeping up tissue perfusion.

In this way, tissues may well be additional powerless to the ischemic heart after expanded circumstances of diabetes inferable from debilitated limit of K_{ATP} channels. On the other hand, differentiated impacts on K_{ATP} channel work are represented to occur 2 months after after affirmation of hyperglycemia, most likely to a limited degree reflecting a ceaseless drop of vascular systems the middle of improvement of disease.

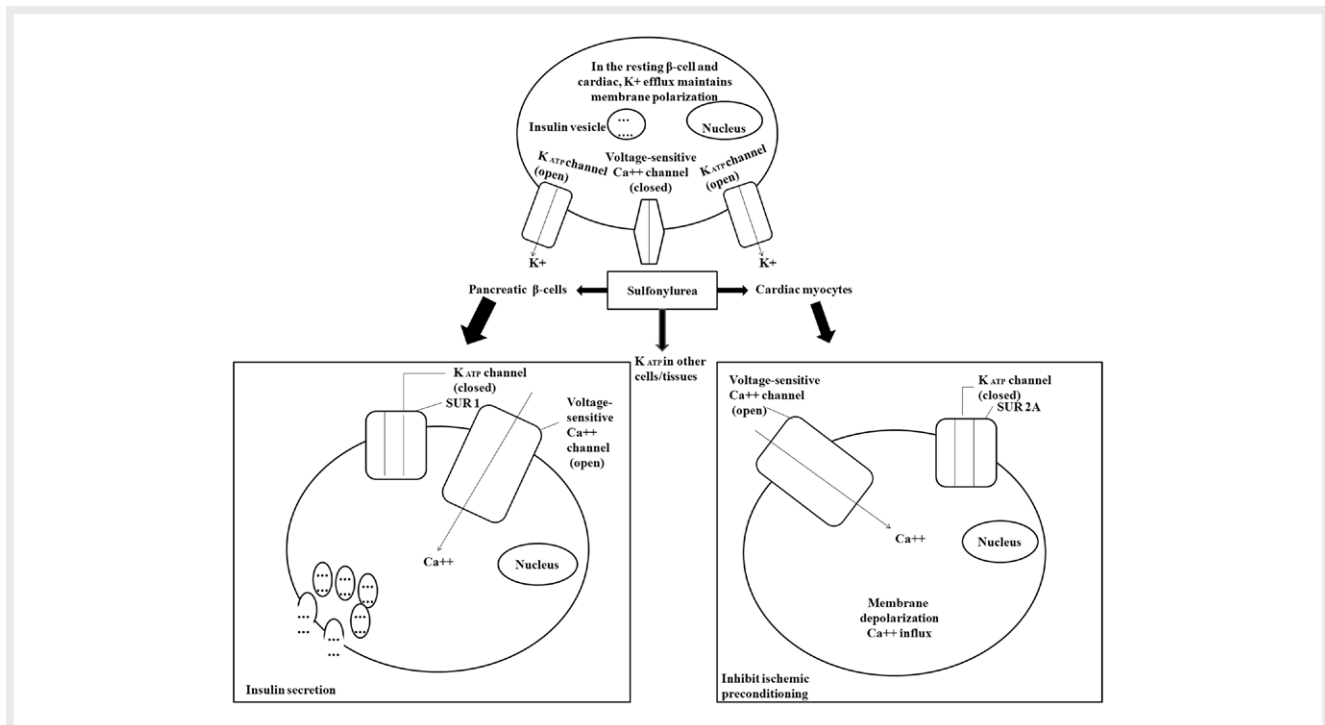
Since, Bouchard et al. reported that vasorelaxant response to lemakalim is weakend in coronary vessels, not in the vascular bed following 2 months of hyperglycemia. Pinacidil-started hyperpolarization of the mesenteric vein was represented to be protected consequent to 8 to 12 weeks. Zimmerman et al. [79] revealed that dilator activities leads to K_{ATP} opening were obstructed in cerebral courses from 4 to 8-week-diabetic rats coming about from lessened basal complimentary of endothelium-induced NO. The K_{ATP} channels restrain was re-established in diabetic vessels associated with consistent NO support [80].

