Duration of a "Brown-Like" Phenotype of White Adipose Tissue Induced by the β3 Agonist CL-316,243

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Introduction
Adipose tissue can be divided into the brown adipose tissue (BAT) type and white adipose tissue (WAT) type, both of which have different physiological functions. WAT is loaded with fat molecules and its main function is to store energy in such lipid droplets. Excess WAT leads to obesity, diabetes and other related diseases [1,2]. BAT is strongly expressed in children, but much less so in adults, and its role is to quickly burn energy to generate heat to support the body to adapt to the cold environment via non-shivering thermogenesis. The transformation of WAT into BAT has been suggested to be a very attractive treatment option for obesity and diabetes [1]. BAT activation resulting in increased "burning" of stored fat, heat production and a resultant decrease in fat depots has been observed in man in several clinical studies after exposure to cold [3–5]. It has been reported that WAT in patients with severe burns gradually shifted to a BAT-like phenotype in both molecular and functional properties, suggesting that white fat in humans can undergo gradual “browning” to cope with such burn traumas and transform from an energy-storing to an energy-dissipating tissue [6]. Pharmacological treatments such as prednisolone or capsinoids under certain conditions activate BAT and increase energy expenditure in humans [2,7–9]. Recently, WAT to BAT transformation after pharmacological treatment i.e. phosphodiesterase inhibitor Sildenafil has been demonstrated in man [10].

In conclusion, the present results indicate that browning-like changes of white fat may be short lasting after treatment termination and could require maintenance treatment of inductor to achieve desired therapeutic effect. This might be a serious shortcoming of potential therapeutic use.

ABSTRACT
"Browning" i.e. the transformation of white adipose tissue into brown-like adipose tissue could induce efficient burning of excess fat reserves via induction of non-shivering thermogenesis. For example, activation of β3 adrenergic receptors has been show to induce such changes, however, it is still not clear, how long after termination of such a treatment, beneficial effects might be maintained. To address this question, we treated rats s.c. for 2 weeks with the β3 agonist CL-316,243 at 1 mg/kg and assessed interscapular brown fat and inguinal white fat pads weight, UCP-1 (a marker for the brown-like fat phenotype) using immunohistochemistry and H&E staining, at different intervals after treatment termination.

One day after the treatment cessation there was a decrease of inguinal white fat pad weight and increase of interscapular fat pad. This change vanished at 7 days for inguinal pad and at 14 days for interscapular pad. Histological analysis of interscapular pads showed increased UCP-1 staining and brown-like morphology in H&E staining slices at 1 day, but not other time points. In case of inguinal pad there were brown-like features in H&E slices at 1 day and less after 7 days, but absent at 14 days. UCP-1 staining was only detected 1 day after the treatment.

In conclusion, the present results indicate that browning-like changes of white fat may be short lasting after treatment termination and could require maintenance treatment of inductor to achieve desired therapeutic effect. This might be a serious shortcoming of potential therapeutic use.

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features [11–15]. Similar effects have been shown for activation of β3 receptors by selective agonists such as CL-316,243 [12, 16–18].

However, it is still not known, how long such a brown fat-like phenotype in WAT adipocytes remains after termination of the treatment. This is crucial for therapeutic use since constant, long term treatment with e. g. some β3 agonists may result in an increase in side-effects and loss of efficacy [19].

To address this issue we treated rats s.c. with CL-316,243, the activity of which we previously verified in our model [20], for 2 weeks at 1 mg/kg and assessed effects at different intervals after treatment termination. In such animals we assessed interscapular (BAT) and inguinal (WAT) fat pads weight and histology using UCP-1 and H&E staining.

Methods

Animals

Male SD rats (200–300 g, Vital River Laboratories Animal Ltd.) were housed in Plexiglas cages (3 rats per cage) at a temperature (22 ± 3 ºC) and humidity (55 ± 15 %) controlled rooms of an AAALAC accredited animal facility. Animals were provided with food (Kaooxieli certified Rodent Diet) and sterile water ad libitum and kept on a 12-h light/dark (6:00 am /6:00 pm) cycle acclimated for 1 week prior to the study.

Reagents and consumables

CL-316,243 disodium was purchased from Sequoia Research Products Ltd. (Pangbourne, UK) and dissolved in phosphate buffered saline with pH of 7.4. All other reagents were purchased from Sigma.

Experimental design

Rats were injected s.c. with sterile vehicle or CL-316,243 (1 mg/kg) daily for 14 days (4 groups, N = 8 per group) as shown in ▶ Fig. 1. On sacrifice day, interscapular and inguinal fat pads were gently isolated, weighed and processed further for histological analysis (see below).

Fat histology analysis

Following completion of the experimental protocol, rats were deeply anesthetized and their fat pads (FP) were removed and placed in 10 % neutral buffered formalin at room temperature for 24 h. The FP were then transferred to 15 % sucrose in a phosphate buffer (PB, 0.1 M, pH 7.3) solution at 4 ºC for 24 h, and finally placed in 30 % sucrose solution in PB at 4 ºC for 48 h. After de-paraffinization /rehydration of tissue sections, the FP was histologically sectioned at 5μm using a cryostat and sections mounted directly onto gelatin coated slides. Heat mediated antigen retrieval conditioning in citrate buffer (pH 6) was performed on tissue sections before commencing with a standard IHC staining protocol. Briefly, slides were immerse in 3 % hydrogen peroxide solution at room temperature for 15 min and next given a 20 min incubation in protein block, serum-free (Dako, Cat #X0909). Afterwards, a further incubation for 1 h at RT in rabbit anti-UCP1 antibody (cat #ab10983, Abcam) was applied at 1:500 dilution in antibody diluent (Dako, Cat #S2022). Slides were then exposed to three 10 min washes in PB and incubated for 30 min in HRP Labelled polymer anti-rabbit (Dako, Cat #K4003) followed by another three 10 min washes with PB. The distribution of peroxidase was revealed by incubating the sections in a DAB solution (Dako, Cat#K8002) for 5 min before being counterstained with hematoxylin. Slides were allowed to dry and cover-slipped with neutral balsam and sealed. Positive IHC area was quantified by IPP 6.0 software. The whole slide was scanned and all the positive area was selected with the same magnification. The pen tool was used to circle the positive area and view the statistics.