Of course, such an instrument of endothelial NO-subordinate K_{ATP} channel activation couldn't occur in different stages, as no certification of an area was found for NO dilator activities in rodent cerebral vessels through K_{ATP} channel opening [81].

Hypercholesterolemia, Atherosclerosis, Cardiomyocyte Swelling and Heart Failure

It is particularly seen that vascular blocking occurred in atherosclerosis and hypercholesterolemia. Such contamination is associated with handicap of endothelial limit. The diminished vascular enlargement of endothelium-affected NO is presumably going to participate a vital part in change of atherosclerosis [82]. As a result, supply courses may exhibit an extended vascular tone and may react deficiently to endothelium-subordinate vasodilators. Under such condition of changed vascular reactivity, K_1 channel development or limit may similarly be unordinary [83].

The hypertrophy of heart is enacted by an expanded in addition to cardiovascular workload. While, transverse aortic fixing was apply in mice with heart specific over expression of SUR1 which limitlessly disturbs cardiovascular sarcolemmal K_{ATP} channel utility, extended hypertrophy of left ventricle was viewed [84]. Inquisitively, there is



► **Fig. 2** Schematic depiction of the affect of SUs on pancreatic β-cells and cardiovascular myocytes. Sulfonylureas tie to sulfonylurea receptor proteins (SURs), subunits of the K_{ATP} channels.

in every way a correspondence between cardiovascular K_{ATP} channel enunciation and the action of the PPAR-γ co-activator, PGC-1α. Reduced channel reason prompts less activity at PGC-1α promoter to some degree through FOXO-1 concealment. Restored ventricular cardiomyocytes are subjected to coronary hindrance in rats show up control of KIR6.1, in the demand of the infarct zone [85]. Congestive heart dissatisfaction or restricted corruption in human hearts prompts an upgraded AP term and affectability to potassium channel openers in the 2 atria and ventricles [86].

Angiotensin II and TNF-α enunciation is insistently related to that of KIR6.1 in failing rat myocardium or created cardiomyocytes and antagonistically related with KIR6.2 [87]. Moreover, cardiomyocytes were treated with diazoxide, exhibiting extended enunciation of KIR6.1/SUR2B in these cells as a noteworthy part of hypertrophy development. Over the best changes in cell volume in the heart item would have capacity to alter the essential cell credibility, cell limits and cell destruction. These movements can raise because of an intracellular metabolite sign that extension cell osmolarity, empower water to enter the cell, augment the cell volume and alter molecule channel work. K_{ATP} channel has been seemed to occur in cellular changes with atrial K_{ATP} coordinates opening in view of cell swelling provoking AP shortening [88]. The non-attendance of KIR6.2 in cardiovascular myocytes led to cell swelling while in WT mice cell swelling exaggerated which is bothered by extension of diazoxide [89].

Cardioprotective Effect of Ischemic Preconditioning in Ovariectomized Rat Heart

The affirmation of caveolin., a membrane protein and and a unsafe controller of nitric oxide (NO), saises after menopause Examination was anticipated to choose the impact of daidzein (DDZ), a phytoestrogen is included to appear cardioprotective effect of ischemic preconditioning (IPC) in ovariectomized rat heart.

IPC helped cardioprotection was broadly fixed in ovariectomized rats when stood out from run of the process rats, which recovered, was reestablished by the treatment of DDZ, a caveolin inhibitor (0.2 mg/kg subcutaneously) for for numerous weeks. In any case, this observed cardioprotection was amazingly diminished by perfusion of L-nitroarginine methyl ester, an endothelial nitric oxide synthase (eNOS) inhibitor (100 mM/L) and glibenclamide, an adenosine triphosphate-fragile potassium channel blocker (10 mM/L) alone or in mix, noted that expansion in myocardial infarct assess, landing of LDH and CK-MB, and and other than diminish within the section of NO.

Opening of mitochondrial K_{ATP} channels shields the myocardium from I/R-affected damage [90]. Different mediators like adenosine, bradykinin, angiotensin, prostaglandins and NO, which are increased in IPC, pass on cardioprotection by the opening of mitochondrial K_{ATP} channel [91, 92]. Further, perfusion of glibenclamide, a K_{ATP} involve blocker in DDZ-pretreated ovariectomized rat heart in a general sense, dropped the cardioprotective effect of IPC without influencing the IPC-intervened entry of NO.

It is discretionary that the watched cardioprotective impact of IPC in rodent and DDZ-pretreated rodent may be a coordinate result of opening of mitochondrial K_{ATP} channel. Also, perfusion of L-NAME with Glibenclamide in DDZ-pretreated ovariectomized rodent heart was not capable to create any extra effect in examination with specific drugs. These revelations reflect that NO conveyed in light of IPC in DDZ-pretreated ovariectomized rodent heart make cardioprotection by the opening of K_{ATP} channels [93].

Conclusion

In summary, we have demonstrated the role of the potassium channel in mediating delayed ischemic preconditioning. It is confirmed that blockers were acting at potassium channel. Further studies will be necessary to unravel the specific signal transduction mechanism by which delayed preconditioning leads to the opening of these channels and differentiating the specific role played by each of the sarcolemmal and mitochondrial potassium channels in mediating delayed cardio protective effect in vivo.

Clinical Implications

It has been indicated from various disorders that mitochondrial ATP sensitive potassium channels are critical which trigger IPC as opposed to sarcolemmal potassium channels. The increment in potassium channel mediations decides metabolic changes and energy required within the dissipation.

These channels can be determined with a role in pharmacological and biophysical properties. Nevertheless, the importance of the potassium channels reconsidered in cardio-protection in present findings. The usefulness of such studies indicates a fundamental role of potassium channels in a variety of CVS disorders including arrhythmias, hypertension and heart failures.

These findings suggest that potassium channels within the adjusting ischemic-reperfusion injury in mice. The heart rate of the mouse occurred during ischemia and probably enhances vigilant extrapolation applied to larger warm blooded animals.

Limitations of Study

A study of mechanistic pathways mediating potassium channels provides the knowledge of cell functions and cardio protection and limits to cardiovascular functions and pathways. ATP sensitive potassium channels adjust excitability of membrane and control metabolic functions. Limitations of this study include missing of experimental protocols with justification and involvement of potassium channels. A lot of advancements in this field those are not proved till date which are not included. A lack of wide versatility of potassium channels in the study for physiological functions.

Acknowledgements

We are grateful to Prof Pradeep Mishra, Director, Institute of Pharmaceutical Research, GLA University, Mathura (U.P.) and Prof Anup Matri, Principal, Rajarshi Rananjay Sinh college of Pharmacy, Amethi, (U.P.) for providing e-resources and library facilities for this study.

Conflict of Interest

Authors did not have any conflict of interest in writing and publishing the manuscript.