Statistical analysis

Results were expressed as mean ± SEM or medians in case of non-Gaussian distribution. Statistical analysis was performed using Student t-test or ANOVA on ranks followed by Man-Whitney test. The difference was considered significant when p < 0.05.

Results

Body weight

There was no significant difference of body weight between CL-316,243 and vehicle treated rats on the last day of treatment i. e. the values were: 389.4 ± 8.5, 387.5 ± 5.1, 387.5 ± 8.2 and 394.4 ± 6.1 g. for saline, and the three (▶ Fig. 1) CL-316,243 treated groups respectively (mean ± SEM).

Inguinal and interscapular fat pad weight/volume

After 2 weeks of treatment with CL-316,243 there was a significant decrease in inguinal fat pad (IFP) weight one day (group 2) after termination.
treatment termination (Fig. 1; One way ANOVA). This effect vanished on day 7 and 14. Interestingly, an increase in interscapular fat pad weight was observed on day 1 and 7 (but not 14) after treatment termination (Fig. 2).

**Fat pads histology analysis**

In animals treated with CL-316,243 there were signs of BAT activation in interscapular fat pad seen in H&E slices and also increase in UCP-1 staining 1 day, but not 7 or 14 days after treatment termination (Fig. 3).

In inguinal fat pad, treatment with CL-316,243 increased BAT like multilocular lipid droplets in inguinal fat as seen in slices stained with H&E 1 day after treatment termination, less evident at 7 days and absent after 14 days (Fig. 4).

CL-316,243 also increased staining for UCP-1 staining in inguinal fat pad suggesting transformation into BAT-like phenotype one day after treatment but not at other tested intervals. Respective median values were 1.1 (% of UCP-1 positive cells) for 1 day interval and 0 for all other groups. The difference for 1 day was statistically significant (Kruskal Wallis ANOVA on ranks \( H = 26.65, P < 0.001 \) followed by Mann-Whitney test, \( U = 0.0, p < 0.001 \)). It should be noted that in two animals in 7 days group there was increase in UCP-1 observed (Fig. 5).

**Discussion**

The present study showed that systemic treatment with CL-316,243 for 2 weeks (1 mg/kg) produced changes in WAT (inguinal fat pad) indicative transformation into a BAT-like phenotype or showing of beige like features such as characteristic adipocytes morphology and UCP-1 staining. Moreover, signs of native BAT activation were also obtained. This is in line with published literature on CL-316,243 showing after chronic treatment hypertrophy of BAT and BAT-like changes of WAT in rats on a fatty diet [21] as well as in mice [22]. Also, at the histological level, browning-like changes were observed in WAT of diabetic Zucker rats after similar treatment [23].

In addition to changes at the histological level, we also observed a decrease in inguinal fat pad weight indicating that enhanced energy expenditure may have taken place. Such a decrease in WAT pads weight has been previously described after chronic treatment with CL-316,243 in rats [21] and in mice [22].

Interestingly, the degree of cold-induced browning of WAT in various body sites of mice seems to correlate with sympathetic innervation [24] which suggests the role of \( \beta3 \) receptors as one of the major mediators. Therefore, data obtained in the present study with the \( \beta3 \) agonist may have some implication to other treatments/inductors producing browning. The formation of brown-like adipocytes in reaction to chronic cold stress in mice is reversed...
within 5 weeks of warm adaptation [25]. This is in concert with the present data suggesting that WAT to BAT-like transformation seems to be rather short lasting as it disappeared 7 or 14 days after the treatment had been stopped. Parallel to that observation, in native BAT (interscapular fat pad) activation was observed accompanied by an increase in weight one day after treatment termination but was absent 7 or 14 days later.

This may indicate some limitations of browning induction by pharmacological treatment for therapeutic effect as it may be short lasting and may require continues treatment. However, such long term treatment may have some limitations since for some pharmacological approaches tolerance to the therapeutic effects and significant increase of side-effects have been observed [19]. It should, however, be considered that this conclusion may not apply to all types of treatment (here specifically β3 agonist CL-316,243) and not to all species (including man). It does however point to such a risk potential and it adds to other pitfalls connected with concept of “browning” of WAT in man e.g. it has so far been shown in man mainly after severe “traumatic events” like extensive skin burns or cachexia [6, 26].

Conclusions
The present study shows that the in vivo conversion of the WAT to BAT-like phenotype is short lasting and requires maintenance treatment with the activator (here β3 agonist). This would be an important shortcoming of clinical application of WAT to BAT-like transformation, however species differences and specificity of type of the pharmacological target should be also taken into account.

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Fig. 5 Effect of two weeks treatment with CL-316,243 on inguinal fat pad histology visualized as UCP-1 staining 1, 7 and 14 days after treatment termination. Four representative pictures from each treatment group are shown. Red arrows point to UCP-1 stained cells.

Conflict of Interest

The Authors have no conflict of interest to declare.

References