References

- [1] De Wet H, Proks P. Molecular action of sulphonylureas on K_{ATP} channels: A real partnership between drugs and nucleotides. *Biochem Soc Trans* 2015; 43: 901–907
- [2] MacIntosh BR, Holash RJ, Renaud JM. Skeletal muscle fatigue-regulation of excitation-contraction coupling to avoid metabolic catastrophe. *J Cell Sci* 2012; 125: 2105–2114
- [3] Flagg TP, Enkvetchakul D, Koster JC et al. Muscle K_{ATP} channels: Recent insights to energy sensing and myoprotection. *Physiol Rev* 2010; 90: 799–829
- [4] Nichols CG. Adenosine triphosphate-sensitive potassium currents in heart disease and cardioprotection. *Card Electrophysiol Clin* 2016; 8: 323–335
- [5] Gok S, Vural K, Sekuri C et al. Effects of the blockade of cardiac sarcolemmal ATP-sensitive potassium channels on arrhythmias and coronary flow in ischemia-reperfusion model in isolated rat hearts. *Vascul Pharmacol*. 2006; 44: 197–205
- [6] Alekseev AE, Reyes S, Yamada S et al. Sarcolemmal ATP-sensitive $K(+)$ channels control energy expenditure determining body weight. *Cell Metab*. 2010; 11: 58–69
- [7] Garrott K, Kuzmiak-Glancy S, Wengrowski A et al. K_{ATP} channel inhibition blunts electromechanical decline during hypoxia in left ventricular working rabbit hearts. *J Physiol*. 2017; 595: 3799–3813
- [8] Murry CE, Jennings RB, Reimer KA. Preconditioning with ischemia: A delay of lethal cell injury in ischemic myocardium. *Circulation*. 1986; 74: 1124–1136
- [9] Blondeau N, Plamondon H, Richelme C et al. $K(ATP)$ channel openers, adenosine agonists and epileptic preconditioning are stress signals inducing hippocampal neuroprotection. *Neuroscience*. 2000; 100: 465–474
- [10] Grover GJ, D'Alonzo AJ, Parham CS et al. Cardioprotection with the K_{ATP} opener cromakalim is not correlated with ischemic myocardial action potential duration. *J Cardiovasc Pharmacol* 1995; 26: 145–152
- [11] Schultz JE, Kwok WM, Hsu AK et al. Terikalant, an inward-rectifier potassium channel blocker, does not abolish the cardioprotection induced by ischemic preconditioning in the rat. *J Mol Cell Cardiol* 1998; 30: 1817–1825
- [12] Sato T, Sasaki N, Seharaseyon J et al. Selective pharmacological agents implicate mitochondrial but not sarcolemmal K_{ATP} channels in ischemic cardioprotection. *Circulation*. 2000; 101: 2418–2423
- [13] Jovanovic S, Jovanovic A. Delivery of genes encoding cardiac $K(ATP)$ channel subunits in conjunction with pinacidil prevents membrane depolarization in cells exposed to chemical hypoxia-reoxygenation. *Biochem Biophys Res Commun* 2001; 282: 1098–1102
- [14] Er F, Michels G, Gassanov N et al. Testosterone induces cytoprotection by activating ATP-sensitive $K+$ channels in the cardiac mitochondrial inner membrane. *Circulation*. 2004; 110: 3100–3107
- [15] Ohtsuka T, Ishiwa D, Kamiya Y et al. Effects of barbiturates on ATP-sensitive K channels in rat substantia nigra. *Neuroscience*. 2006; 137: 573–581
- [16] Gong B, Miki T, Seino S et al. A K_{ATP} channel deficiency affects resting tension, not contractile force, during fatigue in skeletal muscle. *Am J Physiol* 2000; 279: C1351–C1358
- [17] Chen X, Minatoguchi S, Wang N et al. Quinaprilat reduces myocardial infarct size involving nitric oxide production and mitochondrial K_{ATP} channel in rabbits. *J Cardiovasc Pharmacol* 2003; 41: 938–945

- [18] Ghosh S, Standen NB, Galiñanes M. Evidence for mitochondrial KATP channels as effectors of human myocardial preconditioning. *Cardiovasc. Res* 2000; 45: 934–940
- [19] Gross GJ, Fryer RM. Sarcolemmal versus mitochondrial ATP-sensitive K⁺ channels and myocardial preconditioning. *Circ Res.* 1999; 84: 973–979
- [20] Das B, Sarkar C. Similarities between ischemic preconditioning and 17β-estradiol mediated cardiomyocyte KATP channel activation leading to cardioprotective and antiarrhythmic effects during ischemia/reperfusion in the intact rabbit heart. *J Cardiovasc Pharmacol* 2006; 47: 277–286
- [21] Das B, Sarkar C. Is the sarcolemmal or mitochondrial K(ATP) channel activation important in the antiarrhythmic and cardioprotective effects during acute ischemia/reperfusion in the intact anesthetized rabbit model? *Life Sci.* 2005; 77: 1226–1248
- [22] Toller WG, Gross ER, Kersten JR et al. Sarcolemmal and mitochondrial adenosine triphosphate-dependent potassium channels: Mechanism of desflurane-induced cardioprotection. *Anesthesiology.* 2000; 92: 1731–1739
- [23] Bever L, Poitry S, Faure C et al. Pore loop-mutated rat Kir6.1 and Kir6.2 suppress KATP current in rat cardiomyocytes. *Am J Physiol Heart Circ Physiol* 2004; 287: H850–H859
- [24] Miura H, Wachtel RE, Loberiza FR Jr. et al. Diabetes mellitus impairs vasodilation to hypoxia in human coronary arterioles: Reduced activity of ATP-sensitive potassium channels. *Circ Res.* 2003; 92: 151–158
- [25] Singh H, Hudman D, Lawrence CL et al. Distribution of Kir6.0 and SUR2 ATP-sensitive potassium channel subunits in isolated ventricular myocytes. *J Mol Cell Cardiol* 2003; 35: 445–459
- [26] Zhou M, Tanaka O, Suzuki M et al. Localization of pore-forming subunit of the ATP-sensitive K⁽⁺⁾-channel, Kir6.2, in rat brain neurons and glial cells. *Brain Res Mol Brain Res* 2002; 101: 23–32
- [27] Teramoto N. Pharmacological profile of U-37883A, a channel blocker of smooth muscle-type ATP-sensitive K channels. *Cardiovasc Drug Rev* 2006; 24: 25–32
- [28] Seharaseyon J, Ohler A, Sasaki N et al. Molecular composition of mitochondrial ATP-sensitive potassium channels probed by viral Kir gene transfer. *J Mol Cell Cardiol* 2000; 32: 1923–1930
- [29] Kuniyasu A, Kaneko K, Kawahara K et al. Molecular assembly and subcellular distribution of ATP-sensitive potassium channel proteins in rat hearts. *FEBS Lett* 2003; 552: 259–263
- [30] Lacza Z, Snipes JA, Miller AW et al. Heart mitochondria contain functional ATP-dependent K channels. *J Mol Cell Cardiol* 2003; 35: 1339–1347
- [31] Alexander SP, Benson HE, Faccenda E et al. The concise guide to pharmacology 2013/14: Overview. *Br J Pharmacol* 2013; 170: 1449–1867
- [32] Oketani N, Kakei M, Ichinari K et al. Regulation of K(ATP) channels by P(2Y) purinoceptors coupled to PIP(2) metabolism in guinea pig ventricular cells. *Am J Physiol Heart Circ Physiol* 2002; 282: H757–H765
- [33] Trube G, Hescheler J. Inward-rectifying channels in isolated patches of the heart cell membrane: ATP-dependence and comparison with cell-attached patches. *Pflügers Arch* 1984; 401: 178–184
- [34] Spruce AE, Standen NB, Stanfield PR. Voltage-dependent ATP-sensitive potassium channels of skeletal muscle membrane. *Nature.* 1985; 316: 736–738
- [35] Standen NB, Quayle JM, Davies NW et al. Hyperpolarizing vasodilators activate ATP-sensitive K⁺ channels in arterial smooth muscle. *Science.* 1989; 245: 177–180
- [36] Noma A, Takano M. The ATP-sensitive K⁺ channel. *Jpn J Physiol* 1991; 41: 177–187
- [37] Wakeno-Takahashi M, Otani H, Nakao S et al. Adenosine and a nitric oxide donor enhances cardioprotection by preconditioning with isoflurane through mitochondrial adenosine triphosphate-sensitive K⁺ channel-dependent and -independent mechanisms. *Anesthesiology.* 2004; 100: 515–524
- [38] Mannhold R. Structure-activity relationships of K(ATP) channel openers. *Curr Top Med Chem* 2006; 6: 1031–1047
- [39] Saks V, Kaambre T, Guzun R et al. The creatine kinase phosphotransfer network: Thermodynamic and kinetic considerations, the impact of the mitochondrial outer membrane and modelling approaches. *Subcell Biochem* 2007; 46: 27–65
- [40] Garlid KD, Pauczek P. Mitochondrial potassium transport: The K⁺ cycle. *Biochim Biophys Acta.* 2003; 1606: 23–41
- [41] Rodrigo GC, Standen NB. ATP-sensitive potassium channels. *Curr Pharm Des* 2005; 11: 1915–1940
- [42] Shi Y, Wu Z, Cui N et al. PKA phosphorylation of SUR2B subunit underscores vascular KATP channel activation by beta-adrenergic receptors. *Am J Physiol Regul Integr Comp Physiol* 2007; 293: R1205–R1214
- [43] Orié NN, Thomas AM, Perrino BA et al. Ca²⁺ /calineurin regulation of cloned vascular K ATP channels: Crosstalk with the protein kinase A pathway. *Br J Pharmacol* 2009; 157: 554–564
- [44] Sampson LJ, Davies LM, Barrett-Jolley R et al. Angiotensin II-activated protein kinase C targets caveolae to inhibit aortic ATP-sensitive potassium channels. *Cardiovasc Res.* 2007; 76: 61–70
- [45] Jiao J, Garg V, Yang B et al. Protein kinase C-epsilon induces caveolin-dependent internalization of vascular adenosine 5'-triphosphate-sensitive K⁺ channels. *Hypertension.* 2008; 52: 499–506
- [46] Hayabuchi Y, Davies NW, Standen NB. (2001b) Angiotensin II inhibits rat arterial KATP channels by inhibiting steady-state protein kinase A activity and activating protein kinase C. *J Physiol* 2001; 530: 193–205
- [47] Yellon DM, Downey JM. Preconditioning the myocardium: From cellular physiology to clinical cardiology. *Physiol Rev.* 2003; 83: 1113–1151
- [48] Light PE, Sabir AA, Allen BG et al. Protein kinase C-induced changes in the stoichiometry of ATP binding activate cardiac ATP-sensitive K⁺ channels. A possible mechanistic link to ischemic preconditioning. *Circ Res.* 1996; 79: 399–406
- [49] Aziz Q, Thomas AM, Khambra T et al. Regulation of the ATP-sensitive potassium channel subunit, Kir6.2, by a Ca²⁺ -dependent protein kinase C. *J Biol Chem* 2012; 287: 6196–6207
- [50] Korchev YE, Negulyaev YA, Edwards CR et al. Functional localization of single active ion channels on the surface of a living cell. *Nat Cell Biol* 2000; 2: 616–619
- [51] Murphy E, Steenbergen C. Preconditioning: The mitochondrial connection. *Annu Rev Physiol* 2007; 69: 51–67
- [52] Forbes RA, Steenbergen C, Murphy E. Diazoxide-induced cardioprotection requires signaling through a redox-sensitive mechanism. *Circ Res.* 2001; 88: 802–809
- [53] Murphy MP. How mitochondria produce reactive oxygen species. *Biochem J.* 2009; 417: 1–13
- [54] Fornazari M, De Paula JG, Castilho RF et al. Redox properties of the adenosine triphosphate-sensitive K⁺ channel in brain mitochondria. *J Neurosci Res* 2008; 86: 1548–1556
- [55] Burwell LS, Brookes PS. Mitochondria as a target for the cardioprotective effects of nitric oxide in ischemia–reperfusion injury. *Antioxid Redox Signal* 2007; 10: 579–600
- [56] Giulivi C, Poderoso JJ, Boveris A. Production of nitric oxide by mitochondria. *J Biol Chem* 1998; 273: 11038–11043
- [57] Rudolph V, Freeman BA. Cardiovascular consequences when nitric oxide and lipid signaling converge. *Circ Res.* 2009; 105: 511–522

- [58] Sasaki N, Sato T, Ohler A et al. Activation of mitochondrial ATP-dependent potassium channels by nitric oxide. *Circulation*. 2000; 101: 439–445
- [59] Ljubkovic M, Shi Y, Cheng Q et al. Cardiac mitochondrial ATP-sensitive potassium channel is activated by nitric oxide in vitro. *FEBS Lett* 2007; 581: 4255–4259
- [60] Nelson MT, Quayle JM. Physiological roles and properties of potassium channels in arterial smooth muscle. *Am J Physiol* 1995; 268: C799–C822
- [61] Faraci FM, Heistad DD. Regulation of the cerebral circulation: Role of endothelium and potassium channels. *Physiol Rev*. 1998; 78: 53–97
- [62] Harder DR, Smeda J, Lombard J. Enhanced myogenic depolarization in hypertensive cerebral arterial muscle. *Circ Res*. 1985; 57: 319–322
- [63] Kitazono T, Heistad DD, Faraci FM. ATP-sensitive potassium channels in the basilar artery during chronic hypertension. *Hypertension*. 1993; 22: 677–681
- [64] Sobey CG, Heistad DD, Faraci FM. Effect of subarachnoid hemorrhage on cerebral vasodilatation in response to activation of ATP-sensitive K1 channels in chronically hypertensive rats. *Stroke*. 1997; 28: 392–397
- [65] Ohya Y, Setoguchi M, Fujii K et al. Impaired action of levromakalim on ATP-sensitive K1 channels in mesenteric artery cells from spontaneously hypertensive rats. *Hypertension*. 1996; 27: 1234–1239
- [66] Kalliovalkama J, Jolma P, Tolvanen J-P et al. Arterial function in nitric oxide-deficient hypertension: Influence of long-term angiotensin II receptor antagonism. *Cardiovasc Res* 1999; 42: 773–782
- [67] Miyata N, Tsuschida K, Otomo S. Functional changes in potassium channels in carotid arteries from stroke-prone spontaneously hypertensive rats. *Eur J Pharmacol* 1990; 182: 209–210
- [68] Kamata K, Miyata N, Kasuya Y. Functional changes in potassium channels in aortas from rats with streptozotocin-induced diabetes. *Eur J Pharmacol* 1989; 166: 319–323
- [69] Bouchard J-F, Dumont EC, Lamontagne D. Modification of vasodilator response in streptozotocin-induced diabetic rat. *Can J Physiol Pharmacol* 1999; 77: 980–985
- [70] Mayhan WG. Effect of diabetes mellitus on response of the basilar artery to activation of ATP-sensitive potassium channels. *Brain Res*. 1994; 636: 35–39
- [71] Mayhan WG, Faraci FM. Responses of cerebral arterioles in diabetic rats to activation of ATP-sensitive potassium channels. *Am J Physiol* 1993; 265: H152–H157
- [72] Tsuura Y, Ishida H, Okamoto Y et al. Impaired glucose sensitivity of ATP-sensitive K1 channels in pancreatic b-cells in streptozotocin-induced NIDDM rats. *Diabetes*. 1992; 41: 861–865
- [73] Shimoni Y, Firek L, Severson D et al. Short-term diabetes alters K1 currents in rat ventricular myocytes. *Circ Res*. 1994; 74: 620–628
- [74] Makino A, Ohuchi K, Kamata K. Mechanisms underlying the attenuation of endothelium-dependent vasodilatation in the mesenteric arterial bed of the streptozotocin-induced diabetic rat. *Br J Pharmacol* 2000; 130: 549–556
- [75] Kersten JR, Brooks LA, Dellsperger KC. Impaired microvascular response to graded coronary occlusion in diabetic and hyperglycemic dogs. *Am J Physiol* 1995; 268: H1667–H1674
- [76] Ikenaga H, Bast JP, Fallet RW et al. Exaggerated impact of ATP-sensitive K1 channels on afferent arteriolar diameter in diabetes mellitus. *J Am Soc Nephrol* 2000; 11: 1199–1207
- [77] Hostetter TH, Troy JL, Brenner BM. Glomerular hemodynamics in experimental diabetes mellitus. *Kidney Int*. 1981; 19: 410–415
- [78] Fukao M, Hattori Y, Kanno M et al. Alterations in endothelium-dependent hyperpolarization and relaxation in mesenteric arteries from streptozotocin-induced diabetic rats. *Br J Pharmacol* 1997; 121: 1383–1391
- [79] Zimmerman PA, Knot HJ, Stevenson AS et al. Increased myogenic tone and diminished responsiveness to ATP-sensitive K1 channel openers in cerebral arteries from diabetic rats. *Circ Res*. 1997; 81: 996–1004
- [80] Sobey CG, Faraci FM. Effect of nitric oxide and potassium channel agonists and inhibitors on basilar artery diameter. *Am J Physiol* 1997; 272: H256–H262
- [81] Faraci FM, Orgren K, Heistad DD. Impaired relaxation of the carotid artery during activation of ATP-sensitive potassium channels in atherosclerotic monkeys. *Stroke*. 1994; 25: 178–182
- [82] Taguchi H, Faraci FM, Kitazono T et al. Relaxation of the carotid artery to hypoxia is impaired in Watanabe Heritable Hyperlipidemic rabbits. *Arterioscler Thromb Vasc Biol* 1995; 15: 1641–1645
- [83] Yamada S, Kane GC, Behfar A et al. Protection conferred by myocardial ATP-sensitive K⁺ channels in pressure overload-induced congestive heart failure revealed in KCNJ11 Kir6.2-null mutant. *J Physiol* 2006; 577: 1053–1065
- [84] Isidoro TN, Philip-Couderc P, Papageorgiou I et al. Expression and function of ATP-dependent potassium channels in late post-infarction remodeling. *J Mol Cell Cardiol* 2007; 42: 1016–1025
- [85] Fedorov VV, Glukhov AV, Ambrosi CM et al. Effects of KATP channel openers diazoxide and pinacidil in coronary-perfused atria and ventricles from failing and non-failing human hearts. *J Mol Cell Cardiol* 2009; 51: 215–225
- [86] Isidoro TN, Philip-Couderc P, Baertschi AJ et al. Angiotensin II and tumour necrosis factor alpha as mediators of ATP-dependent potassium channel remodelling in post-infarction heart failure. *Cardiovasc Res*. 2009; 83: 726–736
- [87] Saegusa N, Sato T, Saito T et al. Kir 6.2-deficient mice are susceptible to stimulated ANP secretion: K (ATP) channel acts as a negative feedback mechanism. *Cardiovasc Res*. 2005; 67: 60–68
- [88] Prasad SM, Al-Dadah AS, Byrd GD et al. Role of the sarcolemmal adenosine triphosphatase-sensitive potassium channel in hyperkalemic cardioplegia-induced myocyte swelling and reduced contractility. *Ann Thorac* 2006; 81: 148–153
- [89] Taliyan R, Singh M, Sharma PL et al. Possible involvement of α 1 adrenergic receptor and KATP channels in cardioprotective effect of remote aortic preconditioning in isolated rat heart. *J Cardiovasc Dis Res* 2010; 1: 145–151
- [90] Baines CP, Cohen MV, Downey JM. Signal transduction in ischemic preconditioning: The role of kinases and mitochondrial KATP channels. *J Cardiovasc Electr* 1999; 10: 741–754
- [91] Murphy E. Primary and secondary signalling pathways in early preconditioning that converge on the mitochondria to produce cardioprotection. *Circ Res*. 2004; 94: 7–16
- [92] Goyal A, Semwal B, Yadav HN. Abrogated cardioprotective ischemic preconditioning in ovariectomized heart. *Human & experimental toxicology* 2015; 10: 1–10
- [93] Oldenburg O, Cohen MV, Yellon DM et al. Mitochondrial K(ATP) channels: Role in cardioprotection. *Cardiovasc Res* 2002; 55: 429–437